

CANCER

Targeting the renin-angiotensin system to improve cancer treatment: Implications for immunotherapy

Matthias Pinter^{1,2} and Rakesh K. Jain^{1*}

Renin-angiotensin system (RAS) inhibitors (RASi)—widely prescribed for the treatment of cardiovascular diseases—have considerable potential in oncology. The RAS plays a crucial role in cancer biology and affects tumor growth and dissemination directly and indirectly by remodeling the tumor microenvironment. We review clinical data on the benefit of RASi in primary and metastatic tumors and propose that, by activating immunostimulatory pathways, these inhibitors can enhance immunotherapy of cancer.

INTRODUCTION

The circulating renin-angiotensin system (RAS) is mainly known for its pivotal role in maintaining cardiovascular homeostasis and fluid and electrolyte balance. In addition, a local RAS is expressed in many tissues and mainly acts at the cellular level, where it mediates cell proliferation, growth, and metabolism. The local RAS works synergistically and independently of the systemic RAS. Angiotensin II (AngII) is the main effector and maintains tissue homeostasis by exerting regulatory and counterregulatory effects through its different receptors. Alternative peptide-receptor axes also assist in maintaining this balance (1–7). Figure 1 provides an overview of the main components of the RAS. Dysregulation of the RAS, for example, by overexpression of certain RAS components [such as renin, Ang-converting enzyme (ACE), or AngII type 1 receptor (AT1R)], can be involved in the pathophysiology and progression of a broad range of diseases, such as arterial hypertension, kidney disease, and other cardiovascular conditions (5, 8, 9).

The discoveries of captopril—the first orally active ACE inhibitor (ACEi)—in the mid-1970s (10) and losartan—the first orally active, selective AT1R blocker (ARB)—around a decade later (11) represent milestones in the history of the RAS. Numerous ACEis and ARBs have been developed since then. Now, ACEis and ARBs are the most common inhibitors of the RAS and are widely used in the management of several diseases, such as arterial hypertension, heart failure, myocardial infarction, and chronic kidney disease (12–15). Direct renin inhibitors (such as aliskiren) represent a third class of RAS-acting agents and have been added to the armamentarium more recently (16). A list of RAS inhibitors (RASi) approved by the U.S. Food and Drug Administration (FDA) is provided in table S1.

After being in clinical use for more than two decades in non-malignant diseases, ACEi/ARBs have recently received considerable attention in oncology. A large-scale meta-analysis (17), published in 2010, found an increased overall occurrence of cancer in ARB users. However, two other meta-analyses published subsequently did not confirm these data (18, 19). The FDA also rebutted these findings with their own meta-analysis (20) and an integrated analysis of all 19 rodent carcinogenicity assays of ARBs (21). Thus, the data to date do not support an association between ACEi/ARB use and an increased cancer risk. However, they do not suggest a reduced occurrence of cancer either.

¹Edwin L. Steele Laboratories for Tumor Biology, Department of Radiation Oncology, Harvard Medical School and Massachusetts General Hospital, Boston, MA 02114, USA.

²Division of Gastroenterology and Hepatology, Department of Internal Medicine III, Medical University of Vienna, Vienna, A-1090, Austria.

*Corresponding author. Email: jain@steele.mgh.harvard.edu

Copyright © 2017
The Authors, some
rights reserved;
exclusive licensee
American Association
for the Advancement
of Science. No claim to
original U.S. Government
Works. Distributed
under a Creative
Commons Attribution
License 4.0 (CC BY).

Of interest, an increasing number of preclinical studies support the involvement of RAS signaling in cancer development, growth, and progression (4). These data have led to investigations of the effects of RASi—both retrospectively and prospectively—in patients with different types of cancer. Interim analysis of a recent phase 2 trial—stemming from our preclinical findings (22)—showed encouraging R0 (microscopically margin-negative) resection rates in patients with locally advanced pancreatic ductal adenocarcinoma (PDAC) receiving neoadjuvant losartan plus chemoradiation (23). Moreover, our recent retrospective analysis indicated that RASi use is associated with improved survival of patients with nonmetastatic PDAC, presumably by stimulating the tumor's immune microenvironment, normalizing its extracellular matrix (ECM), and reducing the malignant potential of cancer cells (24).

In light of these emerging data, we discuss the role of the RAS in cancer biology with a special emphasis on tumor immunity. In addition, by carefully analyzing the studies with positive versus negative outcomes, we make a case for targeting the RAS to improve treatment of certain malignancies. Moreover, RASi may not only improve the outcome of immunotherapies but also reduce or even prevent adverse effects associated with these therapies.

The AngII/AT1R axis shapes the tumor microenvironment and promotes an immunosuppressive milieu

Components of the RAS are expressed in various human cancers and cell lines (4). Overexpression of AT1R is typically associated with more aggressive tumor features (larger tumors, higher grade, and higher vascular density) and worse outcomes (25–29).

Moreover, RAS components are also expressed in many cell types of the tumor microenvironment, such as endothelial cells, fibroblasts, monocytes, macrophages, neutrophils, dendritic cells, and T cells (4, 30–34). RAS signaling in these cells can facilitate or hinder growth and dissemination and has been shown to affect cell proliferation, migration, invasion, metastasis, apoptosis, angiogenesis, cancer-associated inflammation, immunomodulation, and tumor fibrosis/desmoplasia (1, 4). Generally, the AngII/AT1R axis is considered to favor tumor growth, whereas AngII/AT2R and Ang(1–7)/MAS signaling have opposing effects (1, 4). However, there are also conflicting reports suggesting potential tumor type-specific differences (35–39).

The tumor-promoting actions of the ACE/AngII/AT1R axis, the main target of classical RASi, have been reviewed elsewhere (1, 4). In this section, we focus on its role in tumor immunity and propose RASi as an adjunct for immunotherapy. Immune checkpoint inhibitors have recently achieved compelling success in melanoma and other solid tumors (40). However, their efficacy is diminished by a major barrier—the immunosuppressive tumor microenvironment (41). Here, we review

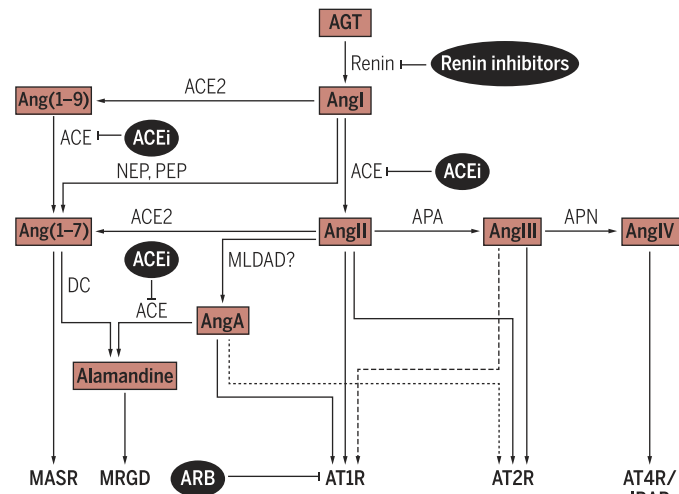


Fig. 1. The RAS is a complex system whose bioactive peptides signal through different receptors. Angiotensinogen (AGT), generated and released into circulation by the liver, is hydrolyzed by renin, a product of the kidneys' juxtaglomerular cells, to form AngI. AngI is then hydrolyzed by ACE, predominantly expressed by endothelial cells in the vascular territory of the lungs, to form the biologically active AngII. In addition to AngII, other truncated bioactive peptides have been identified, such as AngIII, AngIV, Ang(1-7), Ang(1-9), AngA, and alamandine. AngII interacts with two seven-transmembrane receptors, AT1R and AT2R, both of which also mediate the effects of AngA. Ang(1-7) mainly acts via the MAS receptor (MASR), and alamandine binds and signals through MRGD (MAS-related G protein-coupled receptor D). IRAP (insulin-regulated membrane aminopeptidase; also known as AT4R) is a binding site for AngIV (1-7). APA, aminopeptidase A; APN, aminopeptidase N; DC, decarboxylase; MLDAD?, mononuclear leukocyte-derived aspartate DC; NEP, neutral endopeptidase; PEP, prolylendopeptidase.

how AngII/AT1R signaling shapes the tumor immune microenvironment by modulating desmoplasia, vasculature, inflammation, and immune cells. We also discuss how RASi could alleviate immunosuppression and enhance the outcome of immunotherapy.

Tumor desmoplasia and solid stress

By regulating cancer-associated fibroblasts (CAFs) and profibrogenic pathways [such as transforming growth factor- β (TGF- β)], the RAS plays a key role in establishing a desmoplastic environment (22, 42), which affects the immune response in multiple ways (Fig. 2). CAFs can manipulate the immune system directly by inhibiting T and NK (natural killer) cell functions, promoting accumulation of suppressive cell types, and maintaining an inflammatory protumorigenic milieu (43). TGF- β can also directly induce immune suppression by inhibiting the T cell response (44). Dense tumor fibrosis represents a physical barrier to T cell infiltration (45). It also compresses blood vessels by increasing solid stress (46, 47). The reduced tumor perfusion results in a hypoxic and acidic milieu, which promotes reprogramming of macrophages into an immunosuppressive phenotype, impairs tumor killing functions of immune cells, and up-regulates the expression of inhibitory immune checkpoint molecules, such as programmed death-ligand 1 (PD-L1), by immune, stromal, and tumor cells (Fig. 3) (46-51). Normalizing the desmoplastic milieu (for example, by targeting profibrotic pathways and CAFs) can improve the efficacy of immunotherapy (52-54).

Several studies have demonstrated that RASi can successfully normalize the fibrotic stroma. Co-injection of cancer cells with stromal

cells increases tumor size and fibrosis, and treatment with ARBs attenuates these effects (55, 56). Losartan inhibits collagen I production by CAFs and reduces stromal collagen and hyaluronic acid (HA) in several desmoplastic tumor models by decreasing profibrotic signaling via TGF- β , connective tissue growth factor, HA synthase 1 and 3, and endothelin-1 (22). Therefore, losartan reduces solid stress and improves vascular perfusion, resulting in decreased tumor hypoxia and improved distribution and efficacy of anticancer drugs and nanotherapeutics (22, 42). Similarly, inhalation delivery of losartan and telmisartan reduces active TGF- β and collagen I expression and increases the intratumoral distribution of nanoparticles (57, 58). Moreover, the cross-talk between tumor-associated neutrophils (TANs), adipocytes, and pancreatic stellate cells (PSCs) promotes tumor desmoplasia and pancreatic cancer growth in obesity (59). AT1R inhibition attenuates obesity-induced fibrosis and tumor progression and improves response to chemotherapy (CHT). The AT1R blockade also reduces TANs and regulatory T cells (T_{regs}) and increases $CD8^+$ T cells through inhibition of PSC activation and subsequent reduction of interleukin-1 β (IL-1 β) expression (59). In another orthotopic model of pancreatic cancer, inhibition of aberrant TGF- β activity by losartan reduced collagen deposition and accumulation of T_{regs} (60).

Collectively, these data support the idea that targeting AngII/AT1R signaling with RASi can effectively reduce tumor desmoplasia and thereby decrease solid stress, increase tumor perfusion, reduce hypoxia, enhance T cell infiltration and antitumor immunity, and improve delivery and efficacy of anticancer drugs. Thus, inhibiting the AngII/AT1R axis appears to be an attractive strategy, especially for highly desmoplastic tumors, such as PDAC and some subtypes of breast and lung cancer, and RASi may represent a promising combination partner for immunotherapy.

Angiogenesis and tumor vasculature

Considerable evidence suggests that AngII/AT1R signaling promotes VEGF-mediated angiogenesis in solid tumors. AT1R expression correlates with VEGF and VEGF receptor (VEGFR) expression and microvessel density (MVD) in different human tumors (26, 27, 29). In experimental studies, AngII promoted VEGF expression in tumor (61-63) and stromal cells (64). Treatment with either ACEi or ARB reduced VEGF expression and decreased MVD and neovascularization in vivo (65, 66).

VEGF also induces vascular hyperpermeability, one of the main characteristics of the abnormal tumor vasculature (46, 48). Tumor vessel leakiness promotes tumor hypoxia and acidosis by impairing tumor blood flow (Fig. 2) (48, 67). As mentioned above, hypoxia helps to create an immunosuppressive milieu (Fig. 3) and promotes tumor progression and dissemination (48, 68). Tumor vessel normalization can alleviate hypoxia, reprogram the immunosuppressive microenvironment, and improve the efficacy of immunotherapy in mice (68, 69). Glioblastoma patients who show enhanced tumor blood perfusion under anti-angiogenic therapy have markedly prolonged survival compared to subjects who experience no change or a decrease in perfusion (70-72). RASi also reduces VEGF-mediated vascular leakiness in the dermis and retina of rodents (73, 74).

In an orthotopic model of PDAC, inhibition of aberrant TGF- β signaling by losartan restored vessel diameter and permeability (60). In a retrospective study of glioblastoma patients receiving anticancer therapy, a concomitant treatment with matrix-depleting antihypertensive drugs improved vascular function as assessed by magnetic resonance imaging (75).

The impaired perfusion and hypoxic condition of tumors can be further aggravated by AngII-induced vasoconstriction and increased

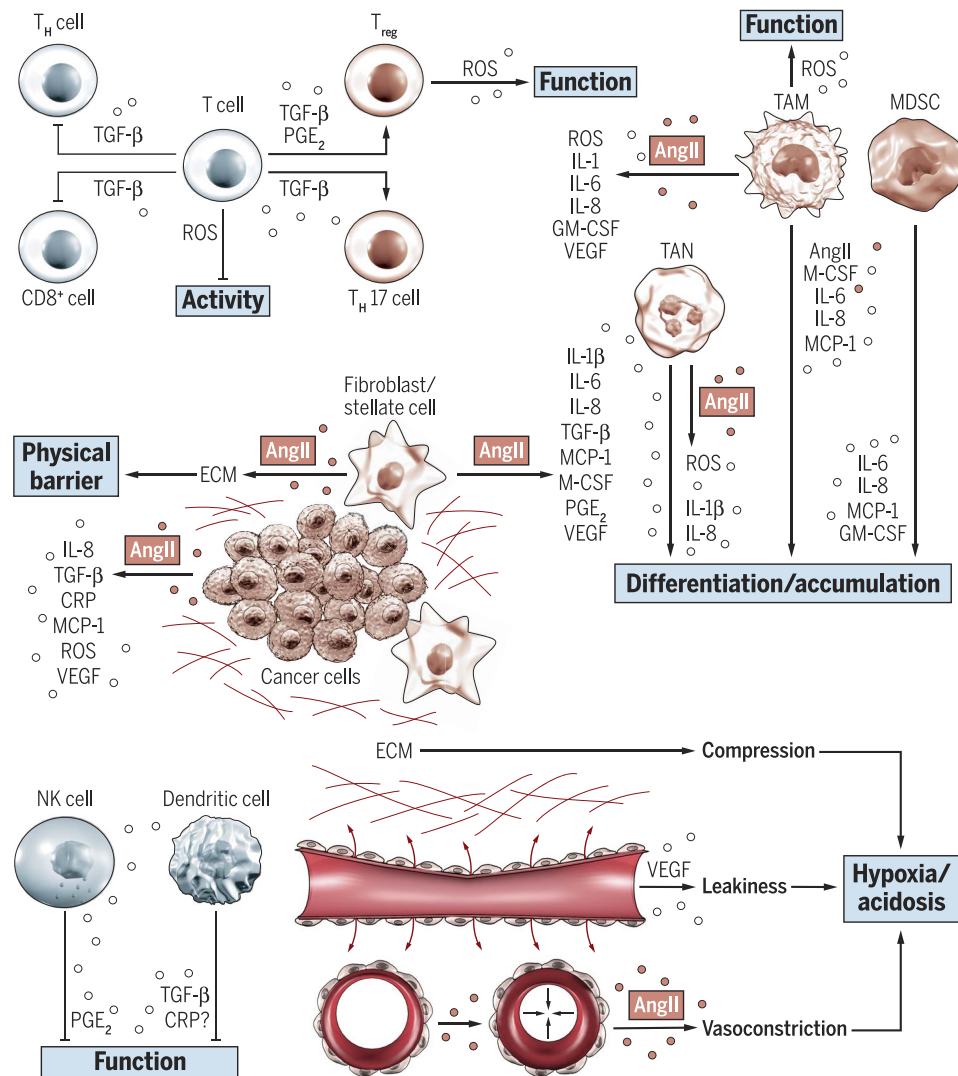


Fig. 2. The AngII/AT1R axis regulates the tumor stroma and contributes to an immunosuppressive micro-environment. AngII/AT1R signaling can increase production and release of several proinflammatory cytokines in both tumor and stromal cells. Immunomodulatory cytokines regulate a myriad of immunosuppressive immune responses by modulating differentiation, recruitment, and function of both myeloid and lymphoid immune cell types (4, 43, 44). More precisely, these cytokines suppress the differentiation and function of immunostimulatory cell types [for example, T_H (T helper) and CD8⁺ cells, NK cells, and dendritic cells] and activate recruitment and function of tumor-promoting cell types [such as T_{reg}, T_H17 cells, TANs, TAMs (tumor-associated macrophages), and MDSCs (myeloid-derived suppressor cells)]. Fibroblasts are a major source of cytokines and also play a key role in establishing a desmoplastic stroma by production and deposition of ECM. The dense tumor fibrosis represents a physical barrier to immune cell infiltration (45) and compresses blood vessels by increasing tissue stiffness and solid stress. The reduced tumor perfusion results in a hypoxic and acidic milieu, which further promotes immunosuppression (46–48). Vascular endothelial growth factor (VEGF)-induced vascular leakiness (48) and AngII-mediated vasoconstriction (76, 77, 80) further impair tumor perfusion and aggravate hypoxia. GM-CSF, granulocyte-macrophage colony-stimulating factor. PGE₂, prostaglandin E₂.

vascular resistance (Fig. 2) (76, 77). Our laboratory has shown that AngII transiently enhanced tumor blood flow and interstitial fluid pressure by increasing the mean arterial blood pressure in different tumor types (78, 79). However, Thews and colleagues (80) found that AngII infusion decreased tumor perfusion and oxygenation in small subcutaneous sarcomas but increased both parameters in large tumors. They concluded that perfusion decreased due to vasoconstriction of preexisting functionally intact host vessels in small sarcomas, whereas

the newly formed tumor vessels in large tumors did not seem to have this vaso-responsive capability, possibly due to lack of smooth muscle cells and/or angiotensin (AT) receptors (80).

Together, available data indicate that AngII/AT1R signaling impairs tumor blood supply through multiple mechanisms, such as desmoplasia-mediated vessel compression, VEGF-induced vessel leakiness and abnormal morphology, and AngII-mediated vasoconstriction of host vessels. The resulting tumor hypoxia aggravates immunosuppression and evasion. Although RASi can reduce VEGF-mediated angiogenesis and desmoplasia, additional studies are needed to ascertain whether RASi have the ability to normalize the tumor vasculature, similar to anti-VEGF agents (48).

Inflammation and immune cell modulation

The RAS promotes cancer-related inflammation and infiltration of tumor-promoting immune cells (1, 4, 81), both of which enhance the immunosuppressive micro-environment (41, 82). Here, we discuss how the RAS modulates the expression of inflammatory cytokines and orchestrates the recruitment of cancer-associated immune cells to the tumor microenvironment.

Inflammatory cytokines

A number of studies have shown that AngII/AT1R signaling can increase the production and release of several proinflammatory cytokines in both tumor and stromal cells (4). Fibroblasts represent a main target of the RAS and play a pivotal role in maintaining an inflammatory response. Cytokines released from tumor and stromal cells upon AT1R activation by AngII include TGF-β, IL-1α, IL-1β, IL-6, IL-8, MCP-1 (monocyte chemoattractant protein-1), M-CSF, COX-2 (cyclooxygenase-2), and CRP (C-reactive protein) (Fig. 2) (4, 22, 42, 56, 59, 65, 83–87). Immunomodulatory cytokines (such as TGF-β, IL-1β, MCP-1, IL-6, and IL-8) can up-regulate multiple—mostly immunosuppressive—pathways by modulating the differentia-

tion and recruitment of both myeloid and lymphoid immune cell types (Fig. 2) (44, 82, 88–91). COX-2 suppresses antitumor immunity and contributes to resistance to immunotherapy, mainly through prostaglandin E₂ synthesis (92, 93). The role of tumor-derived CRP in tumor immunity is less clear, but it may impair dendritic cell function by reducing their migration activity (94).

Oxidative stress represents another aspect of cancer-related inflammation. Although reactive oxygen species (ROS) are involved in T cell

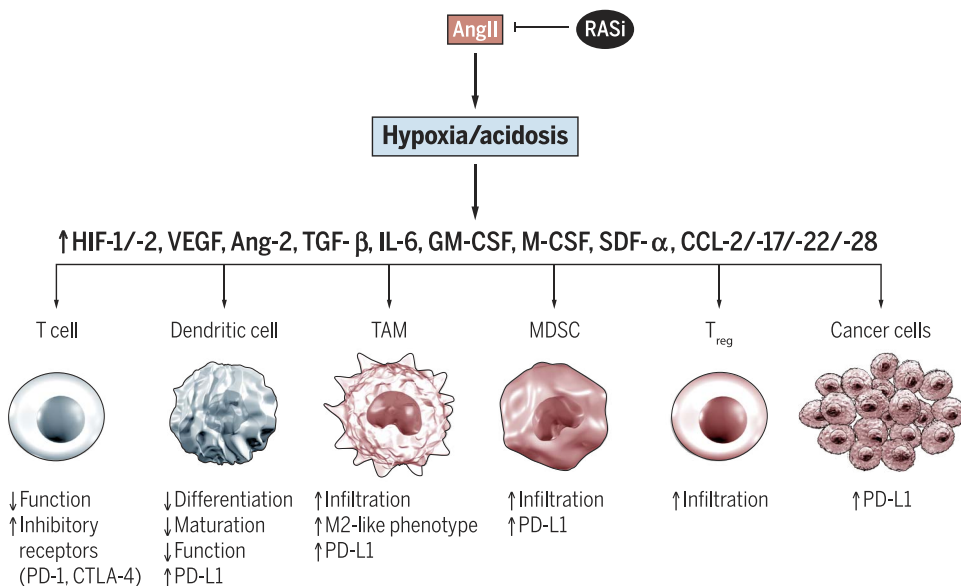


Fig. 3. Tumor hypoxia and acidosis promote immunosuppression. AngII/AT1R-mediated effects on tumor vasculature (shown in Fig. 2) can impair tumor perfusion and oxygenation, resulting in hypoxia and acidosis within the tumor stroma. The resulting up-regulation of various cytokines, growth factors, and transcription factors [including HIF (hypoxia-inducible factor), VEGF, and TGF- β] enhances an immunosuppressive microenvironment, characterized by impaired T and dendritic cell function, accumulation of immunosuppressive cell types (M2-like macrophages, MDSCs, and T_{regs}), and increased expression of inhibitory immune checkpoint molecules such as PD-L1 in tumor and immune cell types (48–50, 68). Ang-2, angiotensin-2; CCL, CC chemokine ligand; CTLA-4, cytotoxic T lymphocyte-associated protein 4; SDF, stromal cell-derived factor.

activation (95, 96), exposure to ROS can reduce T cell fitness (90, 97, 98) and enhance the function of T_{regs} (99) and TAMs (100). TAMs typically show a polarized M2-like phenotype and contribute to immunosuppression, whereas M1-like macrophages are known to induce anti-tumor immunity (101). AngII/AT1R signaling induces ROS generation in tumor cells and stromal cells (4). In prostate cancer cells, AngII-mediated expression of oxidative stress-related proteins (such as inducible nitric oxide synthase) and the generation of the ROS family member O₂⁻ radical are attenuated by the ARB candesartan (102).

Immune cells

Several studies have shown that RASi can reduce infiltration of TAMs. In human prostate cancer, high MCP-1 and macrophage infiltration are associated with more aggressive tumor features, and MCP-1 independently correlates with prostate-specific antigen recurrence (103). AngII/AT1R signaling promotes production and infiltration of TAMs in experimental tumor models; inhibition of AngII production or AT1R signaling down-regulates MCP-1, restrains tumor-induced TAM response, reduces tumor growth, and prolongs survival (34, 103–105).

AngII/AT1R signaling is also important for myeloid differentiation and functional maturation (106). ACE knockout mice show enhanced extramedullary myelopoiesis and increased numbers of cells with MDSC phenotype (32). In contrast, cultured bone marrow from ACE 10/10 mice, a mouse line overexpressing ACE in monocytic cells, demonstrates enhanced myeloid maturation and reduced MDSC production; macrophages from these mice have a more proinflammatory phenotype and more antitumor activity compared to those from wild-type mice (107). Similarly, tumor-bearing ACE 10/10 mice showed enhanced immune response, which ultimately resulted in a reduced tumor growth. Notably, ACEi reversed the beneficial effects on tumor growth, but AT1R blockade did not, suggesting that the effects of

ACE overexpression were not dependent on AngII/AT1R signaling (108, 109).

Together, available data clearly demonstrate that AngII/AT1R signaling stimulates the expression of different cytokines and growth factors from tumor and stromal cells, which enhance cancer-related inflammation and promote an immunosuppressive microenvironment (Fig. 2). Beyond the tumor immune microenvironment, the AngII/AT1R axis is also crucial for the maturation and function of immunostimulatory myeloid cells, and ACE overexpression in monocytic cells enhances antitumor immunity, although the latter effect seems to be independent of the AngII/AT1R axis. These conflicting data highlight the complexity of the RAS in cancer immunity. However, because studies supporting a stimulatory role of RAS in tumor immunosuppression considerably outweigh opposing data, we propose that RASi can effectively reprogram the tumor microenvironment toward an immunostimulatory milieu and enhance the efficacy of immunotherapy.

RASi to reduce side effects of immunotherapy

As discussed above, RASi may increase the intratumoral delivery of T cells and immunotherapeutic agents by modulating tumor vasculature and desmoplasia. This may allow for reduction in the dose of immunotherapeutic agents without decreasing the therapeutic benefit and could ultimately result in a decreased number of severe (grades 3 and 4) immunotherapy-related adverse effects. These side effects can occur in more than 50% of patients, especially if certain checkpoint blockers are combined, and some can be even life-threatening (110, 111).

Obesity and associated chronic inflammation seem to play a critical role in inducing immunotherapy-associated toxicities (112, 113). Systemic stimulatory immunotherapy, such as α CD40/IL-2, can cause a cytokine storm, characterized by high tumor necrosis factor- α (TNF- α) and IL-6, resulting in multiorgan pathologies and lethality in obese but not in lean mice (112, 113). The TNF blockade ameliorates the observed toxicities in obese mice (113). Inhibition of the RAS can also ameliorate chronic inflammation, as shown by reduced serum concentrations of proinflammatory cytokines (TNF- α and IL-6) in patients with hypertension and diabetes (114–116). This represents another way that RASi may help to reduce or even prevent immunotherapy-induced toxicity.

RAS inhibition can improve treatment of certain tumors

The effect of RASi on the clinical outcome of patients with different tumor types has been extensively studied in recent years. Tables S2 and S3 provide an overview of the published prospective (117–126) and retrospective studies (24, 127–175), respectively. Here, we summarize the main conclusions based on the available data.

RASi usage in conjunction with CHT

Available clinical data suggest that RASi may potentiate the effect of certain systemic antitumor therapies. The use of RASi was associated

with better outcomes in patients with different solid tumors who received platinum-based CHT (142, 143, 149, 165, 172). The gain in overall survival (OS; the length of time from either the date of diagnosis or the start of treatment that patients are still alive) ranged from ~3 months in advanced non-small cell lung cancer (NSCLC) to 5.7 months in advanced gastric cancer and even 11 months in metastatic colorectal cancer (CRC) (142, 149, 165, 172). In line with the clinical data, experimental studies showed that platinum-based CHT can increase VEGF production through up-regulation of AT1R expression. This seems to represent a mechanism for platinum resistance that can be successfully targeted by RASi (176, 177).

In addition, concomitant RASi treatment was associated with better survival in patients with metastatic renal cell carcinoma (RCC; gain in OS, 7 to 26 months) (137–140), metastatic CRC (gain in OS, ~11 months) (172), glioblastoma (175), and advanced hepatocellular carcinoma (HCC; gain in OS, ~5 months) (173) who received VEGF-targeted therapies. Because AngII/AT1R signaling promotes VEGF-mediated angiogenesis (4), RASi may potentiate the effect of anti-VEGF therapy. In a mouse model of Ehrlich's ascites carcinoma, the ARB olmesartan augmented the anti-angiogenic effect of the tyrosine kinase inhibitor (TKI) sorafenib (178). RASi may also represent a strategy to inhibit rapid revascularization (179, 180) and regrowth of tumors (181, 182) after cessation of anti-VEGF therapy, which is often necessary due to treatment-related side effects, especially with VEGFR TKIs (183, 184). Notably, arterial hypertension is a common side effect of anti-VEGF therapy and can be associated with better survival outcomes (185). VEGF-targeted therapy-induced hypertension is often treated with RASi, which could represent a potential confounder for the reported beneficial survival results associated with RASi use in patients who received anti-VEGF therapies. However, two points suggest otherwise: First, some studies reported the number of patients who received RASi either at baseline or after initiation of anti-VEGF therapy and showed that most of the patients were taking RASi already at baseline (137, 139). Second, McKay and colleagues (140) demonstrated that even in the subgroup of patients who developed anti-VEGF therapy-induced hypertension, RASi users had improved survival compared to nonusers.

Finally, two studies suggested a putative clinical benefit of RASi use in patients who received epidermal growth factor receptor (EGFR) TKIs (128, 143). This could be explained by the preclinical finding that AT1R signaling can regulate proliferation and migration of cancer cells through transactivation of the EGFR by metalloproteinase-dependent shedding of EGF ligands (4).

Tumor characteristics as determinants of RASi efficacy

RASi use was associated with better outcomes in multiple studies, whereas no association was found in others. This suggests that response to RASi treatment may also vary by tumor type and depend on certain tumor characteristics, as discussed below.

In breast cancer, only 2 of 13 studies shown in tables S2 and S3 reported beneficial effects of RASi use, whereas 3 studies found worse outcomes. A meta-analysis found no association of ACEi/ARB use with disease-free survival (DFS; the length of time after primary treatment for a cancer ends that the patient survives without any signs or symptoms of that cancer) or OS in breast cancer (186). The heterogeneity in terms of tumor stage, hormone receptor status, human epidermal growth factor receptor 2 overexpression, and (neo)adjuvant treatment regimen could have masked a potential benefit of RASi in certain subgroups and highlights the need for careful patient selection to obtain homogenous and comparable study cohorts.

The use of RASi was associated with better outcomes in patients with RCC, CRC, and HCC (tables S2 and S3). These tumors are well known to respond to anti-VEGF therapy (187–189). As discussed earlier, RASi may enhance the efficacy of VEGF-targeted therapies and thereby improve clinical outcome. However, in HCC (125, 126, 159, 164) and some CRC (167) and RCC (144) studies listed in tables S2 and S3, most of the patients were not treated with anti-VEGF treatment, suggesting that anti-VEGF-responsive tumors generally seem to be more sensitive to RASi.

RASi therapy had a clinical benefit in both slowly progressing cancers, such as prostate cancer, and highly aggressive tumor types, such as glioblastoma and pancreatic cancer (tables S2 and S3). A phase 2 study at the Massachusetts General Hospital (MGH) is currently investigating whether adding losartan to CHT (FOLFIRINOX), followed by chemoradiation, can convert locally advanced PDAC to resectable tumors (23). Preliminary results from this trial showed that R0 resection was achieved in 13 of 25 patients (52%), which is a major improvement compared to previously reported R0 resection rates obtained with neoadjuvant FOLFIRINOX and radiation in locally advanced PDAC (23 to 24%) (190, 191). The median OS was 33 months, with a 2-year survival rate of 65% for all patients and 83% for resected patients (23).

In addition, RASi use was effective in both early and advanced tumor stages. In some tumor types, the effect of RASi was investigated primarily in either early tumors (such as resected urinary tract cancer) (130, 147, 150, 151) or advanced stages (such as metastatic NSCLC) (142, 149). In RCC and CRC, positive outcomes were reported for both early (144, 167) and metastatic diseases (137–140, 172). Notably, in PDAC, a survival benefit in RASi users was only shown for locally advanced/metastatic diseases treated with CHT (168–170) but not for resected early/locally advanced tumors (174).

In contrast, in our own retrospective analysis, RASi use was associated with longer OS in pancreatic cancer patients with resected primary tumors (median OS, 36.3 versus 19.3 months) and locally advanced tumors (median OS, 11.3 versus 9.3 months) but not in metastatic patients. To obtain mechanistic insights, we performed RNA sequencing expression profiling of prospectively collected cancer treatment-naïve pancreatic cancer samples (four lisinopril-treated patients versus four controls). Our data suggest that lisinopril, which was the most commonly used ACEi in our cohort, normalized the ECM, down-regulated genes involved in cancer progression (such as Wnt and Notch signaling), and up-regulated genes associated with the activity of T cells and antigen-presenting cells. In addition, we identified a predictive gene signature for RASi-mediated survival, which was validated in two publicly available cohorts (24). A recently published meta-analysis pooling data on different solid tumor types (192) showed that the use of ACEi or ARB was associated with improved DFS and OS. After pooling studies that were classified as early (I/II) or advanced (III/IV) stage-dominant, the association with DFS remained significant in both stages ($P = 0.04$ and $P = 0.03$, respectively); a positive association with OS was only observed in advanced tumor stage (192).

Finally, HCC usually develops in patients with underlying liver fibrosis/cirrhosis (193). The peritumoral liver tissue and the severity of liver dysfunction determine prognosis of HCC, and complications of cirrhosis (portal hypertension and variceal bleeding) are a common cause of death in patients with HCC (193). The AngII/AT1R axis plays a crucial role in the pathophysiology of liver cirrhosis (194), and RASi can improve both liver fibrosis (195) and portal hypertension (196). These effects, in addition to the direct antitumor effects of RASi,

may also contribute to the improved outcome observed in HCC patients treated with RASi (125, 126, 159, 164, 173).

CONCLUSIONS

Preclinical studies have provided compelling evidence that the AngII/AT1R axis regulates almost all hallmarks of cancer. RASi can directly attenuate tumor growth and dissemination and improve the efficacy of systemic therapies by increasing drug delivery to the tumor tissue. The latter should help to reduce the dose of CHT and immunotherapy without decreasing the benefit and consequently decrease the anti-cancer therapy-induced side effects.

It is also clear that AngII/AT1R signaling contributes to the immunosuppressive tumor microenvironment in multiple ways. The immunosuppressive milieu is a major barrier for immunotherapy and may explain why immune checkpoint inhibitors have failed in some tumor types, such as PDAC, and have benefited only a fraction of patients in other indications where these agents are approved. Studies have shown that AT1R inhibition can decrease infiltration of immunosuppressive cell types and increase the number of effector T cells. This could also help to reduce the dose of immunotherapy without lowering drug efficacy, eventually resulting in a decreased number of severe immunotherapy-induced side effects. Although not yet studied in the context of tumor immunity, the AngII/AT1R axis is also important for the maturation of immune effector cells.

Multiple clinical studies have also revealed that RASi may have beneficial effects in a broad range of malignancies. The gain in survival is tumor type- and stage-dependent and ranged from 3 months (advanced NSCLC) to more than 25 months (metastatic RCC) in retrospective studies. However, response to RASi treatment may not only vary with tumor types but also depend on certain tumor characteristics, cancer treatment, and RASi type and dosing. More precisely, RCC, HCC, PDAC, glioblastoma, urinary tract cancer, and NSCLC seem to belong to the responsive tumor types, whereas breast cancer is rather unresponsive to RASi. With respect to cancer treatment, RASi use was associated with better outcomes in patients with NSCLC, gastric cancer, and CRC who received platinum-based CHT and in those with RCC, HCC, and CRC treated with anti-VEGF therapy (for example, sunitinib). More data are needed for other tumor types, such as melanoma, thyroid cancer, head and neck cancer, and hematologic malignancies.

Because the clinical evidence largely came from retrospective studies and small prospective pilot trials, these findings should be considered as hypothesis-generating. However, given the large amount of preclinical and clinical data suggesting a beneficial effect of RASi in different cancer types, we propose that RASi have a great potential to become an adjunct within the oncological armamentarium. Ongoing trials testing whether RASi can improve the antitumor effect of certain anticancer treatments are listed in table S4.

Future perspectives and translational challenges

Advancing the promising strategy to reprogram the tumor microenvironment with RASi to enhance anticancer treatment will require a close interplay between basic and clinical research and addressing a number of outstanding questions. Preclinical research should combine immune checkpoint inhibitors or other immunotherapy approaches with RASi to confirm whether RASi have the potential to reprogram the immunosuppressive microenvironment and eventually render tumors more sensitive to immunotherapies. In addition, mechanistic studies should not only focus on effects of RASi on the tumor stroma

but also investigate treatment-related changes within immune cell populations in the bone marrow and lymphoid organs. This will help to better understand the role of the RAS in cancer immunity.

Moreover, clinical pilot studies focusing on biological readouts—such as intratumoral ECM deposition, immune cell infiltration, and drug distribution—should be designed to confirm the available pre-clinical data and to pave the way for large randomized controlled efficacy trials. These studies should seek to identify those patients who may benefit most from concomitant RASi use. Such personalized approaches require a tight integration between measurements of various biomarkers—circulating (profibrotic molecules, immune cells, and chemokines), tissue (profibrotic molecules, collagen, and HA), and imaging (perfusion, oxygenation, and drug distribution)—and the treatment outcome (197). Assessing the intratumoral expression of the components of the RAS may also have the potential to predict response to RASi treatment.

Finally, the beneficial response of tumors to RASi is dose-dependent. For example, the collagen content of desmoplastic tumors decreases with an increasing dose of ARBs (42). However, increasing the dose can cause hypotension and other adverse effects. One potential solution to this challenge is to develop nanoformulations of RASi that will preferentially deliver RASi to the tumor microenvironment. Addressing these issues and challenges will unravel the complexity of RAS signaling and its role in different malignancies and enable development of new strategies to deliver RASi to tumors in safe doses with an even better outcome.

SUPPLEMENTARY MATERIALS

www.sciencetranslationalmedicine.org/cgi/content/full/9/410/ean5616/DC1

Table S1. RASi approved by the FDA.

Table S2. Published prospective studies using RASi in different types of cancer.

Table S3. Published retrospective studies using RASi in different types of cancer.

Table S4. Ongoing prospective studies investigating the effect of RASi in solid malignant tumors.

REFERENCES AND NOTES

- E. I. Ager, J. Neo, C. Christophi, The renin-angiotensin system and malignancy. *Carcinogenesis* **29**, 1675–1684 (2008).
- M. Bader, Tissue renin-angiotensin-aldosterone systems: Targets for pharmacological therapy. *Annu. Rev. Pharmacol. Toxicol.* **50**, 439–465 (2010).
- M. de Gasparo, K. J. Catt, T. Inagami, J. W. Wright, T. Unger, International union of pharmacology. XXIII. The angiotensin II receptors. *Pharmacol. Rev.* **52**, 415–472 (2000).
- A. J. George, W. G. Thomas, R. D. Hannan, The renin-angiotensin system and cancer: Old dog, new tricks. *Nat. Rev. Cancer* **10**, 745–759 (2010).
- M. Paul, A. Poyan Mehr, R. Kreutz, Physiology of local renin-angiotensin systems. *Physiol. Rev.* **86**, 747–803 (2006).
- T. Qaradakhli, V. Apostolopoulos, A. Zulli, Angiotensin (1–7) and alamandine: Similarities and differences. *Pharmacol. Res.* **111**, 820–826 (2016).
- S. Rodrigues-Ferreira, C. Nahmias, G-protein coupled receptors of the renin-angiotensin system: New targets against breast cancer? *Front. Pharmacol.* **6**, 24 (2015).
- H. Kobori, M. Nangaku, L. G. Navar, A. Nishiyama, The intrarenal renin-angiotensin system: From physiology to the pathobiology of hypertension and kidney disease. *Pharmacol. Rev.* **59**, 251–287 (2007).
- C. M. Ferrario, Role of angiotensin II in cardiovascular disease—Therapeutic implications of more than a century of research. *J. Renin Angiotensin Aldosterone Syst.* **7**, 3–14 (2006).
- D. W. Cushman, M. A. Ondetti, History of the design of captopril and related inhibitors of angiotensin converting enzyme. *Hypertension* **17**, 589–592 (1991).
- G. Bhardwaj, How the antihypertensive losartan was discovered. *Expert Opin. Drug Discov.* **1**, 609–618 (2006).
- P. Ponikowski, A. A. Voors, S. D. Anker, H. Bueno, J. G. F. Cleland, A. J. S. Coats, V. Falk, J. R. González-Juanatey, V.-P. Harjola, E. A. Jankowska, M. Jessup, C. Linde, P. Nihoyannopoulos, J. T. Parissis, B. Pieske, J. P. Riley, G. M. C. Rosano, L. M. Ruilope, F. Ruschitzka, F. H. Rutten, P. van der Meer; Authors/Task Force Members, 2016 ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure: The Task

- Force for the diagnosis and treatment of acute and chronic heart failure of the European Society of Cardiology (ESC) Developed with the special contribution of the Heart Failure Association (HFA) of the ESC. *Eur. Heart J.* **37**, 2129–2200 (2016).
13. P. A. James, S. Oparil, B. L. Carter, W. C.ushman, C. Dennison-Himmelfarb, J. Handler, D. T. Lackland, M. L. LeFevre, T. D. MacKenzie, O. Ogedegbe, S. C. Smith Jr., L. P. Svetkey, S. J. Taler, R. R. Townsend, J. T. Wright Jr., A. S. Narva, E. Ortiz, 2014 evidence-based guideline for the management of high blood pressure in adults: Report from the panel members appointed to the Eighth Joint National Committee (JNC 8). *JAMA* **311**, 507–520 (2014).
 14. A. Levin, P. E. Stevens, Summary of KDIGO 2012 CKD Guideline: Behind the scenes, need for guidance, and a framework for moving forward. *Kidney Int.* **85**, 49–61 (2014).
 15. P. T. O'Gara, F. G. Kushner, D. D. Ascheim, D. E. Casey Jr., M. K. Chung, J. A. de Lemos, S. M. Ettinger, J. C. Fang, F. M. Fesmire, B. A. Franklin, C. B. Granger, H. M. Krumholz, J. A. Linderbaum, D. A. Morrow, L. K. Newby, J. P. Ornato, N. Ou, M. J. Radford, J. E. Tamis-Holland, C. L. Tommaso, C. M. Tracy, Y. J. Woo, D. X. Zhao, J. L. Anderson, A. K. Jacobs, J. L. Halperin, N. M. Albert, R. G. Brindis, M. A. Creager, D. DeMets, R. A. Guyton, J. S. Hochman, R. J. Kovacs, E. M. Ohman, W. G. Stevenson, C. W. Yancy; American College of Cardiology Foundation/American Heart Association Task Force on Practice Guidelines, 2013 ACCF/AHA guideline for the management of ST-elevation myocardial infarction: A report of the American College of Cardiology Foundation/American Heart Association Task Force on Practice Guidelines. *Circulation* **127**, e362–e425 (2013).
 16. M. M. Shafiq, D. V. Menon, R. G. Victor, Oral direct renin inhibition: Premise, promise, and potential limitations of a new antihypertensive drug. *Am. J. Med.* **121**, 265–271 (2008).
 17. I. Sipahi, S. M. Debanne, D. Y. Rowland, D. I. Simon, J. C. Fang, Angiotensin-receptor blockade and risk of cancer: Meta-analysis of randomised controlled trials. *Lancet Oncol.* **11**, 627–636 (2010).
 18. ARB Trialists Collaboration, Effects of telmisartan, irbesartan, valsartan, candesartan, and losartan on cancers in 15 trials enrolling 138,769 individuals. *J. Hypertens.* **29**, 623–635 (2011).
 19. S. Bangalore, S. Kumar, S. E. Kjeldsen, H. Makani, E. Grossman, J. Wetterslev, A. K. Gupta, P. S. Sever, C. Gluud, F. H. Messerli, Antihypertensive drugs and risk of cancer: Network meta-analyses and trial sequential analyses of 324,168 participants from randomised trials. *Lancet Oncol.* **12**, 65–82 (2011).
 20. FDA Drug Safety Communication: No increase in risk of cancer with certain blood pressure drugs—Angiotensin Receptor Blockers (ARBs), 15 July 2010; www.fda.gov/Drugs/DrugSafety/ucm257516.htm.
 21. W. T. Link, A. De Felice, An FDA overview of rodent carcinogenicity studies of angiotensin II AT-1 receptor blockers: Pulmonary adenomas and carcinomas. *Regul. Toxicol. Pharmacol.* **70**, 555–563 (2014).
 22. V. P. Chauhan, J. D. Martin, H. Liu, D. A. Lacorre, S. R. Jain, S. V. Kozin, T. Stylianopoulos, A. S. Mousa, X. Han, P. Adstamongkonkul, Z. Popović, P. Huang, M. G. Bawendi, Y. Boucher, R. K. Jain, Angiotensin inhibition enhances drug delivery and potentiates chemotherapy by decompressing tumour blood vessels. *Nat. Commun.* **4**, 2516 (2013).
 23. J. E. Murphy, J. Y.-L. Wo, C. Ferrone, W. Jiang, B. Y. Yeap, L. S. Blaszkowsky, E. L. Kwak, J. N. Allen, J. W. Clark, J. E. Faris, A. X. Zhu, L. Goyal, H. J. Mamon, K. D. Lillemoe, D. P. Ryan, T. F. DeLaney, C. Fernandez-del Castillo, Y. Boucher, T. S. Hong, TGF- β 1 inhibition with losartan in combination with FOLFIRINOX (F-NOX) in locally advanced pancreatic cancer (LAPC): Preliminary feasibility and R0 resection rates from a prospective phase II study. *J. Clin. Oncol.* **35** (suppl. 4S), 386 (2017).
 24. H. Liu, K. Naxerova, M. Pinter, J. Incio, H. Lee, K. Shigeta, W. W. Ho, J. A. Crain, A. Jacobson, T. Michelakos, D. Dias-Santos, A. Zanonato, T. S. Hong, J. W. Clark, J. E. Murphy, D. P. Ryan, V. Deshpande, K. D. Lillemoe, C. Fernandez-del Castillo, M. Downes, R. M. Evans, J. Michaelson, C. R. Ferrone, Y. Boucher, R. K. Jain, Use of angiotensin system inhibitors is associated with immune activation and longer survival in non-metastatic pancreatic ductal adenocarcinoma. *Clin. Cancer Res.* (2017).
 25. O. Arrieta, B. Pineda-Olvera, P. Guevara-Salazar, N. Hernández-Pedro, D. Morales-Espinosa, T. L. Cerón-Lizarraga, C. H. González-De la Rosa, D. Rembao, B. Segura-Pacheco, J. Sotelo, Expression of AT1 and AT2 angiotensin receptors in astrocytomas is associated with poor prognosis. *Br. J. Cancer* **99**, 160–166 (2008).
 26. O. Arrieta, C. Villarreal-Garza, G. Vizcaino, B. Pineda, N. Hernández-Pedro, P. Guevara-Salazar, T. Wegman-Ostrosky, G. Villanueva-Rodríguez, A. Gamboa-Domínguez, Association between AT1 and AT2 angiotensin II receptor expression with cell proliferation and angiogenesis in operable breast cancer. *Tumour Biol.* **36**, 5627–5634 (2015).
 27. K. Ino, K. Shibata, H. Kajiyama, E. Yamamoto, T. Nagasaka, A. Nawa, S. Nomura, F. Kikkawa, Angiotensin II type 1 receptor expression in ovarian cancer and its correlation with tumour angiogenesis and patient survival. *Br. J. Cancer* **94**, 552–560 (2006).
 28. C. Rocken, F. W. Rohl, E. Diebler, U. Lendeckel, M. Pross, S. Carl-McGrath, M. P. Ebert, The angiotensin II/angiotensin II receptor system correlates with nodal spread in intestinal type gastric cancer. *Cancer Epidemiol. Biomarkers Prev.* **16**, 1206–1212 (2007).
 29. S. Shirotake, A. Miyajima, T. Kosaka, N. Tanaka, T. Maeda, E. Kikuchi, M. Oya, Angiotensin II type 1 receptor expression and microvessel density in human bladder cancer. *Urology* **77**, 1009.e19–1009.e25 (2011).
 30. I. V. Balyasnikova, S. M. Danilov, V. R. Muzykantov, A. B. Fisher, Modulation of angiotensin-converting enzyme in cultured human vascular endothelial cells. *In Vitro Cell. Dev. Biol. Anim.* **34**, 545–554 (1998).
 31. S. M. Danilov, E. Sadovnikova, N. Scharenborg, I. V. Balyasnikova, D. A. Svinareva, E. L. Semikina, E. N. Parovichnikova, V. G. Savchenko, G. J. Adema, Angiotensin-converting enzyme (CD143) is abundantly expressed by dendritic cells and discriminates human monocyte-derived dendritic cells from acute myeloid leukemia-derived dendritic cells. *Exp. Hematol.* **31**, 1301–1309 (2003).
 32. C. Lin, V. Datta, D. Okwan-Duodu, X. Chen, S. Fuchs, R. Alsabeh, S. Billet, K. E. Bernstein, X. Z. Shen, Angiotensin-converting enzyme is required for normal myelopoiesis. *FASEB J.* **25**, 1145–1155 (2011).
 33. X. Z. Shen, S. Billet, C. Lin, D. Okwan-Duodu, X. Chen, A. E. Lukacher, K. E. Bernstein, The carboxypeptidase ACE shapes the MHC class I peptide repertoire. *Nat. Immunol.* **12**, 1078–1085 (2011).
 34. V. Cortez-Retamozo, M. Etzrodt, A. Newton, R. Ryan, F. Pucci, S. W. Sio, W. Kuswanto, P. J. Rauch, A. Chudnovskiy, Y. Iwamoto, R. Kohler, B. Marinelli, R. Gorbатов, G. Wojtkiewicz, P. Panizzi, M. Mino-Kenudson, R. Forghani, J.-L. Figueiredo, J. W. Chen, R. Xavier, F. K. Swirski, M. Nahrendorf, R. Weissleder, M. J. Pittet, Angiotensin II drives the production of tumor-promoting macrophages. *Immunity* **38**, 296–308 (2013).
 35. H. Azevedo, A. Fujita, S. Y. Bando, P. Iamashita, C. A. Moreira-Filho, Transcriptional network analysis reveals that AT1 and AT2 angiotensin II receptors are both involved in the regulation of genes essential for glioma progression. *PLOS ONE* **9**, e110934 (2014).
 36. X. Li, H. Zhang, V. Soledad-Conrad, J. Zhuang, B. D. Uhal, Bleomycin-induced apoptosis of alveolar epithelial cells requires angiotensin synthesis de novo. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **284**, L501–L507 (2003).
 37. L. Nguyen, E. I. Ager, J. Neo, C. Christophi, Regulation of colorectal cancer cell epithelial to mesenchymal transition by the renin angiotensin system. *J. Gastroenterol. Hepatol.* **31**, 1773–1782 (2016).
 38. M. Papp, X. Li, J. Zhuang, R. Wang, B. D. Uhal, Angiotensin receptor subtype AT₁ mediates alveolar epithelial cell apoptosis in response to ANG II. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **282**, L713–L718 (2002).
 39. S. Zheng, Y. Yang, R. Song, X. Yang, H. Liu, Q. Ma, L. Yang, R. Meng, T. Tao, S. Wang, J. He, Ang-(1–7) promotes the migration and invasion of human renal cell carcinoma cells via Mas-mediated AKT signaling pathway. *Biochem. Biophys. Res. Commun.* **460**, 333–340 (2015).
 40. M. J. Smyth, S. F. Ngiew, A. Ribas, M. W. L. Teng, Combination cancer immunotherapies tailored to the tumour microenvironment. *Nat. Rev. Clin. Oncol.* **13**, 143–158 (2016).
 41. D. H. Munn, V. Bronte, Immune suppressive mechanisms in the tumor microenvironment. *Curr. Opin. Immunol.* **39**, 1–6 (2016).
 42. B. Diop-Frimpong, V. P. Chauhan, S. Krane, Y. Boucher, R. K. Jain, Losartan inhibits collagen I synthesis and improves the distribution and efficacy of nanotherapeutics in tumors. *Proc. Natl. Acad. Sci. U.S.A.* **108**, 2909–2914 (2011).
 43. D. Öhlund, E. Elyada, D. Tuveson, Fibroblast heterogeneity in the cancer wound. *J. Exp. Med.* **211**, 1503–1523 (2014).
 44. M. O. Li, R. A. Flavell, TGF- β : A master of all T cell trades. *Cell* **134**, 392–404 (2008).
 45. J. Watt, H. M. Kocher, The desmoplastic stroma of pancreatic cancer is a barrier to immune cell infiltration. *Oncotmunology* **2**, e26788 (2013).
 46. R. K. Jain, Normalizing tumor microenvironment to treat cancer: Bench to bedside to biomarkers. *J. Clin. Oncol.* **31**, 2205–2218 (2013).
 47. R. K. Jain, J. D. Martin, T. Stylianopoulos, The role of mechanical forces in tumor growth and therapy. *Annu. Rev. Biomed. Eng.* **16**, 321–346 (2014).
 48. R. K. Jain, Antiangiogenesis strategies revisited: From starving tumors to alleviating hypoxia. *Cancer Cell* **26**, 605–622 (2014).
 49. M. Z. Noman, M. Hasmim, Y. Messai, S. Terry, C. Kieda, B. Janji, S. Chouaib, Hypoxia: A key player in antitumor immune response. A review in the theme: Cellular responses to hypoxia. *Am. J. Physiol. Cell Physiol.* **309**, C569–C579 (2015).
 50. A. Palazón, J. Aragonés, A. Morales-Kastresana, M. O. de Landázuri, I. Melero, Molecular pathways: Hypoxia response in immune cells fighting or promoting cancer. *Clin. Cancer Res.* **18**, 1207–1213 (2012).
 51. T. Voron, O. Colussi, E. Marcheteau, S. Pernot, M. Nizard, A.-L. Pointet, S. Latreche, S. Bergaya, N. Benhamouda, C. Tanchot, C. Stockmann, P. Combe, A. Berger, F. Zinzindohoue, H. Yagita, E. Tartour, J. Taieb, M. Terme, VEGF-A modulates expression of inhibitory checkpoints on CD8⁺ T cells in tumors. *J. Exp. Med.* **212**, 139–148 (2015).
 52. Y. Chen, R. R. Ramjiawan, T. Reiberger, M. R. Ng, T. Hato, Y. Huang, H. Ochiai, S. Kitahara, E. C. Unan, T. P. Reddy, C. Fan, P. Huang, N. Bardeesy, A. X. Zhu, R. K. Jain, D. G. Duda, CXCR4 inhibition in tumor microenvironment facilitates anti-programmed death receptor-1 immunotherapy in sorafenib-treated hepatocellular carcinoma in mice. *Hepatology* **61**, 1591–1602 (2015).

53. C. Feig, J. O. Jones, M. Kraman, R. J. B. Wells, A. Deonarine, D. S. Chan, C. M. Connell, E. W. Roberts, Q. Zhao, O. L. Caballero, S. A. Teichmann, T. Janowitz, D. I. Jodrell, D. A. Tuveson, D. T. Fearon, Targeting CXCL12 from FAP-expressing carcinoma-associated fibroblasts synergizes with anti-PD-L1 immunotherapy in pancreatic cancer. *Proc. Natl. Acad. Sci. U.S.A.* **110**, 20212–20217 (2013).
54. H. Jiang, S. Hegde, B. L. Knolhoff, Y. Zhu, J. M. Herndon, M. A. Meyer, T. M. Nywening, W. G. Hawkins, I. M. Shapiro, D. T. Weaver, J. A. Pachter, A. Wang-Gillam, D. G. DeNardo, Targeting focal adhesion kinase renders pancreatic cancers responsive to checkpoint immunotherapy. *Nat. Med.* **22**, 851–860 (2016).
55. A. Masamune, S. Hamada, K. Kikuta, S. Miura, E. Nakano, T. Shimosegawa, The angiotensin II type 1 receptor blocker olmesartan inhibits the growth of pancreatic cancer by targeting stellate cell activities in mice. *Scand. J. Gastroenterol.* **48**, 602–609 (2013).
56. M. Okazaki, S. Fushida, S. Harada, T. Tsukada, J. Kinoshita, K. Oyama, H. Tajima, I. Ninomiya, T. Fujimura, T. Ohta, The angiotensin II type 1 receptor blocker candesartan suppresses proliferation and fibrosis in gastric cancer. *Cancer Lett.* **355**, 46–53 (2014).
57. C. Godugu, A. R. Patel, R. Doddapaneni, S. Marepally, T. Jackson, M. Singh, Inhalation delivery of Telmisartan enhances intratumoral distribution of nanoparticles in lung cancer models. *J. Control. Release* **172**, 86–95 (2013).
58. K. Patel, R. Doddapaneni, N. Chowdhury, C. H. A. Boakye, G. Behl, M. Singh, Tumor stromal disrupting agent enhances the anticancer efficacy of docetaxel loaded STGylated liposomes in lung cancer. *Nanomedicine* **11**, 1377–1392 (2016).
59. J. Incio, H. Liu, P. Suboj, S. M. Chin, I. X. Chen, M. Pinter, M. R. Ng, H. T. Nia, J. Grahovac, S. Kao, S. Babykutty, Y. Huang, K. Jung, N. N. Rahbari, X. Han, V. P. Chauhan, J. D. Martin, J. Kahn, P. Huang, V. Desphande, J. Michaelson, T. P. Michelakos, C. R. Ferrone, R. Soares, Y. Boucher, D. Fukumura, R. K. Jain, Obesity-induced inflammation and desmoplasia promote pancreatic cancer progression and resistance to chemotherapy. *Cancer Discov.* **6**, 852–869 (2016).
60. S. A. Arnold, L. B. Rivera, J. G. Carbon, J. E. Toombs, C.-L. Chang, A. D. Bradshaw, R. A. Brekken, Losartan slows pancreatic tumor progression and extends survival of SPARC-null mice by abrogating aberrant TGF β activation. *PLOS ONE* **7**, e31384 (2012).
61. R. Anandanadesan, Q. Gong, G. Chipitsyna, A. Witkiewicz, C. J. Yeo, H. A. Arafat, Angiotensin II induces vascular endothelial growth factor in pancreatic cancer cells through an angiotensin II type 1 receptor and ERK1/2 signaling. *J. Gastrointest. Surg.* **12**, 57–66 (2008).
62. Y. Ji, Z. Wang, Z. Li, K. Li, X. Le, T. Zhang, Angiotensin II induces angiogenic factors production partly via AT1/JAK2/STAT3/SOCS2 signaling pathway in MHCC97H cells. *Cell. Physiol. Biochem.* **29**, 863–874 (2012).
63. T. Kosaka, A. Miyajima, S. Shirotake, E. Kikuchi, M. Hasegawa, S. Mikami, M. Oya, Ets-1 and hypoxia inducible factor-1 α inhibition by angiotensin II type-1 receptor blockade in hormone-refractory prostate cancer. *Prostate* **70**, 162–169 (2010).
64. M. Fujita, I. Hayashi, S. Yamashina, A. Fukamizu, M. Itoman, M. Majima, Angiotensin type 1a receptor signaling-dependent induction of vascular endothelial growth factor in stroma is relevant to tumor-associated angiogenesis and tumor growth. *Carcinogenesis* **26**, 271–279 (2005).
65. M. Kosugi, A. Miyajima, E. Kikuchi, Y. Horiguchi, M. Murai, Angiotensin II type 1 receptor antagonist candesartan as an angiogenic inhibitor in a xenograft model of bladder cancer. *Clin. Cancer Res.* **12**, 2888–2893 (2006).
66. H. Yoshiji, S. Kuriyama, M. Kawata, Y. Yoshii, Y. Ikenaka, R. Noguchi, T. Nakatani, H. Tsujinoue, H. Fukui, The angiotensin-I-converting enzyme inhibitor perindopril suppresses tumor growth and angiogenesis: Possible role of the vascular endothelial growth factor. *Clin. Cancer Res.* **7**, 1073–1078 (2001).
67. R. K. Jain, Determinants of tumor blood flow: A review. *Cancer Res.* **48**, 2641–2658 (1988).
68. Y. Huang, S. Goel, D. G. Duda, D. Fukumura, R. K. Jain, Vascular normalization as an emerging strategy to enhance cancer immunotherapy. *Cancer Res.* **73**, 2943–2948 (2013).
69. Y. Huang, J. Yuan, E. Righi, W. S. Kamoun, M. Ancukiewicz, J. Nezivar, M. Santosuosso, J. D. Martin, M. R. Martin, F. Vianello, P. Leblanc, L. L. Munn, P. Huang, D. G. Duda, D. Fukumura, R. K. Jain, M. C. Poznansky, Vascular normalizing doses of antiangiogenic treatment reprogram the immunosuppressive tumor microenvironment and enhance immunotherapy. *Proc. Natl. Acad. Sci. U.S.A.* **109**, 17561–17566 (2012).
70. T. T. Batchelor, E. R. Gerstner, K. E. Emblem, D. G. Duda, J. Kalpathy-Cramer, M. Snuderl, M. Ancukiewicz, P. Polaskova, M. C. Pinho, D. Jennings, S. R. Plotkin, A. S. Chi, A. F. Eichler, J. Dietrich, F. H. Hochberg, C. Lu-Emerson, A. J. Iafra, S. P. Ivy, B. R. Rosen, J. S. Loeffler, P. Y. Wen, A. G. Sorensen, R. K. Jain, Improved tumor oxygenation and survival in glioblastoma patients who show increased blood perfusion after cediranib and chemoradiation. *Proc. Natl. Acad. Sci. U.S.A.* **110**, 19059–19064 (2013).
71. K. E. Emblem, K. Mouridsen, A. Bjornerud, C. T. Farrar, D. Jennings, R. J. H. Borra, P. Y. Wen, P. Ivy, T. T. Batchelor, B. R. Rosen, R. K. Jain, A. G. Sorensen, Vessel architectural imaging identifies cancer patient responders to anti-angiogenic therapy. *Nat. Med.* **19**, 1178–1183 (2013).
72. A. G. Sorensen, K. E. Emblem, P. Polaskova, D. Jennings, H. Kim, M. Ancukiewicz, M. Wang, P. Y. Wen, P. Ivy, T. T. Batchelor, R. K. Jain, Increased survival of glioblastoma patients who respond to antiangiogenic therapy with elevated blood perfusion. *Cancer Res.* **72**, 402–407 (2012).
73. R. E. Gilbert, D. J. Kelly, A. J. Cox, J. L. Wilkinson-Berka, J. R. Rumble, T. Osicka, S. Panagiotopoulos, V. Lee, E. C. Hendrich, G. Jerums, M. E. Cooper, Angiotensin converting enzyme inhibition reduces retinal overexpression of vascular endothelial growth factor and hyperpermeability in experimental diabetes. *Diabetologia* **43**, 1360–1367 (2000).
74. H. Sano, K. Hosokawa, H. Kidoya, N. Takakura, Negative regulation of VEGF-induced vascular leakage by blockade of angiotensin II type 1 receptor. *Arterioscler. Thromb. Vasc. Biol.* **26**, 2673–2680 (2006).
75. K. E. Emblem, E. R. Gerstner, G. Sorensen, B. R. Rosen, P. Y. Wen, T. T. Batchelor, R. K. Jain, Abstract 3975: Matrix-depleting anti-hypertensives decompress tumor blood vessels and improve perfusion in patients with glioblastoma receiving anti-angiogenic therapy. *Cancer Res.* **76** (suppl. 14), 3975 (2016).
76. K. M. Bell, V. E. Prise, K. M. Shaffi, D. J. Chaplin, G. M. Tozer, A comparative study of tumour-blood-flow modification in two rat-tumour systems using endothelin-1 and angiotensin II: Influence of tumour size on angiotensin-II response. *Int. J. Cancer* **67**, 730–738 (1996).
77. G. M. Tozer, K. M. Shaffi, The response of tumour vasculature to angiotensin II revealed by its systemic and local administration to 'tissue-isolated' tumours. *Br. J. Cancer* **72**, 595–600 (1995).
78. R. A. Zlotnicki, L. T. Baxter, Y. Boucher, R. K. Jain, Pharmacologic modification of tumor blood flow and interstitial fluid pressure in a human tumor xenograft: Network analysis and mechanistic interpretation. *Microvasc. Res.* **50**, 429–443 (1995).
79. R. A. Zlotnicki, Y. Boucher, I. Lee, L. T. Baxter, R. K. Jain, Effect of angiotensin II induced hypertension on tumor blood flow and interstitial fluid pressure. *Cancer Res.* **53**, 2466–2468 (1993).
80. O. Thews, D. K. Kelleher, P. Vaupel, Disparate responses of tumour vessels to angiotensin II: Tumour volume-dependent effects on perfusion and oxygenation. *Br. J. Cancer* **83**, 225–231 (2000).
81. D. Okwan-Duodu, J. Landry, X. Z. Shen, R. Diaz, Angiotensin-converting enzyme and the tumor microenvironment: Mechanisms beyond angiogenesis. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **305**, R205–R215 (2013).
82. F. R. Balkwill, A. Mantovani, Cancer-related inflammation: Common themes and therapeutic opportunities. *Semin. Cancer Biol.* **22**, 33–40 (2012).
83. Y. Ji, Z. Wang, Z. Li, A. Zhang, Y. Jin, H. Chen, X. Le, Angiotensin II enhances proliferation and inflammation through AT1/PKC/NF- κ B signaling pathway in hepatocellular carcinoma cells. *Cell. Physiol. Biochem.* **39**, 13–32 (2016).
84. H. Uemura, H. Ishiguro, Y. Nagashima, T. Sasaki, N. Nakaigawa, H. Hasumi, S. Kato, Y. Kubota, Antiproliferative activity of angiotensin II receptor blocker through cross-talk between stromal and epithelial prostate cancer cells. *Mol. Cancer Ther.* **4**, 1699–1709 (2005).
85. T. Matsuzuka, K. Miller, L. Pickel, C. Doi, R. Ayuzawa, M. Tamura, The synergistic induction of cyclooxygenase-2 in lung fibroblasts by angiotensin II and pro-inflammatory cytokines. *Mol. Cell. Biochem.* **320**, 163–171 (2009).
86. H. Pham, B. Chong, R. Vincenti, L. W. Slice, Ang II and EGF synergistically induce COX-2 expression via CREB in intestinal epithelial cells. *J. Cell. Physiol.* **214**, 96–109 (2008).
87. L. W. Slice, T. Chiu, E. Rozengurt, Angiotensin II and epidermal growth factor induce cyclooxygenase-2 expression in intestinal epithelial cells through small GTPases using distinct signaling pathways. *J. Biol. Chem.* **280**, 1582–1593 (2005).
88. J. M. David, C. Dominguez, D. H. Hamilton, C. Palena, The IL-8/IL-8R axis: A double agent in tumor immune resistance. *Vaccines* **4**, E22 (2016).
89. F. Martin, L. Apetoh, F. Ghiringhelli, Controversies on the role of Th17 in cancer: A TGF- β -dependent immunosuppressive activity? *Trends Mol. Med.* **18**, 742–749 (2012).
90. S. Ugel, F. De Sanctis, S. Mandruzzato, V. Bronte, Tumor-induced myeloid deviation: When myeloid-derived suppressor cells meet tumor-associated macrophages. *J. Clin. Invest.* **125**, 3365–3376 (2015).
91. D. S. Vinay, E. P. Ryan, G. Pawelec, W. H. Talib, J. Stagg, E. Elkord, T. Lichter, W. K. Decker, R. L. Whelan, H. M. C. S. Kumara, E. Signori, K. Honoki, A. G. Georgakilas, A. Amin, W. G. Helferich, C. S. Boosani, G. Guha, M. R. Ciriolo, S. Chen, S. I. Mohammed, A. S. Azmi, W. N. Keith, A. Bilisland, D. Bhakta, D. Halicka, H. Fujii, K. Aquilano, S. S. Ashraf, S. Nowshheen, X. Yang, B. K. Choi, B. S. Kwon, Immune evasion in cancer: Mechanistic basis and therapeutic strategies. *Semin. Cancer Biol.* **35** (Suppl.), S185–S198 (2015).
92. J. R. Brown, R. N. DuBois, COX-2: A molecular target for colorectal cancer prevention. *J. Clin. Oncol.* **23**, 2840–2855 (2005).
93. S. Zelenay, A. G. van der Veer, J. P. Böttcher, K. J. Snelgrove, N. Rogers, S. E. Acton, P. Chakravarty, M. R. Girotti, R. Marais, S. A. Quezada, E. Sahai, C. Reis e Sousa, Cyclooxygenase-dependent tumor growth through evasion of immunity. *Cell* **162**, 1257–1270 (2015).

94. H. Frenzel, R. Pries, C. P. Brocks, W. J. Jabs, N. Wittkopf, B. Wollenberg, Decreased migration of myeloid dendritic cells through increased levels of C-reactive protein. *Anticancer Res.* **27**, 4111–4115 (2007).
95. S. H. Jackson, S. Devadas, J. Kwon, L. A. Pinto, M. S. Williams, T cells express a phagocyte-type NADPH oxidase that is activated after T cell receptor stimulation. *Nat. Immunol.* **5**, 818–827 (2004).
96. L. A. Sena, S. Li, A. Jairaman, M. Prakriya, T. Ezponda, D. A. Hildeman, C.-R. Wang, P. T. Schumacker, J. D. Licht, H. Perlman, P. J. Bryce, N. S. Chandel, Mitochondria are required for antigen-specific T cell activation through reactive oxygen species signaling. *Immunity* **38**, 225–236 (2013).
97. S. I. Gringhuis, A. Leow, E. A. M. Papendrecht-van der Voort, P. H. J. Remans, F. C. Breedveld, C. L. Verweij, Displacement of linker for activation of T cells from the plasma membrane due to redox balance alterations results in hyporesponsiveness of synovial fluid T lymphocytes in rheumatoid arthritis. *J. Immunol.* **164**, 2170–2179 (2000).
98. N. Lahdenperhja, K. Savinainen, M. Hurme, Pre-exposure to oxidative stress decreases the nuclear factor- κ B-dependent transcription in T lymphocytes. *J. Immunol.* **160**, 1354–1358 (1998).
99. H.-R. Kim, A. Lee, E.-J. Choi, M.-P. Hong, J.-H. Kie, W. Lim, H. K. Lee, B.-I. Moon, J.-Y. Seoh, Reactive oxygen species prevent imiquimod-induced psoriatic dermatitis through enhancing regulatory T cell function. *PLOS ONE* **9**, e91146 (2014).
100. X. Lin, W. Zheng, J. Liu, Y. Zhang, H. Qin, H. Wu, B. Xue, Y. Lu, P. Shen, Oxidative stress in malignant melanoma enhances tumor necrosis factor- α secretion of tumor-associated macrophages that promote cancer cell invasion. *Antioxid. Redox Signal.* **19**, 1337–1355 (2013).
101. A. Sica, T. Schioppa, A. Mantovani, P. Allavena, Tumour-associated macrophages are a distinct M2 polarised population promoting tumour progression: Potential targets of anti-cancer therapy. *Eur. J. Cancer* **42**, 717–727 (2006).
102. H. Uemura, H. Ishiguro, Y. Ishiguro, K. Hoshino, S. Takahashi, Y. Kubota, Angiotensin II induces oxidative stress in prostate cancer. *Mol. Cancer Res.* **6**, 250–258 (2008).
103. S. Shirotake, A. Miyajima, T. Kosaka, N. Tanaka, E. Kikuchi, S. Mikami, Y. Okada, M. Oya, Regulation of monocyte chemoattractant protein-1 through angiotensin II type 1 receptor in prostate cancer. *Am. J. Pathol.* **180**, 1008–1016 (2012).
104. K. Egami, T. Murohara, T. Shimada, K.-i. Sasaki, S. Shintani, T. Sugaya, M. Ishii, T. Akagi, H. Ikeda, T. Matsuishi, T. Imaizumi, Role of host angiotensin II type 1 receptor in tumor angiogenesis and growth. *J. Clin. Invest.* **112**, 67–75 (2003).
105. N. Chehl, Q. Gong, G. Chipitsyna, T. Aziz, C. J. Yeo, H. A. Arafat, Angiotensin II regulates the expression of monocyte chemoattractant protein-1 in pancreatic cancer cells. *J. Gastrointest. Surg.* **13**, 2189–2200 (2009).
106. X. Z. Shen, K. E. Bernstein, The peptide network regulated by angiotensin converting enzyme (ACE) in hematopoiesis. *Cell Cycle* **10**, 1363–1369 (2011).
107. X. Z. Shen, D. Okwan-Duodu, W.-L. Blackwell, F. S. Ong, T. Janjulia, E. A. Bernstein, S. Fuchs, S. Alkan, K. E. Bernstein, Myeloid expression of angiotensin-converting enzyme facilitates myeloid maturation and inhibits the development of myeloid-derived suppressor cells. *Lab. Invest.* **94**, 536–544 (2014).
108. X. Z. Shen, P. Li, D. Weiss, S. Fuchs, H. D. Xiao, J. A. Adams, I. R. Williams, M. R. Capecchi, W. R. Taylor, K. E. Bernstein, Mice with enhanced macrophage angiotensin-converting enzyme are resistant to melanoma. *Am. J. Pathol.* **170**, 2122–2134 (2007).
109. X. Z. Shen, A. E. Lukacher, S. Billet, I. R. Williams, K. E. Bernstein, Expression of angiotensin-converting enzyme changes major histocompatibility complex class I peptide presentation by modifying C termini of peptide precursors. *J. Biol. Chem.* **283**, 9957–9965 (2008).
110. L. Hofmann, A. Forschner, C. Loquai, S. M. Goldinger, L. Zimmer, S. Ugurel, M. I. Schmidgen, R. Gutzmer, J. S. Utikal, D. Göppner, J. C. Hassel, F. Meier, J. K. Tietze, I. Thomas, C. Weishaupt, M. Leverkus, R. Wahl, U. Dietrich, C. Garbe, M. C. Kirchberger, T. Eigentler, C. Berking, A. Gesierich, A. M. Krackhardt, D. Schadendorf, G. Schuler, R. Dummer, L. M. Heinzerling, Cutaneous, gastrointestinal, hepatic, endocrine, and renal side-effects of anti-PD-1 therapy. *Eur. J. Cancer* **60**, 190–209 (2016).
111. M. A. Postow, J. Chesney, A. C. Pavlick, C. Robert, K. Grossmann, D. McDermott, G. P. Linette, N. Meyer, J. K. Giguere, S. S. Agarwala, M. Shaheen, M. S. Ernstoff, D. Minor, A. K. Salama, M. Taylor, P. A. Ott, L. M. Rollin, C. Horak, P. Gagnier, J. D. Wolchok, F. S. Hodi, Nivolumab and ipilimumab versus ipilimumab in untreated melanoma. *N. Engl. J. Med.* **372**, 2006–2017 (2015).
112. M. N. Bouchlaka, G. D. Skisel, M. Chen, A. Mirsoian, A. E. Zamora, E. Mavarakis, D. E. C. Wilkins, K. L. Alderson, H.-H. Hsiao, J. M. Weiss, A. M. Monjazeb, C. Hesdorffer, L. Ferrucci, D. L. Longo, B. R. Blazar, R. H. Wiltrout, D. Redelman, D. D. Taub, W. J. Murphy, Aging predisposes to acute inflammatory induced pathology after tumor immunotherapy. *J. Exp. Med.* **210**, 2223–2237 (2013).
113. A. Mirsoian, M. N. Bouchlaka, G. D. Skisel, M. Chen, C.-C. S. Pai, E. Mavarakis, R. G. Spencer, K. W. Fishbein, S. Siddiqui, A. M. Monjazeb, B. Martin, S. Maudsley, C. Hesdorffer, L. Ferrucci, D. L. Longo, B. R. Blazar, R. H. Wiltrout, D. D. Taub, W. J. Murphy, Adiposity induces lethal cytokine storm after systemic administration of stimulatory immunotherapy regimens in aged mice. *J. Exp. Med.* **211**, 2373–2383 (2014).
114. D. Fliser, K. Buchholz, H. Haller; European Trial on Olmesartan and Pravastatin in Inflammation and Atherosclerosis (EUROPIA) Investigators, Antiinflammatory effects of angiotensin II subtype 1 receptor blockade in hypertensive patients with microinflammation. *Circulation* **110**, 1103–1107 (2004).
115. S. Manabe, T. Okura, S. Watanabe, T. Fukuoka, J. Higaki, Effects of angiotensin II receptor blockade with valsartan on pro-inflammatory cytokines in patients with essential hypertension. *J. Cardiovasc. Pharmacol.* **46**, 735–739 (2005).
116. M. G. Pavlatou, G. Mastorakos, A. Margeli, E. Kouskouni, N. Tentolouris, N. Katsilambros, G. P. Chrousos, I. Papassotiropoulos, Angiotensin blockade in diabetic patients decreases insulin resistance-associated low-grade inflammation. *Eur. J. Clin. Invest.* **41**, 652–658 (2011).
117. M. D. Holmes, S. E. Hankinson, D. Feskanich, W. Y. Chen, Beta blockers and angiotensin-converting enzyme inhibitors' purported benefit on breast cancer survival may be explained by aspirin use. *Breast Cancer Res. Treat.* **139**, 507–513 (2013).
118. P. H. Jones, K. Christodoulos, N. Dobbs, P. Thavasu, F. Balkwill, A. D. Blann, G. J. Caine, S. Kumar, A. J. Kakkar, N. Gompertz, D. C. Talbot, T. S. Ganesan, A. L. Harris, Combination antiangiogenesis therapy with marimastat, captopril and fragmin in patients with advanced cancer. *Br. J. Cancer* **91**, 30–36 (2004).
119. Y. Nakai, H. Isayama, H. Ijichi, T. Sasaki, H. Kogure, H. Yagioka, K. Miyabayashi, S. Mizuno, K. Yamamoto, D. Mouri, K. Kawakubo, N. Yamamoto, K. Hirano, N. Sasahira, K. Tateishi, M. Tada, K. Koike, Phase I trial of gemcitabine and candesartan combination therapy in normotensive patients with advanced pancreatic cancer: GECA1. *Cancer Sci.* **103**, 1489–1492 (2012).
120. Y. Nakai, H. Isayama, H. Ijichi, T. Sasaki, N. Takahara, Y. Ito, S. Matsubara, R. Uchino, H. Yagioka, T. Arizumi, T. Hamada, K. Miyabayashi, S. Mizuno, K. Yamamoto, H. Kogure, N. Yamamoto, K. Hirano, N. Sasahira, K. Tateishi, M. Tada, K. Koike, A multicenter phase II trial of gemcitabine and candesartan combination therapy in patients with advanced pancreatic cancer: GECA2. *Invest. New Drugs* **31**, 1294–1299 (2013).
121. G. Ronquist, G. Frithz, Y.-H. Wang, T. Lindeborg, Captopril may reduce biochemical (prostate-specific antigen) failure following radical prostatectomy for clinically localized prostate cancer. *Scand. J. Urol. Nephrol.* **43**, 32–36 (2009).
122. G. V. Sørensen, P. A. Ganz, S. W. Cole, L. A. Pedersen, H. T. Sørensen, D. P. Cronin-Fenton, J. P. Garne, P. M. Christiansen, T. L. Lash, T. P. Ahern, Use of β -blockers, angiotensin-converting enzyme inhibitors, angiotensin II receptor blockers, and risk of breast cancer recurrence: A Danish nationwide prospective cohort study. *J. Clin. Oncol.* **31**, 2265–2272 (2013).
123. M. Tatokoro, Y. Fujii, S. Kawakami, K. Saito, F. Koga, Y. Matsuoka, Y. Iimura, H. Masuda, K. Kihara, Phase-II trial of combination treatment of interferon- α , cimetidine, cyclooxygenase-2 inhibitor and renin-angiotensin-system inhibitor (I-CCA therapy) for advanced renal cell carcinoma. *Cancer Sci.* **102**, 137–143 (2011).
124. H. Uemura, H. Hasumi, T. Kawahara, S. Sugiura, Y. Miyoshi, N. Nakaigawa, J.-i. Teranishi, K. Noguchi, H. Ishiguro, Y. Kubota, Pilot study of angiotensin II receptor blocker in advanced hormone-refractory prostate cancer. *Int. J. Clin. Oncol.* **10**, 405–410 (2005).
125. H. Yoshiji, R. Noguchi, Y. Ikenaka, K. Kaji, Y. Aihara, M. Yamazaki, J. Yamao, M. Toyohara, A. Mito, M. Sawai, M. Yoshida, C. Morioka, M. Fujimoto, M. Uemura, H. Fukui, Combination of branched-chain amino acids and angiotensin-converting enzyme inhibitor suppresses the cumulative recurrence of hepatocellular carcinoma: A randomized control trial. *Oncol. Rep.* **26**, 1547–1553 (2011).
126. H. Yoshiji, R. Noguchi, M. Toyohara, Y. Ikenaka, M. Kitade, K. Kaji, M. Yamazaki, J. Yamao, A. Mito, M. Sawai, M. Yoshida, M. Fujimoto, T. Tsujimoto, H. Kawarata, M. Uemura, H. Fukui, Combination of vitamin K₂ and angiotensin-converting enzyme inhibitor ameliorates cumulative recurrence of hepatocellular carcinoma. *J. Hepatol.* **51**, 315–321 (2009).
127. A. Alashkham, C. Paterson, P. Windsor, A. Struthers, P. Rauchhaus, G. Nabi, The incidence and risk of biochemical recurrence following radical radiotherapy for prostate cancer in men on angiotensin-converting enzyme inhibitors (ACEIs) or angiotensin receptor blockers (ARBs). *Clin. Genitourin. Cancer* **14**, 398–405 (2016).
128. A. Aydiner, R. Ciftci, F. Sen, Renin-Angiotensin system blockers may prolong survival of metastatic non-small cell lung cancer patients receiving erlotinib. *Medicine* **94**, e887 (2015).
129. T. Babacan, O. Balakan, T. Y. Kuzan, F. Sarici, E. Koca, N. Kertmen, I. Petekaya, K. Altundag, The effect of renin-angiotensin-system inhibition on survival and recurrence of N3+ breast cancer patients. *J. BUON* **20**, 50–56 (2015).
130. M. L. Blute Jr., T. J. Rushmer, F. Shi, B. J. Fuller, E. J. Abel, D. F. Jarrard, T. M. Downs, Renin-angiotensin inhibitors decrease recurrence after transurethral resection of bladder tumor in patients with nonmuscle invasive bladder cancer. *J. Urol.* **194**, 1214–1219 (2015).
131. E. Botteri, E. Munzone, N. Rotmensz, C. Cipolla, V. De Giorgi, B. Santillo, A. Zanelotti, L. Adami, M. Colleoni, G. Viale, A. Goldhirsch, S. Gandini, Therapeutic effect of β -blockers in triple-negative breast cancer postmenopausal women. *Breast Cancer Res. Treat.* **140**, 567–575 (2013).

132. D. M. Boudreau, O. Yu, J. Chubak, H. S. Wirtz, E. J. A. Bowles, M. Fujii, D. S. M. Buist, Comparative safety of cardiovascular medication use and breast cancer outcomes among women with early stage breast cancer. *Breast Cancer Res. Treat.* **144**, 405–416 (2014).
133. Y. K. Chae, E. N. Brown, X. Lei, A. Melhem-Bertrandt, S. H. Giordano, J. K. Litton, G. N. Hortobagyi, A. M. Gonzalez-Angulo, M. Chavez-MacGregor, Use of ACE inhibitors and angiotensin receptor blockers and primary breast cancer outcomes. *J. Cancer* **4**, 549–556 (2013).
134. Y. K. Chae, M. E. Valsecchi, J. Kim, A. L. Bianchi, D. Khemasuwan, A. Desai, W. Tester, Reduced risk of breast cancer recurrence in patients using ACE inhibitors, ARBs, and/or statins. *Cancer Invest.* **29**, 585–593 (2011).
135. P. A. Ganz, L. A. Habel, E. K. Weltzien, B. J. Caan, S. W. Cole, Examining the influence of beta blockers and ACE inhibitors on the risk for breast cancer recurrence: Results from the LACE cohort. *Breast Cancer Res. Treat.* **129**, 549–556 (2011).
136. H. Goldvaser, S. Rizel, D. Hendler, V. Neiman, D. Shepshelovich, T. Shochat, A. Sulkes, B. Brenner, R. Yerushalmi, The association between angiotensin receptor blocker usage and breast cancer characteristics. *Oncology* **91**, 217–223 (2016).
137. H. Izzedine, L. Derosa, G. Le Teuff, L. Albiges, B. Escudier, Hypertension and angiotensin system inhibitors: Impact on outcome in sunitinib-treated patients for metastatic renal cell carcinoma. *Ann. Oncol.* **26**, 1128–1133 (2015).
138. D. Keizman, M. Gottfried, M. Ish-Shalom, N. Maimon, A. Peer, A. Neumann, H. Hammers, M. A. Eisenberger, V. Sinibaldi, R. Pili, H. Hayat, S. Kovel, A. Sella, B. Boursi, R. Weitzen, W. Mermershtain, K. Rouvinov, R. Berger, M. A. Carducci, Active smoking may negatively affect response rate, progression-free survival, and overall survival of patients with metastatic renal cell carcinoma treated with sunitinib. *Oncologist* **19**, 51–60 (2014).
139. D. Keizman, P. Huang, M. A. Eisenberger, R. Pili, J. J. Kim, E. S. Antonarakis, H. Hammers, M. A. Carducci, Angiotensin system inhibitors and outcome of sunitinib treatment in patients with metastatic renal cell carcinoma: A retrospective examination. *Eur. J. Cancer* **47**, 1955–1961 (2011).
140. R. R. McKay, G. E. Rodriguez, X. Lin, M. D. Kaymakcalan, O.-P. R. Hamnvik, V. S. Sabbiseti, R. S. Bhatt, R. Simantov, T. K. Choueiri, Angiotensin system inhibitors and survival outcomes in patients with metastatic renal cell carcinoma. *Clin. Cancer Res.* **21**, 2471–2479 (2015).
141. A. Melhem-Bertrandt, M. Chavez-MacGregor, X. Lei, E. N. Brown, R. T. Lee, F. Meric-Bernstam, A. K. Sood, S. D. Conzen, G. N. Hortobagyi, A. M. Gonzalez-Angulo, Beta-blocker use is associated with improved relapse-free survival in patients with triple-negative breast cancer. *J. Clin. Oncol.* **29**, 2645–2652 (2011).
142. A. R. Menter, N. M. Carroll, L. C. Sakoda, T. Delate, M. C. Hornbrook, R. K. Jain, L. H. Kushi, V. P. Quinn, D. P. Ritzwoller, Effect of angiotensin system inhibitors on survival in patients receiving chemotherapy for advanced non-small-cell lung cancer. *Clin. Lung Cancer* **18**, 189–197 (2017).
143. L. Miao, W. Chen, L. Zhou, H. Wan, B. Gao, Y. Feng, Impact of angiotensin I-converting enzyme inhibitors and angiotensin II type-1 receptor blockers on survival of patients with NSCLC. *Sci. Rep.* **6**, 21359 (2016).
144. A. Miyajima, S. Yazawa, T. Kosaka, N. Tanaka, S. Shirotake, R. Mizuno, E. Kikuchi, M. Oya, Prognostic impact of renin-angiotensin system blockade on renal cell carcinoma after surgery. *Ann. Surg. Oncol.* **22**, 3751–3759 (2015).
145. M. A. N. Sendur, S. Aksoy, S. Yaman, N. Y. Ozdemir, N. Zengin, K. Altundag, Efficacy of angiotensin-receptor blockers on demographic and clinico-pathological characteristics of breast cancer. *Breast* **21**, 419–420 (2012).
146. M. J. Sorch, G. Kichenadasse, A. Rowland, R. J. Woodman, A. A. Mangoni, Angiotensin system inhibitors and survival in patients with metastatic renal cell carcinoma treated with VEGF-targeted therapy: A pooled secondary analysis of clinical trials. *Int. J. Cancer* **138**, 2293–2299 (2016).
147. N. Tanaka, A. Miyajima, E. Kikuchi, K. Matsumoto, M. Hagiwara, H. Ide, T. Kosaka, T. Masuda, S. Nakamura, M. Oya, Prognostic impact of renin-angiotensin system blockade in localised upper-tract urothelial carcinoma. *Br. J. Cancer* **106**, 290–296 (2012).
148. H. Wang, Z. Liao, Y. Zhuang, Y. Liu, L. B. Levy, T. Xu, S. W. Yusuf, D. R. Gomez, Incidental receipt of cardiac medications and survival outcomes among patients with stage III non-small-cell lung cancer after definitive radiotherapy. *Clin. Lung Cancer* **16**, 128–136 (2015).
149. S. Wilop, S. von Hobe, M. Crysandt, A. Esser, R. Osieka, E. Jost, Impact of angiotensin I converting enzyme inhibitors and angiotensin II type 1 receptor blockers on survival in patients with advanced non-small-cell lung cancer undergoing first-line platinum-based chemotherapy. *J. Cancer Res. Clin. Oncol.* **135**, 1429–1435 (2009).
150. T. Yoshida, H. Kinoshita, K. Fukui, T. Matsuzaki, K. Yoshida, T. Mishima, M. Yanishi, Y. Komai, M. Sugi, T. Inoue, T. Murota, T. Matsuda, Prognostic impact of renin-angiotensin inhibitors in patients with bladder cancer undergoing radical cystectomy. *Ann. Surg. Oncol.* **24**, 823–831 (2017).
151. K. Yuge, A. Miyajima, N. Tanaka, S. Shirotake, T. Kosaka, E. Kikuchi, M. Oya, Prognostic value of renin-angiotensin system blockade in non-muscle-invasive bladder cancer. *Ann. Surg. Oncol.* **19**, 3987–3993 (2012).
152. T. Buchler, M. Krejci, A. Svobodnik, Z. Adam, J. Minarik, J. Bacovsky, V. Scudla, J. Mayer, J. Vorlicek, R. Hajek, Outcome of patients with multiple myeloma and hypertension treated with angiotensin-I-converting enzyme inhibitors during high-dose chemotherapy. *Hematol. J.* **5**, 559–564 (2005).
153. C. R. Cardwell, U. C. Mc Menamin, B. M. Hicks, C. Hughes, M. M. Cantwell, L. J. Murray, Drugs affecting the renin-angiotensin system and survival from cancer: A population based study of breast, colorectal and prostate cancer patient cohorts. *BMC Med.* **12**, 28 (2014).
154. A. F. Carpentier, D. Ferrari, O. Bailon, R. Ursu, C. Banissi, A.-L. Dubessy, C. Belin, C. Levy, Steroid-sparing effects of angiotensin-II inhibitors in glioblastoma patients. *Eur. J. Neurol.* **19**, 1337–1342 (2012).
155. Y. K. Chae, A. Dimou, S. Pierce, H. Kantarjian, M. Andreeff, The effect of calcium channel blockers on the outcome of acute myeloid leukemia. *Leuk. Lymphoma* **55**, 2822–2829 (2014).
156. Y.-H. Chen, C.-H. Huang, H.-I. Lu, C.-H. Chen, W.-T. Huang, M.-J. Hsieh, K.-M. Rau, A. Y. W. Chang, W.-C. Lin, S.-H. Li, Prognostic impact of renin-angiotensin system blockade in esophageal squamous cell carcinoma. *J. Renin Angiotensin Aldosterone Syst.* **16**, 1185–1192 (2015).
157. V. De Giorgi, S. Gandini, M. Grazzini, S. Benemei, N. Marchionni, P. Peppetti, Effect of β -blockers and other antihypertensive drugs on the risk of melanoma recurrence and death. *Mayo Clin. Proc.* **88**, 1196–1203 (2013).
158. D. R. Engineer, B. O. Burney, T. G. Hayes, J. M. Garcia, Exposure to ACEI/ARB and β -blockers is associated with improved survival and decreased tumor progression and hospitalizations in patients with advanced colon cancer. *Transl. Oncol.* **6**, 539–545 (2013).
159. A. Facciorusso, V. Del Prete, N. Crucinio, N. Muscatello, B. I. Carr, A. Di Leo, M. Barone, Angiotensin receptor blockers improve survival outcomes after radiofrequency ablation in hepatocarcinoma patients. *J. Gastroenterol. Hepatol.* **30**, 1643–1650 (2015).
160. L.-R. He, W. Qiao, Z.-X. Liao, R. Komaki, L. Ho, W. L. Hofstetter, S. H. Lin, Impact of comorbidities and use of common medications on cancer and non-cancer specific survival in esophageal carcinoma. *BMC Cancer* **15**, 1095 (2015).
161. J. H. Heinzerling, T. Anthony, E. H. Livingston, S. Huerta, Predictors of distant metastasis and mortality in patients with stage II colorectal cancer. *Am. Surg.* **73**, 230–238 (2007).
162. S. Holmes, E. J. Griffith, G. Musto, G. Y. Minuk, Antihypertensive medications and survival in patients with cancer: A population-based retrospective cohort study. *Cancer Epidemiol.* **37**, 881–885 (2013).
163. E. Januel, R. Ursu, A. Alkhafaji, A. Marantidou, J. Doridam, C. Belin, C. Levy-Piedbois, A. F. Carpentier, Impact of renin-angiotensin system blockade on clinical outcome in glioblastoma. *Eur. J. Neurol.* **22**, 1304–1309 (2015).
164. M. Kaibori, M. Ishizaki, K. Matsui, H. Kitade, Y. Matsui, A.-H. Kwon, Evaluation of metabolic factors on the prognosis of patients undergoing resection of hepatocellular carcinoma. *J. Gastroenterol. Hepatol.* **26**, 536–543 (2011).
165. S. T. Kim, K. H. Park, S. C. Oh, J. H. Seo, J. S. Kim, S. W. Shin, Y. H. Kim, How does inhibition of the renin-angiotensin system affect the prognosis of advanced gastric cancer patients receiving platinum-based chemotherapy? *Oncology* **83**, 354–360 (2012).
166. A. Kourilsky, G. Bertrand, R. Ursu, J. Doridam, C. Barlog, T. Faillot, E. Mandonnet, C. Belin, C. Levy, A. F. Carpentier, Impact of angiotensin-II receptor blockers on vasogenic edema in glioblastoma patients. *J. Neurol.* **263**, 524–530 (2016).
167. Z. S. Morris, S. Saha, W. J. Magnussen, B. A. Morris, J. F. Borkenhagen, A. Ching, G. Hirose, V. McMurry, D. M. Francis, P. M. Harari, R. Chappell, S. Tsujii, M. A. Ritter, Increased tumor response to neoadjuvant therapy among rectal cancer patients taking angiotensin-converting enzyme inhibitors or angiotensin receptor blockers. *Cancer* **122**, 2487–2495 (2016).
168. Y. Nakai, H. Isayama, H. Ijichi, T. Sasaki, N. Sasahira, K. Hirano, H. Kogure, K. Kawakubo, H. Yagioka, Y. Yashima, S. Mizuno, K. Yamamoto, T. Arizumi, O. Togawa, S. Matsubara, T. Tsujino, K. Tateishi, M. Tada, M. Omata, K. Koike, Inhibition of renin-angiotensin system affects prognosis of advanced pancreatic cancer receiving gemcitabine. *Br. J. Cancer* **103**, 1644–1648 (2010).
169. Y. Nakai, H. Isayama, T. Sasaki, S. Mizuno, N. Sasahira, H. Kogure, K. Kawakubo, N. Yamamoto, K. Hirano, H. Ijichi, K. Tateishi, M. Tada, K. Koike, Clinical outcomes of chemotherapy for diabetic and nondiabetic patients with pancreatic cancer: Better prognosis with statin use in diabetic patients. *Pancreas* **42**, 202–208 (2013).
170. Y. Nakai, H. Isayama, T. Sasaki, N. Takahara, K. Saito, K. Ishigaki, T. Hamada, S. Mizuno, K. Miyabayashi, K. Yamamoto, D. Mohri, H. Kogure, N. Yamamoto, H. Ijichi, K. Tateishi, M. Tada, K. Koike, The inhibition of renin-angiotensin system in advanced pancreatic cancer: An exploratory analysis in 349 patients. *J. Cancer Res. Clin. Oncol.* **141**, 933–939 (2015).
171. Y. Nakai, H. Isayama, T. Sasaki, N. Takahara, K. Saito, T. Takeda, G. Umefune, T. Saito, K. Takagi, T. Watanabe, T. Hamada, R. Uchino, S. Mizuno, K. Yamamoto, H. Kogure, S. Matsubara, N. Yamamoto, H. Ijichi, K. Tateishi, M. Tada, K. Koike, No survival benefit from the inhibition of renin-angiotensin system in biliary tract cancer. *Anticancer Res.* **36**, 4965–4970 (2016).

172. H. Osumi, S. Matsusaka, T. Wakatsuki, M. Suenaga, E. Shinozaki, N. Mizunuma, Angiotensin II type-1 receptor blockers enhance the effects of bevacizumab-based chemotherapy in metastatic colorectal cancer patients. *Mol. Clin. Oncol.* **3**, 1295–1300 (2015).
173. M. Pinter, A. Weinmann, M.-A. Wörns, F. Hucke, S. Bota, J. U. Marquardt, D. G. Duda, R. K. Jain, P. R. Galle, M. Trauner, M. Peck-Radosavljevic, W. Sieghart, Use of inhibitors of the renin-angiotensin system is associated with longer survival in patients with hepatocellular carcinoma. *United European Gastroenterol. J.* 10.1177/2050640617695698 (2017).
174. S. J. Tingle, J. A. Moir, S. A. White, Role of anti-stromal polypharmacy in increasing survival after pancreaticoduodenectomy for pancreatic ductal adenocarcinoma. *World J Gastrointest Pathophysiol* **6**, 235–242 (2015).
175. V. A. Levin, J. Chan, M. Datta, J. L. Yee, R. K. Jain, Effect of angiotensin system inhibitors on survival in newly diagnosed glioma patients and recurrent glioblastoma patients receiving chemotherapy and/or bevacizumab. *J. Neurooncol.* **134**, 325–330 (2017).
176. N. Tanaka, A. Miyajima, T. Kosaka, S. Shirotake, M. Hasegawa, E. Kikuchi, M. Oya, Cis-dichlorodiammineplatinum upregulates angiotensin II type 1 receptors through reactive oxygen species generation and enhances VEGF production in bladder cancer. *Mol. Cancer Ther.* **9**, 2982–2992 (2010).
177. N. Tanaka, A. Miyajima, T. Kosaka, Y. Miyazaki, S. Shirotake, H. Shirakawa, E. Kikuchi, M. Oya, Acquired platinum resistance enhances tumour angiogenesis through angiotensin II type 1 receptor in bladder cancer. *Br. J. Cancer* **105**, 1331–1337 (2011).
178. M. M. Abd-Elhaseeb, S. A. Zaitone, S. H. Abou-El-Ela, Y. M. Moustafa, Olmesartan potentiates the anti-angiogenic effect of sorafenib in mice bearing Ehrlich's ascites carcinoma: Role of angiotensin (1–7). *PLOS ONE* **9**, e85891 (2014).
179. M. R. Mancuso, R. Davis, S. M. Norberg, S. O'Brien, B. Sennino, T. Nakahara, V. J. Yao, T. Inai, P. Brooks, B. Freemark, D. R. Shalinsky, D. D. Hu-Lowe, D. M. McDonald, Rapid vascular regrowth in tumors after reversal of VEGF inhibition. *J. Clin. Invest.* **116**, 2610–2621 (2006).
180. A. W. Griffioen, L. A. Mans, A. M. A. de Graaf, P. Nowak-Sliwinska, C. L. M. M. de Hoog, T. A. M. de Jong, F. A. Vyth-Dreese, J. R. van Beijnum, A. Bex, E. Jonasch, Rapid angiogenesis onset after discontinuation of sunitinib treatment of renal cell carcinoma patients. *Clin. Cancer Res.* **18**, 3961–3971 (2012).
181. W. D. Fox, B. Higgins, K. M. Maiese, M. Drobnjak, C. Cordon-Cardo, H. I. Scher, D. B. Agus, Antibody to vascular endothelial growth factor slows growth of an androgen-independent xenograft model of prostate cancer. *Clin. Cancer Res.* **8**, 3226–3231 (2002).
182. T. Powles, C. Blank, S. Chowdhury, S. Horenblas, J. Peters, J. Shamash, N. Sarwar, E. Boleti, A. Sahdev, T. O'Brien, D. Berney, L. Beltran, P. Nathan, J. Haanen, A. Bex, The outcome of patients treated with sunitinib prior to planned nephrectomy in metastatic clear cell renal cancer. *Eur. Urol.* **60**, 448–454 (2011).
183. B. I. Rini, J. Bellmunt, J. Clancy, K. Wang, A. G. Niethammer, S. Hariharan, B. Escudier, Randomized phase III trial of temsirolimus and bevacizumab versus interferon alfa and bevacizumab in metastatic renal cell carcinoma: INTORACT trial. *J. Clin. Oncol.* **32**, 752–759 (2014).
184. N. B. Haas, J. Manola, R. G. Uzzo, K. T. Flaherty, C. G. Wood, C. Kane, M. Jewett, J. P. Dutcher, M. B. Atkins, M. Pins, G. Wilding, D. Cella, L. Wagner, S. Matin, T. M. Kuzel, W. J. Sexton, Y.-N. Wong, T. K. Choueiri, R. Pili, I. Puzanov, M. Kohli, W. Stadler, M. Carducci, R. Coomes, R. S. DiPaola, Adjuvant sunitinib or sorafenib for high-risk, non-metastatic renal-cell carcinoma (ECOG-ACRIN E2805): A double-blind, placebo-controlled, randomised, phase 3 trial. *Lancet* **387**, 2008–2016 (2016).
185. A. Ravaud, M. Schmidinger, Clinical biomarkers of response in advanced renal cell carcinoma. *Ann. Oncol.* **24**, 2935–2942 (2013).
186. S. Raimondi, E. Botteri, E. Munzone, C. Cipolla, N. Rotmensz, A. DeCensi, S. Gandini, Use of beta-blockers, angiotensin-converting enzyme inhibitors and angiotensin receptor blockers and breast cancer survival: Systematic review and meta-analysis. *Int. J. Cancer* **139**, 212–219 (2016).
187. European Association For The Study Of The Liver; European Organisation For Research And Treatment Of Cancer, EASL-EORTC clinical practice guidelines: Management of hepatocellular carcinoma. *J. Hepatol.* **56**, 908–943 (2012).
188. M. G. Fakih, Metastatic colorectal cancer: Current state and future directions. *J. Clin. Oncol.* **33**, 1809–1824 (2015).
189. B. Escudier, C. Porta, M. Schmidinger, N. Rioux-Leclercq, A. Bex, V. Khoo, V. Gruenvald, A. Horwich; ESMO Guidelines Committee, Renal cell carcinoma: ESMO clinical practice guidelines for diagnosis, treatment and follow-up. *Ann. Oncol.* **27** (suppl. 5), v58–v68 (2016).
190. J. E. Faris, L. S. Blaszkowsky, S. McDermott, A. R. Guimaraes, J. Szymonifka, M. A. Huynh, C. R. Ferrone, J. A. Wargo, J. N. Allen, L. E. Dias, E. L. Kwak, K. D. Lillemo, S. P. Thayer, J. E. Murphy, A. X. Zhu, D. V. Sahani, J. Y. Wo, J. W. Clark, C. Fernandez-del Castillo, D. P. Ryan, T. S. Hong, FOLFIRINOX in locally advanced pancreatic cancer: The Massachusetts General Hospital Cancer Center experience. *Oncologist* **18**, 979–985 (2015).
191. E. A. Mellon, S. E. Hoffe, G. M. Springett, J. M. Frakes, T. J. Strom, P. J. Hodul, M. P. Malafa, M. D. Chuong, R. Shridhar, Long-term outcomes of induction chemotherapy and neoadjuvant stereotactic body radiotherapy for borderline resectable and locally advanced pancreatic adenocarcinoma. *Acta Oncol.* **54**, 979–985 (2015).
192. T. Song, C. H. Choi, M. K. Kim, M.-L. Kim, B. S. Yun, S. J. Seong, The effect of angiotensin system inhibitors (angiotensin-converting enzyme inhibitors or angiotensin receptor blockers) on cancer recurrence and survival: A meta-analysis. *Eur. J. Cancer Prev.* **26**, 78–85 (2017).
193. M. Pinter, M. Trauner, M. Peck-Radosavljevic, W. Sieghart, Cancer and liver cirrhosis: Implications on prognosis and management. *ESMO Open* **1**, e000042 (2016).
194. H. Yoshiji, S. Kuriyama, J. Yoshii, Y. Ikenaka, R. Noguchi, T. Nakatani, H. Tsujinoue, H. Fukui, Angiotensin-II type 1 receptor interaction is a major regulator for liver fibrosis development in rats. *Hepatology* **34**, 745–750 (2001).
195. J. R. Jonsson, A. D. Clouston, Y. Ando, L. I. Kelemen, M. J. Horn, M. D. Adamson, D. M. Purdie, E. E. Powell, Angiotensin-converting enzyme inhibition attenuates the progression of rat hepatic fibrosis. *Gastroenterology* **121**, 148–155 (2001).
196. P. Tandon, J. G. Abraldes, A. Berzigotti, J. C. Garcia-Pagan, J. Bosch, Renin-angiotensin-aldosterone inhibitors in the reduction of portal pressure: A systematic review and meta-analysis. *J. Hepatol.* **53**, 273–282 (2010).
197. R. K. Jain, Lessons from multidisciplinary translational trials on anti-angiogenic therapy of cancer. *Nat. Rev. Cancer* **8**, 309–316 (2008).

Acknowledgments: We thank Y. Boucher, I. Chen, A. Crane, M. Datta, M. Khandekar, H. Liu, M. Pittet, and K. Naxerova for helpful comments. We also apologize to all authors whose papers are not cited because of the limitations in number of references. **Funding:** M.P. is supported by an Erwin-Schrodinger Fellowship by the Austrian Science Fund (project no. J 3747-B28). R.K.J. is supported by the National Cancer Institute (NCI; grants P01-CA080124, P50-CA165962, R01-CA129371, R01-CA208205, and U01-CA 224348), NCI Outstanding Investigator Award (R35-CA197743), the Lustgarten Foundation, the Ludwig Center at Harvard, the National Foundation for Cancer Research, and the Gates Foundation. **Author contributions:** M.P. and R.K.J. performed the literature search, wrote the manuscript, and approved the final version of the manuscript. **Competing interests:** M.P. received travel support from Bayer and speaking fees and consultant fees from Bayer and Bristol-Myers Squibb. R.K.J. received consultant fees from Ophthotech, Sun Pharma Advanced Research Corporation (SPARC), SynDevRx, and XTuit; owns equity in Enlight, Ophthotech, SynDevRx, and XTuit; and serves on the Board of Directors of XTuit and the Boards of Trustees of Tekla Healthcare Investors, Tekla Life Sciences Investors, Tekla Healthcare Opportunities Fund, and Tekla World Healthcare Fund. R.K.J. is an inventor on patent application (USSN 61/43 8,240 and USSN 61/643,487) submitted by MGH that covers the use of antihypertensive agents for cancer therapy.

Submitted 1 May 2017
Accepted 25 August 2017
Published 4 October 2017
10.1126/scitranslmed.aan5616

Citation: M. Pinter, R. K. Jain, Targeting the renin-angiotensin system to improve cancer treatment: Implications for immunotherapy. *Sci. Transl. Med.* **9**, ean5616 (2017).

Science Translational Medicine

Targeting the renin-angiotensin system to improve cancer treatment: Implications for immunotherapy

Matthias Pinter and Rakesh K. Jain

Sci Transl Med **9**, eaan5616.
DOI: 10.1126/scitranslmed.aan5616

ARTICLE TOOLS	http://stm.sciencemag.org/content/9/410/eaan5616
SUPPLEMENTARY MATERIALS	http://stm.sciencemag.org/content/suppl/2017/10/02/9.410.eaan5616.DC1
RELATED CONTENT	http://stm.sciencemag.org/content/scitransmed/9/385/eaak9679.full http://stm.sciencemag.org/content/scitransmed/9/391/eaal4682.full http://stm.sciencemag.org/content/scitransmed/9/385/eaak9670.full http://stm.sciencemag.org/content/scitransmed/8/340/340ra73.full http://stm.sciencemag.org/content/scitransmed/10/442/eaah4807.full http://science.sciencemag.org/content/sci/365/6453/544.full
REFERENCES	This article cites 194 articles, 43 of which you can access for free http://stm.sciencemag.org/content/9/410/eaan5616#BIBL
PERMISSIONS	http://www.sciencemag.org/help/reprints-and-permissions

Use of this article is subject to the [Terms of Service](#)

Science Translational Medicine (ISSN 1946-6242) is published by the American Association for the Advancement of Science, 1200 New York Avenue NW, Washington, DC 20005. The title *Science Translational Medicine* is a registered trademark of AAAS.

Copyright © 2017 The Authors, some rights reserved; exclusive licensee American Association for the Advancement of Science. No claim to original U.S. Government Works