

PD-L1 (B7-H1) and PD-1 pathway blockade for cancer therapy: Mechanisms, response biomarkers, and combinations

Weiping Zou,^{1*} Jedd D. Wolchok,^{2*} Lieping Chen^{3*}

PD-L1 and PD-1 (PD) pathway blockade is a highly promising therapy and has elicited durable antitumor responses and long-term remissions in a subset of patients with a broad spectrum of cancers. How to improve, widen, and predict the clinical response to anti-PD therapy is a central theme in the field of cancer immunology and immunotherapy. Oncologic, immunologic, genetic, and biological studies focused on the human cancer microenvironment have yielded substantial insight into this issue. Here, we focus on tumor microenvironment and evaluate several potential therapeutic response markers including the PD-L1 and PD-1 expression pattern, genetic mutations within cancer cells and neoantigens, cancer epigenetics and effector T cell landscape, and microbiota. We further clarify the mechanisms of action of these markers and their roles in shaping, being shaped, and/or predicting therapeutic responses. We also discuss a variety of combinations with PD pathway blockade and their scientific rationales for cancer treatment.

INTRODUCTION

The tumor microenvironment is the primary location in which tumor cells and the host immune system interact. Characterization of the nature of immune responses in the human cancer microenvironment holds the key to understanding protective tumor immunity and improving and empowering current cancer immunotherapy. Accumulating evidence has revealed that the interaction between tumor cells and the host immune system fosters tumor immune evasion and ultimately results in tumor dissemination, relapse, and metastasis (1–3). The study of different cancer-infiltrating immune cell subsets, including CD4⁺Foxp3⁺ regulatory T cells (T_{regs}) (4), antigen-presenting cells (APCs) (5, 6), myeloid-derived suppressor cells (MDSCs) (7), and effector T cell subsets (8–13), and immune signature networks (3) has defined the nature of immune responses in the human cancer microenvironment. These findings have elucidated the critical importance of reversing immunosuppressive mechanisms, including programmed cell death 1 ligand (PD-L1, B7-H1, CD274) and programmed cell death receptor 1 (PD-1, CD279) pathway (herein, PD pathway) blockade (1, 2, 14, 15), to engender potent antitumor immunity. The identification of PD-L1 (16, 17), the finding that PD-1 is a receptor for PD-L1 (18), and the demonstration of the expression, regulation, and function of the PD pathway in the human cancer microenvironment (5, 14, 16, 17, 19–23) have provided scientific rationales and direct support for the current clinical application of PD pathway blockade (Table 1).

B7-H1 was cloned in 1999 (16). PD-1 has been subsequently identified as a counter-receptor for B7-H1 (18), and B7-H1 is therefore also known as PD-L1 to emphasize this receptor-ligand interaction. The expression profile of PD-L1 in human cancers has been previously reviewed (14). In addition to tumor cells (14, 17), high levels of PD-L1 protein expression have been observed in human tumor-associated APCs including tumor environmental dendritic cells (DCs), tumor-

draining lymph node DCs (5, 24), macrophages (20, 25), fibroblasts (26), and T cells (27, 28). PD-L1 expression can be induced or maintained by many cytokines (5, 17, 29, 30), of which interferon- γ (IFN- γ) is the most potent. The association between tumor-infiltrating T cells (TILs), IFN- γ signaling genes, and PD-L1 expression suggests that effector T cell-derived IFN- γ contributes to high levels of PD-L1 expression in the tumor microenvironment (31). Immune-induced tumor PD-L1 expression is considered to be an adaptive resistance mechanism for tumor cells in response to immune challenge (31–33). In addition, oncogenic phosphatase and tensin homolog (PTEN) loss results in enhanced PD-L1 expression in glioma (34) and triple-negative breast cancer cells (35). In human T cell lymphoma, PD-L1 expression may depend on the expression and enzymatic activity of chimeric nucleophosmin (NPM) and anaplastic lymphoma kinase (ALK) (36). Thus, PD-L1 expression can also be regulated by intrinsic oncogenic pathways.

In addition to PD-L1, PD-L2 (B7-DC, CD273) also interacts with PD-1 with similar affinity to deliver a potentially suppressive signal. The immunologic function of this interaction in cancer immunity, however, may not be critical because of the relatively rare expression of PD-L2 on cancer cells and its interaction with a potentially stimulatory receptor, repulsive guidance molecule b (RGMB) (Fig. 1) (15).

PD-1 is absent in resting T cells and was initially found in activated mouse T cells upon T cell receptor (TCR) engagement (37) and subsequently in exhausted T cells in chronic infection models (38, 39). In patients with different types of cancer, high levels of PD-1 expression are detected in TILs including tumor antigen-specific T cells, presumably due to chronic antigenic stimulation. These human tumor-associated PD-1⁺ T cells are functionally impaired, and their biological activity can be partially recovered with PD-1 or PD-L1 blockade (19–23). A recent study reports that a subset of human melanoma cells express PD-1 and melanoma cell-intrinsic PD-1 promotes melanoma cell growth (40). This finding, however, is inconsistent with previous reports that PD-1 expression is predominant on tumor-infiltrating lymphocytes but poorly expressed in melanoma based on immunohistochemistry analysis (41). Indeed, differentiating the role of specific reagents and techniques in detecting PD-1 expression by tumor cells is important to further understand the biologic importance of the data.

¹Department of Surgery, University of Michigan School of Medicine, Ann Arbor, MI 48109, USA. ²Department of Medicine and the Ludwig Center, Memorial Sloan Kettering Cancer Center, New York, NY 10065, USA. ³Department of Immunobiology, Yale University School of Medicine, New Haven CT 06519, USA.

*Corresponding author. E-mail: wzou@med.umich.edu (W.Z.); wolchokj@mskcc.org (J.D.W.); lieping.chen@yale.edu (L.C.)

Table 1. Examples of clinical trials with PD-1 and PD-L1 blockade. Clinical trials with less than 20 cases or published after 1 October 2015 were not included in the table. CRC, colorectal cancer; HSCT, hematopoietic stem cell transplantation.

Target and drug information	Clinical response rate in different cancer types	Phase	Cases	References
Target: PD-1 Name: Nivolumab, Opdivo, BMS-936558, MDX-1106, ONO-4538 Isotype: Humanized IgG4a Source: Bristol-Myers Squibb, Ono Pharmaceuticals	12.8% in treatment-refractory metastatic melanoma, castrate-resistant prostate cancer, RCC, NSCLC, or CRC	1	39	(50)
	28% in advanced melanoma, 18% in NSCLC, 27% in RCC	1	296	(51)
	40% in melanoma treated with nivolumab + ipilimumab, 20% in nivolumab followed by ipilimumab	1	86	(58)
	87% in relapsed or refractory Hodgkin's lymphoma	1	23	(56)
	14.5% in refractory NSCLC	2	117	(70)
	31.7% in advanced melanoma progressed after anti-CTLA-4	3	405	(65)
	40% in previously untreated melanoma without BRAF mutation	3	418	(64)
	17% in previously treated advanced NSCLC	2	129	(69)
	29% in previously treated advanced RCC	1	34	(71)
	20% in advanced squamous-cell NSCLC	3	272	(68)
	57.6% (nivolumab + ipilimumab) versus 19% (ipilimumab) versus 43.7% (nivolumab) in untreated stage III or IV melanoma	3	945	(63)
Target: PD-1 Name: Pembrolizumab, Keytruda, MK-3475, lambrolizumab Isotype: Humanized IgG4κ Source: Merck	38% in melanoma	1	135	(57)
	26% in ipilimumab-refractory advanced melanoma	1	173	(61)
	63% versus 0% in stage IV NSCLC patients with high and low nonsynonymous mutation burden	1	29	(67)
	19.4% in advanced NSCLC	1	495	(66)
	40% and 0% in mismatch repair-deficient/proficient CRC	2	41	(53)
	33% (pembrolizumab) and 11.9% (ipilimumab) in advanced melanoma	3	834	(62)
Target: PD-1 Name: Pidilizumab or CT-011 Isotype: Humanized IgG1 Source: CureTech Ltd.	51% in diffuse large B cell lymphoma (after HSCT)	2	66	(54)
	66% in relapsed follicular lymphoma	2	32	(55)
Target: PD-L1 Name: MPDL3280A, RG7446 Isotype: Fc-modified human IgG1b Source: Genentech/Roche	21% overall response rate in advanced incurable cancer NSCLC, SCLC, melanoma, RCC, CRC, gastric cancer, head and neck squamous cell carcinoma, breast cancer, ovarian, pancreatic cancer, uterine cancer, sarcoma, pancreaticoduodenal cancer	1	277	(49)
	52% in metastatic bladder cancer	1	68	(47)
Target: PD-L1 Name: BMS-936559, MDX-1105 Isotype: Fully human IgG4a Source: Bristol-Myers Squibb	17.3% in melanoma, 11.7% in RCC, 10.2% in NSCLC, 5.9% in ovarian cancer	1	207	(48)

The magnitude of this surprising finding remains to be prospectively determined in additional studies across tumor types in patients. Thus, PD-L1 and PD-1 are expressed by various cellular components in the human tumor microenvironment, where they can inhibit antitumor T cell immunity (Fig. 1). This geographic expression profile is an important feature for PD pathway blockade.

Here, we focus on the human cancer microenvironment and PD pathway blockade. We propose that human antitumor immune responses are controlled and regulated by tumor somatic mutations, epigenetic alterations, and environmental cues. We discuss these three

aspects, emphasize the relevant studies in the human cancer microenvironment, and link scientific rationales to clinical combinational therapy with PD pathway blockade.

MECHANISMS OF ACTION OF THE PD SIGNALING PATHWAY

How does PD signaling mediate dysfunctional tumor immunity? PD-L1⁺ cells, particularly PD-L1-expressing APCs and tumor cells, engage

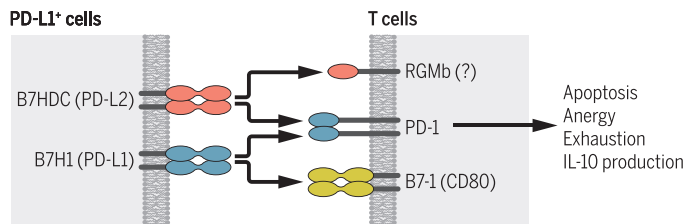


Fig. 1. Mechanisms of action of the PD-L1 and PD-1 pathway. Cells that express high levels of PD-L1 may include tumor cells, APCs (DCs, macrophages, MDSCs, and B cells), T lymphocytes, epithelial cells, fibroblasts, and others. Engagement of PD-L1 by PD-L1⁺ cells induces T cell apoptosis, anergy, functional exhaustion, or IL-10 production.

PD-1⁺ T cells, resulting in T cell dysfunction. Multiple modes of action are thought to explain T cell immune evasion through the PD pathway. The engagement of PD-L1 and PD-1 may cause T cell apoptosis, anergy, exhaustion, and interleukin-10 (IL-10) expression (Fig. 1). PD-L1 may function as a molecular “shield” to protect PD-L1⁺ tumor cells from CD8⁺ T cell-mediated lysis (14, 15). In addition to PD-1, an interaction between PD-L1 and CD80 has been demonstrated in mouse models (Fig. 1) (42, 43). Activated T cells and APCs may express CD80, which may function as a receptor and deliver inhibitory signals when engaged by PD-L1 (42, 43). Moreover, it has been shown that PD-L1 can function as a receptor to “back” transmit signals into T cells (44) and tumor cells (45) to affect their survival, whereas the intracellular biochemistry of this back signaling is yet to be determined. Thus, PD-L1 could act as both ligand and receptor to execute immunoregulatory functions.

In addition to PD-L1, PD-1 is a receptor for PD-L2. RGMb is a binding partner for PD-L2 (Fig. 1) (46). Thus, PD blockade may not be biologically identical, and differing blockade may shift the balance in their interaction with their binding partners, leading to potential varied biological outcomes (Fig. 1). Notably, the relationship between CD80, PD-1 and RGMb, and PD-L1 and PD-L2 in their cellular expression profile, expression regulation, potential molecular interaction, and functional relevance in human tumor immunity and checkpoint immunotherapy has not been completely defined. Furthermore, because PD-L1 is expressed by different types of cells and mediates immunoregulation through different mechanisms, it is not totally understood which major cellular and molecular mechanisms are correlated with clinical responses to PD pathway blockade in patients with cancer. Nonetheless, current clinical trials demonstrate similar clinical response patterns for anti-PD therapy (Table 1), suggesting that major cellular and molecular mechanisms associated with a clinical response may be shared in PD blockade. Future studies of the tumor microenvironment in patients with or without immunotherapy will hopefully demonstrate the dynamics of these various interactions and the underlying cellular and molecular mechanisms, which are relevant for further understanding of how immune elements shape and predict therapeutic response and nonresponse (Fig. 1).

CLINICAL TRIALS AND RESPONSE BIOMARKERS OF PD PATHWAY BLOCKADE

Clinically, PD pathway blockade has demonstrated important activity across a spectrum of different tumor types spanning both solid tumors

and hematologic malignancies including bladder cancer (47), breast cancer (48, 49), colorectal cancer (48, 50–53), diffuse large B cell lymphoma (54), follicular lymphoma (55), gastric cancer (48), head and neck squamous cell carcinoma (49), Hodgkin’s lymphoma (56), melanoma (48–51, 57–65), ovarian cancer (48, 49), non-small cell lung cancer (NSCLC) (7, 48–51, 66, 68–70), pancreatic cancer (48, 49), renal cell carcinoma (RCC) (48–51, 71), prostate cancer (50, 51), sarcoma (49), small cell lung cancer (SCLC) (49), and uterine cancer (49). The objective response rates are varied from different cancer types in different clinical trials (Table 1). On the basis of current clinical data, bladder cancer (47), melanoma (48–52, 57–65), mismatch repair-deficient colorectal cancer (53), and certain hematopoietic malignancies (54, 56) may be among the most responsive cancer types. Specific features of particular clinical trials are summarized (Table 1) and discussed in the following sections.

Although head-to-head comparisons of antibodies to PD-1 and PD-L1 have not been done, levels of clinical response and toxicities appear to be generally consistent with either approach. Immune-related toxicities occur with PD pathway blockade but are much less frequent than those observed with cytotoxic T lymphocyte antigen-4 (CTLA-4) blockade (62, 63). The most frequently observed toxicity encountered with PD pathway blockade is fatigue, which often does not require treatment and does not necessarily limit the duration of therapy. However, inflammatory pneumonitis has been observed, which may be fatal if not addressed promptly with corticosteroids or other means of immunosuppression. Rare, high-grade events like pneumonitis or interstitial nephritis may necessitate cessation of therapy, yet clinical responses remain durable despite cessation of therapy and even after immunosuppression, implying that the optimal duration of therapy with PD pathway blocking agents remains to be determined.

Given that PD pathway blockade induces a clinical response in subsets of cancer patients (Table 1), a central question is whether and how therapeutic responsiveness is predicted and/or shaped by host and tumor components. There are many ongoing efforts to identify predictive biomarkers of PD pathway blockade. Recent studies have provided hints to associate clinical responses with several potential biomarkers (Fig. 2).

Does the expression level of PD-L1 predict clinical response?

Tumor cells and APCs express high levels of PD-L1, and tumor-associated PD-L1⁺ DCs mediate T cell suppression (5, 14). Tumor tissues of RCC (72) and esophageal, gastric (73, 74), and ovarian (75) cancers show that PD-L1 expression is an indicator of poor prognosis for patient survival. It is reasonable to hypothesize that PD-L1 in tumor and/or APCs in the tumor microenvironment may predict or be associated with the clinical response of PD blockade. In support of this, a correlation has been observed between the expression of tumor tissue PD-L1 and the likelihood of the response to anti-PD therapy in patients with melanoma (48, 51), NSCLC (66), and RCC (41). An 87% objective response is observed in patients with relapsed or refractory Hodgkin’s lymphoma treated with anti-PD-1 (Table 1). In line with this, an amplification of PD-L1 and PD-L2 is detected in lymphoma cells in these patients (56). In addition to PD-L1 expression in tumor, expression of PD-L1 in tumor-infiltrating immune cells, particularly myeloid APCs (macrophages and myeloid DCs), is correlated with clinical responses to anti-PD-L1 treatment in several types of cancer (Table 1) (49). In contrast, most progressing patients show a lack of PD-L1 up-regulation by either tumor cells or tumor-infiltrating immune cells (49). However, the results

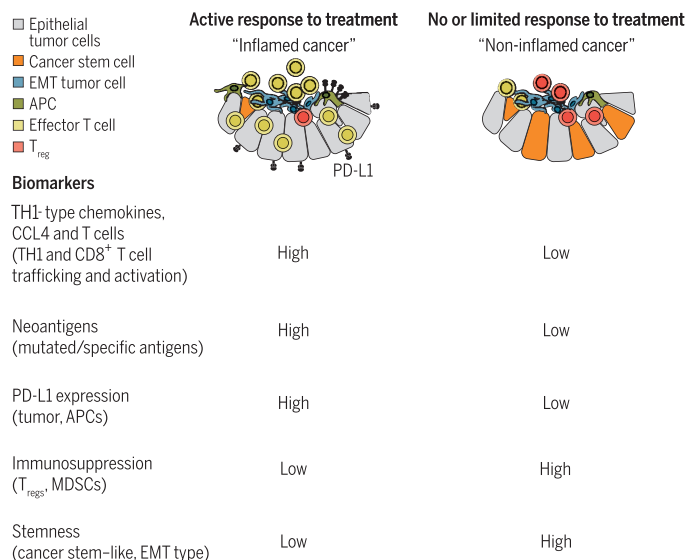


Fig. 2. Proposed potential response biomarkers of PD pathway blockade. Several biomarkers, including high levels of PD-L1 expression, T_H1-type chemokines, infiltrating T cells, mutations, low levels of immunosuppressive elements, and EMT/stem-like features, may be associated with an active response to PD pathway blockade.

in trials with anti-PD-1 (64) and the combination of anti-PD-L1 and anti-CTLA-4 suggest that melanoma patients can have a clinical response regardless of the tumor cell PD-L1 status (58, 76). Notably, the host immune cell PD-L1 expression, particularly PD-L1 expression in DCs in the tumor microenvironment and draining lymph nodes (5), has not been specifically examined in most clinical trials. Furthermore, activated T cells or innate immune cells can release type I and II IFN and stimulate de novo PD-L1 expression. Presumably, blocking newly induced PD-L1 can affect therapeutic responses. In support of this possibility, across multiple cancer types, responses to anti-PD-L1 therapy are frequent in patients with high PD-L1 expression in tumor-infiltrating immune cells, particularly macrophages and DCs, in the course of tumor regression (41, 47, 49). Hence, the relevance of host PD-L1 expression in shaping the clinical response to PD pathway blockade is important to consider.

Furthermore, PD-L1 expression may be clustered rather than diffuse in tumor tissues (17, 31) and is likely localized to the area where IFN- γ ⁺ T cells infiltrate (31). Thus, current needle biopsy-based sampling of human tumor tissues may miss the PD-L1-positive area and give false-negative results. This problem may be overcome by an in vivo imaging method using radiolabeled high-affinity PD-1 variants to assess PD-L1 expression in the entire tumor as shown in a tumor-bearing mouse model (77). In addition, oncogene-driven expression of tumor PD-L1 may not correlate with TILs. It remains to be determined whether the therapeutic efficacy of PD pathway blockade is similar between patients with oncogenic PD-L1⁺ tumors (31, 41, 78) and immune-driven PD-L1⁺ tumors.

With current limitations of clinical sampling methods, the expression of PD-L1 on the surface of tumor cells and immune cells before immunotherapy may be a useful but not a definitive predictive biomarker of the response to the PD pathway blockade. Unlike the presence of oncogenic driver mutations, PD-L1 expression is a dynamic and

inducible biomarker, which is more of a relative indicator of the likelihood of response rather than a binary predictor of response.

Do cancer neoantigens and/or somatic mutations predict a clinical response?

The immune system can recognize developing cancers. Therapeutic manipulation of immunity can induce tumor regression. Whereas tumor-associated antigens (TAAs) are largely self-antigens, cancers contain somatic genetic mutations, which could be specific to the cancer. These mutation-associated antigens can be tumor-specific antigens and be presented to and recognized by T cells in patients with cancer. Aligned with this notion, the tumor mutational antigen (neoantigen) T cell response has been documented in a genetically engineered, autochthonous mouse model of sarcomagenesis (79), in a highly immunogenic methylcholanthrene (MCA)-induced mouse sarcoma model (80), and in mouse vaccine models (2, 81). In patients with melanoma, tumor mutation-specific CD4⁺ (83, 84) and CD8⁺ (85) T cells are found, and these cells can mediate tumor regression. Vaccination with melanoma-derived mutant peptides augments T cell immunity directed at naturally occurring dominant neoantigens and subdominant neoantigens (86). Although the antigen specificity is unknown, the mismatch repair-defective subset of colorectal cancer displays high tumor infiltration of T helper 1 (T_H1)-type T cells and CD8⁺ T cells along with high PD-L1 and PD-1 expression (87). With large-scale genomic data sets of solid tumor tissues, the number of predicted major histocompatibility complex (MHC) class I-associated neoantigens is correlated with the cytolytic activity of CD8⁺ T cells (88).

Several lines of evidence support a link between PD pathway blockade and tumor mutation-derived antigen-specific T cell responses. In the mouse MCA sarcoma model, treatment with anti-PD-1 and/or anti-CTLA-4 activates tumor mutational antigen-specific T cells and results in tumor rejection (89). In patients with colorectal cancers, mismatch repair status predicts clinical benefit of PD-1 blockade (53). In patients with NSCLC treated with anti-PD-1, high nonsynonymous mutation burden is associated with improved objective response, durable clinical benefit, and progression-free survival, and mutated antigen-specific CD8⁺ T cell responses parallel tumor regression (67). Lung cancer and melanoma are found to be clinically responsive in two early clinical trials with anti-PD therapy (51). These two types of cancer have high numbers of somatic mutations as a result of exposure to cigarette smoke and ultraviolet radiation. Biopsied specimens of regressing melanoma lesions are infiltrated by CD8⁺ T cells in patients treated with anti-PD therapy (57). A high objective response rate to anti-PD-1 therapy is observed in patients with microsatellite instable colorectal cancer but not in patients with mismatch repair-proficient colorectal cancer (53). Similarly, in 29 stage IV NSCLC patients, there were 63% and 0% response rates, respectively, with high and low nonsynonymous mutation burden (Table 1) (67). The data suggest that the increased number of mutation-associated neoantigens may be associated with the enhanced responsiveness to PD pathway blockade in some cancer patients.

Notably, we are at the beginning of our understanding of the relationship between mutant tumor neoantigens, their T cell responses, and cancer immunotherapy responses. Several observations are worth noting in this regard: (i) Melanoma and lung cancer naturally possess high levels of mutations. (ii) High levels of mutations may not necessarily form high levels of immunogenic neoantigens; this may simply be a probabilistic phenomenon. (iii) There is no direct evidence demonstrating that neoantigen-specific T cells mediate tumor elimination in

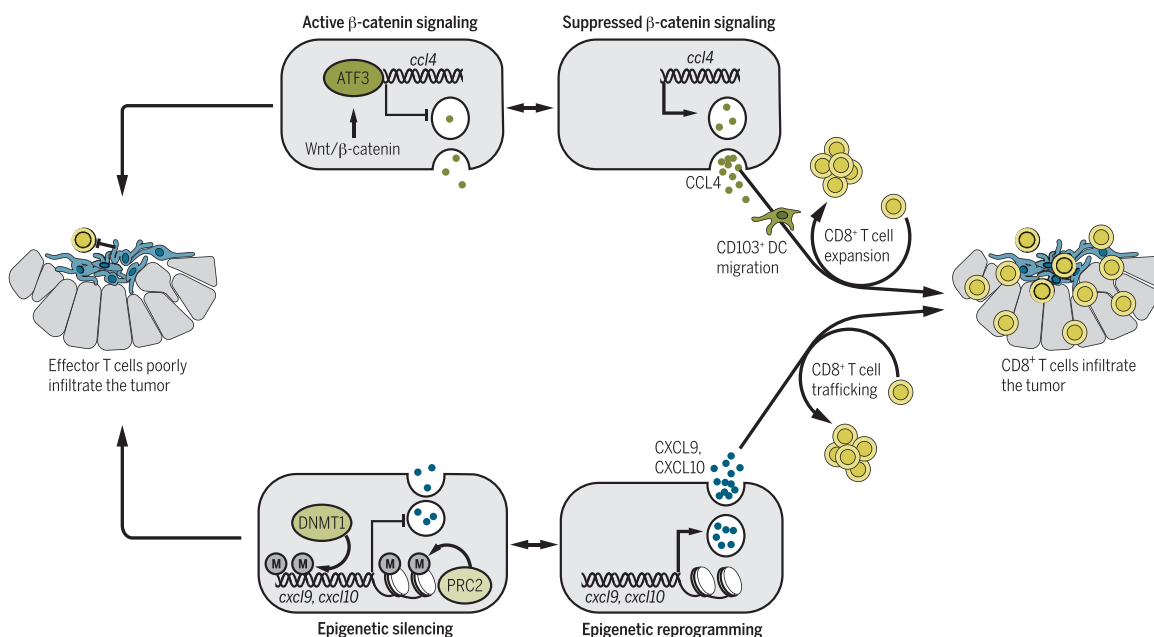
patients treated with PD pathway blockade. (iv) Multiple factors including PD-L1 and PD-1 expression levels, effector T cell tumor trafficking, and environmental cues (microbiota) may contribute to immunotherapeutic responses. Nonetheless, recognizing the intrinsic ability of the immune system to specifically target the unique mutations potentially shared by many cancer types (90) is an important part of understanding the mechanism of cancer immunotherapy (91).

Do T_H1-type chemokines, TILs, and T cell clonality predict clinical responses (Fig. 3)?

T_H1-type chemokines are correlated with effector T cell density in some human tumors and are also positively associated with cancer patient survival (8, 10, 92). However, a crucial question is why some tumors are “inflamed” with effector T cell infiltration, whereas others are not. Two research teams have proposed plausible mechanisms to address this question. In a murine melanoma model, tumor-intrinsic β-catenin signaling negatively controls chemokine CCL4 expression. CCL4 mediates DC tumor trafficking. Thus, tumor β-catenin activation results in poor CCL4 expression and subsequently limits DC recruitment and DC-mediated T cell activation (Fig. 3) (93). In human ovarian cancer (94) and colon cancer (95), poor T cell tumor infiltration is attributed to potent epigenetic silencing of the tumor T_H1-type chemokines CXCL9 and CXCL10, which mediate effector T cell and natural killer (NK) cell tumor migration (Fig. 3) (94, 95). Polycomb repressive complex 2 (PRC2), the demethylase JMJD3-mediated histone H3 lysine 27 trimethylation (H3K27me3), and DNA methylation repress the expression of T_H1-type chemokines and subsequently restrain effector T cell trafficking into the cancer microenvironment (94, 95). These studies suggest that intrinsic tumor oncogenic (93) and epigenetic (94, 95) pathways control T cell activation and/or migration. Both epigenetic silencing (96) and β-catenin signaling are intrinsic tumorigenic mechanisms and are associated with cancer stem cell properties. Thus, oncogenic genetic and epigenetic pathways may play dual biologic and immunologic roles in supporting tumor progression and limiting spontaneous and therapeutic-induced tumor-specific T cell immunity (Fig. 3).

Fig. 3. Mechanisms of poor tumor T cell infiltration.

Active tumor β-catenin inhibits CCL4 expression and limits CD103⁺ DC recruitment and CD8⁺ T cell activation. T_H1-type chemokines CXCL9 and CXCL10 are repressed by EZH2 and DNMT-mediated epigenetic silencing. Consequently, CD8⁺ T cells poorly infiltrate tumor. Therefore, suppressing β-catenin or epigenetic reprogramming could increase CD8⁺ T cell tumor infiltration.



PD-L1 expression is clearly important in the tumor microenvironment (5, 14, 15, 33). A major feature of PD pathway blockade is immunoregulation specifically at the tumor site (15). Hence, it is reasonable to assume that preexisting tumor-infiltrating immune cells and T_H1-type chemokines may correlate with clinical response to PD pathway blockade. In support of this possibility, recent clinical trials suggest that intratumoral T cell infiltration and T_H1-type gene expression and a clonal TCR repertoire are associated with improved clinical responses to anti-PD therapy (47, 97). The frequency of mutational antigen-specific T cells and their functional status in the TIL populations remain to be investigated and compared before and after cancer immunotherapy. Given that oncogenic genetic (93) and epigenetic (94, 95) pathways control effector T cell tumor trafficking, epigenetic marks, enhancer of zeste homolog 2 (EZH2), and DNA methyltransferase 1 (DNMT1) and are negatively associated with CD8⁺ T cells and patient survival (94, 95), and epigenetic reprogramming synergistically increases the effect of PD-L1 blockade (94), it will be interesting to explore whether tumor-specific genetic and epigenetic marks are associated with the clinical response to anti-PD therapy (Fig. 2).

Do microbiota contribute to clinical responses to PD pathway blockade?

Recent studies have shown that the gut microbiome can affect the outcome of cancer chemotherapy in murine models (98, 99). Chemotherapy-induced T_H1 and T_H17 responses are enhanced by translocating commensals and contribute to tumor eradication (98, 99). This is in line with the antitumor role of polyfunctional T_H17 cells in human ovarian cancer (11, 92). Similarly, commensal bacteria can alter the therapeutic effect of anti-PD-L1 therapy in mice bearing subcutaneous tumors, and the response to PD-L1 blockade is enhanced by supplementation with “good” bacteria, *Bifidobacterium*, during treatment (100). The antitumor effects of anti-CTLA-4 also depend on distinct *Bacteroides* species. In mice and patients, T cell responses specific for *B. thetaiotaomicron* or *B. fragilis* are associated with the efficacy of the CTLA-4 blockade (101). It appears that immune responses modulated by the gut microbiome

can have systemic effects on tumor immunity and cancer therapy. It remains to be defined whether the gut microbiome of cancer patients will have an important impact on PD pathway blockade including cancer neoantigen-specific T cell responses and effector T cell tumor infiltration. Nonetheless, these studies raise the possibility that beneficial microorganisms may be an adjuvant for cancer immunotherapy. Thus, it will be scientifically and clinically interesting to profile patient gut microbiota and dissect the relationship with immune responses and clinical outcomes in the course of cancer immunotherapy.

We have discussed several biomarkers in shaping and predicting the clinical response to PD pathway blockade (Fig. 2). Are there definite translational biomarkers for PD pathway blockade? On the basis of the immune profile, cancers may be classified into “inflamed” and “non-inflamed” types. The former is enriched with a T_H1-type immune signature including T_H1-type chemokines and effector T cells (presumably containing mutated antigen-specific T cells) (94) and likely expresses a high amount of PD-L1. The latter is poorly immune-infiltrated and likely expresses a limited amount of PD-L1. Recent clinical studies,

largely from patients with melanoma, suggest that the inflamed, but not the non-inflamed, tumor type is highly responsive to PD pathway blockade (Fig. 2). However, lymphocyte-rich regions may not always be associated with PD-L1 expression (41, 78, 102). Biologically, the non-inflamed tumor type may be closely associated with an epithelial-mesenchymal transition (EMT) and stem-like-type subgroup. In line with this possibility, the T_H1-type immune profile is controlled by stem-like associated oncogenic and epigenetic pathways including β-catenin and PRC2 complex (93–95). Thus, immune inflamed cancers may be a “non-EMT/stem-like type” and are more likely responders to PD blockade therapy. Analogously, the nonresponders (or minimal responders) may be lacking T cell infiltration and T_H1-type chemokines, less specific mutations and neoantigens, and enriched with multiple layers of immunosuppressive mechanisms and potential EMT/stem-like types (Fig. 2). An urgent next step is to define and develop combinatorial therapy to improve and enhance the clinical response in patients with different types of cancer.

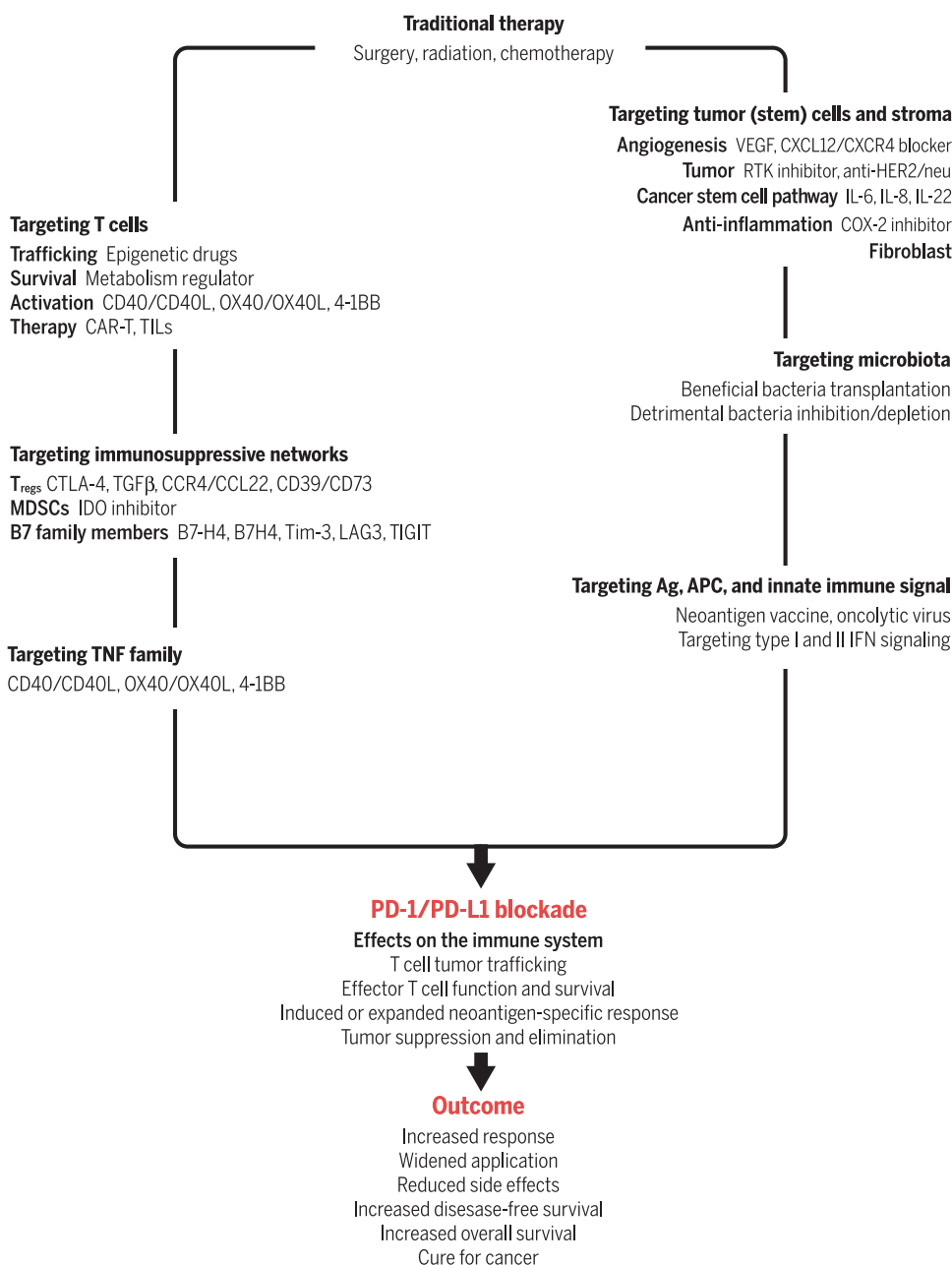


Fig. 4. Scientific rationales of potential therapeutic combinations with PD pathway blockade. Multiple layers of immunosuppressive mechanisms, weak T cell activation, and tumor-intrinsic biological pathways contribute to cancer progression and therapy resistance. The different combinations with PD pathway blockade may yield a synergistic or additive clinical response.

COMBINATORIAL REGIMENS WITH PD PATHWAY BLOCKADE

Because of the complexity of immunoregulatory mechanisms and the heterogeneity of tumor and host, it is envisioned that combination immunotherapies will be required to efficiently treat a larger proportion of cancer patients (1). Continuing advances in our understanding of immunoregulation and tumor immunity will allow for the development of new combination(s) for the treatment of different types of cancer. On the basis of particular limitations of single-agent therapy and combinatorial scientific rationales, therapeutic combinations hold much potential in this area (Fig. 4).

Enforcing effector T cell trafficking with epigenetic reprogramming drugs

T_H1 -type chemokines and effector T cell tumor infiltration are associated with therapeutic responses to PD pathway blockade (Fig. 2). Histone modification and DNA methylation epigenetically repress tumor T_H1 -type chemokines and subsequently determine effector T cell trafficking into the tumor microenvironment (94, 95). It may be reasonable to surmise that cancer epigenetic reprogramming may remove T_H1 -type chemokine repressive marks and promote effector T cell trafficking into the tumor microenvironment and improve the therapeutic efficacy of PD pathway blockade. In support of this, treatment with cancer epigenetic reprogramming drugs including EZH2 inhibitors, DZNep (103) (a selective inhibitor of EZH2 methyltransferase activity), GSK126 (104), and 5-aza-2'-deoxycytidine (5-AZA dC) (a DNMT inhibitor) enhances tumor T_H1 -chemokine production and T cell trafficking into tumor (94, 95) and augments therapeutic effects of PD-L1 blockade and T cell therapy in a preclinical model (94). Furthermore, treatment with azacitidine up-regulates IFN signature genes in several human cancer cell lines (105, 106). 5-AZA dC treatment enhances the cancer and germline antigen NY-ESO-1 expression in human ovarian cancer cells (107) and promotes chemokine expression and T cell tumor trafficking in a mouse ovarian cancer model (108). Thus, epigenetic reprogramming can unlock the repressed T_H1 -type chemokines, IFN signature genes, and tumor antigen expression and may therefore condition tumor from poor T cell infiltration to rich T cell infiltration and ultimately potentiate PD blockade therapy (94, 95, 108).

Supplementation of effector T cells with adoptive T cell therapy

There may be insufficient functional effector T cells in the tumor microenvironment. Adoptive T cell therapy (ACT), including in vitro expanded peripheral blood or TILs and genetically engineered chimeric antigen receptor T cells, is an important therapeutic approach. However, activated human TILs and TAA-specific T cells express PD-1 (19–23, 109). These data suggest a potential benefit for the combination of ACT and PD pathway blockade. In further support of the role of PD-1 blockade in ACT, melanoma TILs with zinc finger nuclease-mediated gene editing of PD-1 display increased effector function in vitro (110). New clinical trials will be warranted to further evaluate this strategy. Mechanistically, PD pathway blockade before ACT may prepare the specific “soil,” the less suppressive tumor microenvironment (1), for the transferred T cells to home and function. Given that transferred T cells are often activated and express PD-1 and immune activation can stimulate PD-L1 expression in tumor cells and APCs in the tumor microenvironment, it is assumed that concurrent or post-PD pathway blockade to ACT may improve the functionality of the transferred T cells.

Promotion of T cell function by targeting TNF family and T cell metabolism

The tumor microenvironment is highly immunosuppressive in patients with advanced cancer. Targeting immunosuppressive mechanisms is considered an effective strategy to treat patients with cancer (1). On the other hand, it remains logical to directly stimulate and activate the immune cells in combination with PD pathway blockade (111). To this end, we can manipulate certain tumor necrosis factor (TNF) family signaling pathways in patients with cancer.

(i) **CD40 and CD40L.** The interaction of CD40 and CD40L delivers a potent costimulatory signal to T cells. The humanized CD40 agonist antibody CP-870893 in combination with chemotherapy is currently in clinical trials to treat patients with pancreatic cancer (112). Two other anti-CD40 antibodies, dacetuzumab and HCD122, are currently being tested in hematologic malignancies (113).

(ii) **OX40 and OX40L.** The interaction of OX40 and OX40L may potentiate T cell activity (114), and agonistic anti-OX40 antibodies and OX40 ligands are currently being studied in clinical trials (115) (NCT02219724, NCT02410512, and NCT02221960).

(iii) **4-1BB (CD137).** CD137 engagement may preferentially stimulate $CD8^+$ T cells and NK cells. Administration of agonistic anti-4-1BB antibody improves T cell immunity in various murine tumor models (116). Recent clinical trials have shown promising results in a single agent or in combination with anti-PD therapy for the treatment of advanced solid tumors (NCT02179918 and NCT02554812).

(iv) **Targeting T cell metabolism.** Recent studies reveal that abnormal metabolism impairs effector T cell function in the tumor microenvironment (13, 117, 118). Reprogramming tumor metabolism would be an interesting option in combination with PD pathway blockade (Fig. 4).

Thus, there are scientific rationales to support the combination with all of these agonistic antibodies and approaches with PD pathway blockade. Clinical studies will be needed to conclusively demonstrate the precise indications, effectiveness, and side effects of given combinations in treating a specific type of cancer.

Subversion of immunosuppressive networks in the tumor microenvironment

Immunosuppressive networks are major obstacles for spontaneous and therapy-induced antitumor immunity (1). The clinical responses observed by blocking inhibitory B7 family members provides solid evidence to target additional immunosuppressive components in the tumor microenvironment.

(i) **Targeting T_{regs} .** T_{regs} actively inhibit T cell-mediated antitumor immunity in the human cancer microenvironment (4). Targeting T_{regs} is proposed as a therapeutic strategy to treat patients with cancer (119–121). T_{regs} express CTLA-4. Anti-CTLA-4 antibody may deplete T_{regs} in the tumor (122). Ipilimumab, a fully human anti-CTLA-4 monoclonal antibody, was approved by the U.S. Food and Drug Administration (FDA) in 2011 for the treatment of patients with advanced melanoma (123). Ipilimumab monotherapy produces clinical responses in 10 to 20% of patients and extends overall survival in metastatic melanoma, with 20% of patients surviving 3 years or longer (123, 124). In a phase 1 trial of anti-PD-1 combined with anti-CTLA-4 in patients with advanced melanoma, rapid and deep tumor regressions were observed in 53% of patients treated at the optimal dose level (58). Phase 2 and 3 clinical trials have confirmed these observations and further demonstrate the beneficial effects on the objective response rate and the progression-free survival among patients with advanced melanoma who had not previously received treatment (Table 1) (63, 76). In patients with tumors demonstrating <5% PD-L1 expression, this combination appears to be more effective than either agent alone (63). However, the response rate for monotherapy with CTLA-4 blockade is generally lower than that with PD blockade in patients with melanoma (Table 1) (13, 62). Durability of responses with each approach is a topic of current study. Nonetheless, anti-CTLA-4 treatment may deplete T_{regs} and potentially increase the ratio between effector T cells and T_{regs} in the

tumor microenvironment. Thus, this combination therapy has been approved by the FDA for the treatment of BRAF V600 wild-type unresectable or metastatic melanoma. This treatment, however, may carry a relatively high frequency of immune-related toxicity. Therefore, a sequenced model with PD pathway blockade therapy first followed by anti-CTLA-4 may later be considered. In addition to anti-CTLA-4, other strategies targeting T_{regs} (121) including transforming growth factor- β signaling blockade may be used in combination with PD blockade (125). T_{regs} migrate into the human cancer microenvironment through CCL22 and CCR4 signaling pathway (4, 119). Blocking this pathway may enhance the effects of PD blockade. An anti-CCR4 antibody (mogamulizumab) can deplete circulating T_{regs} in patients with T cell leukemia and lymphoma (126). Ongoing studies are evaluating the efficacy of anti-CCR4 in combination with PD pathway blockade (NCT02301130). Human tumor-associated T_{regs} express the ectonucleotidases CD39 and CD73, convert adenosine 5'-triphosphate (ATP) to adenosine, and inhibit T cell activation by the adenosinergic pathway (92). A CD73-specific antibody has demonstrated an additive activity when combined with PD-1 antibodies in murine tumor models (127). Thus, CD39, CD73, adenosine, and adenosine A2a receptor (ADORA2A) signaling blockade may be combined with PD pathway therapy to treat patients with cancer.

(ii) Targeting myeloid cells. Human tumor-associated MDSCs and inhibitory APCs including macrophages actively inhibit T cell-mediated antitumor immunity (5–7) and promote cancer stemness (7, 128) in the human cancer microenvironment. Given the roles of indoleamine-2,3-dioxygenase (IDO) in MDSC-mediated T cell suppression (129), the use of IDO inhibitor(s) (INCB024360) (32) with PD blockade is a potential option for combinatorial therapy.

(iii) Targeting additional potentially immune inhibitory immunoglobulin superfamily molecules. The expression of B7-H3 (B7RP-2, CD276) and B7-H4 (B7x, B7S1) is found in different types of human tumor tissues (6, 14, 130, 131). In human hepatocellular carcinoma (HCC), B7-H3 expression is linked to limited T cell proliferation and IFN- γ production (132). B7-H3 blockade resulted in increased CD8⁺ T cell influx in murine pancreatic cancer (133), whereas some studies showed that B7-H3 could promote tumor immunity (134). Tumor-associated macrophages express B7-H4 and are able to suppress tumor-specific T cells (6). Therefore, the immunomodulatory role of B7-H3 and B7-H4 in different types of tumor or host cells is under debate (134, 135). Several clinical studies, however, have been initiated to test the effect of anti-B7-H3 antibodies in a single agent (NCT02628535) or in combination with anti-CTLA-4 (NCT02381314) or with anti-PD-1 (NCT02475213) for treating solid tumors.

PD-1H (Dies1, VISTA, DD1 α) is a more recently identified B7-CD28 family molecule and was shown to be an immune inhibitory ligand in a mouse tumor model and in a mouse experimental autoimmune encephalomyelitis (EAE) model (136). However, using PD-1H-deficient mice and agonistic antibodies, this molecule was also shown to be an immune inhibitory receptor on T cells (137, 138). Thus, VISTA blockade in a single agent or in combination with anti-PD blockade may be potential regimens to be examined in future clinical trials once the necessary reagents are available.

The expression of T cell immunoglobulin mucin 3 (Tim-3) (139, 140), lymphocyte activation gene 3 protein (LAG3) (23), and T cell immunoglobulin and ITIM domain (TIGIT) (141, 142) is reported in human cancer-infiltrating T cells. Blockade of Tim-3 (139, 140), LAG3 (23, 143), and TIGIT (141, 142) with anti-PD antibody in-

creases effector T cell function. Thus, the combinations of PD pathway with Tim-3, LAG3, and TIGIT blockade have been proposed and tested in clinical trials (NCT01968109 and NCT02460224).

Targeting tumor-specific antigen and antigen presentation

(i) Neoantigen vaccine. Traditional vaccines can activate T cells and induce immune responses to targets on tumor cells, but there is not substantial evidence of reproducible clinical responses in patients with established tumors (144). PD pathway blockade can increase the antitumor efficacy of conventional vaccines in animal models (145–147). As somatic genetic mutations could generate unique tumor-specific neoantigens and neoantigen-specific T cell responses (148), vaccination with immunogenic neoantigens has been examined in animal models (81, 82, 89). Mutated antigen-specific T cell responses can be found in patients with cancer (67, 83–86). Thus, a novel vaccination platform will be likely incorporated with specific neoantigens and potentially in combination with PD pathway blockade. Ongoing research efforts are aimed at increasing the efficiency of producing personalized neoepitope vaccines and also possibly identifying shared neoantigens for use along with immune-modulating antibodies. As the tumor microenvironment is immunologically suppressive (1) and metabolically dysregulated (13, 117, 118), in addition to inducing and/or expanding neoantigen-specific T cells through specific vaccination, it is also crucial to ensure that these T cells can efficiently traffic into and survive in the cancer microenvironment (Fig. 4).

(ii) Oncolytic viral therapy. Talimogene laherparepvec (T-VEC) is a herpes simplex virus type 1-derived oncolytic immunotherapy designed to selectively replicate within tumors and produce granulocyte macrophage colony-stimulating factor (GM-CSF) (149). The FDA has approved T-VEC for the local treatment of unresectable cutaneous, subcutaneous, and nodal lesions in patients with recurrent melanoma after initial surgery. T-VEC may likely trigger DC differentiation and enhance antigen presentation, promote T cell activation and IFN- γ production, and induce PD-L1 expression. Thus, T-VEC may be a high-priority agent for combination trials with PD blockade. A phase 3 study is currently exploring T-VEC with pembrolizumab (anti-PD-1) for unresected melanoma (NCT02263508).

Targeting inflammatory mediators with COX-2 inhibitor

Prostaglandin E₂ (PGE₂) and its key synthesizing enzyme cyclooxygenase-2 (COX-2) can directly mediate protumor activities and recruit and induce MDSCs in the tumor microenvironment (150). However, PD pathway blockade may increase the expression of PGE₂ and protumor inflammatory cytokines, which potentially offsets the therapeutic effects of this blockade (151). Several preclinical models demonstrate that inhibition of COX-2 synergizes with PD pathway blockade in eradicating tumors (151, 152), suggesting that COX inhibitors could be useful adjuvants for immune-based therapies including PD blockade in cancer patients.

Targeting innate immune signaling pathway

The innate immune system and its major signaling type I and II IFN contribute to the antitumor immune response (3, 153). Human tumor-associated plasmacytoid DCs induce IL-10⁺ T_{regs} (154, 155). However, after activation, these human tumor-associated plasmacytoid DCs are capable of producing large amount of type I IFN (154, 155). In tumor-bearing mouse models, type I and II IFN signaling pathway is essential

for therapeutic responses to chemotherapy (156, 157), radiation therapy (158, 159), and anti-HER2/neu therapy (160). In line with mouse studies, in women with metastatic breast cancer, response to anti-HER2/neu therapy correlates with NK cell-associated antibody-dependent cell-mediated cytotoxicity (ADCC) (161). Furthermore, PD-L1 expression can be potently stimulated through the IFN signaling pathway. Thus, targeting innate immune signaling pathway in combination with PD pathway blockade is scientifically rationalized.

Targeting cancer cells

(i) Localized radiation. Radiation therapy is a well-recognized means to achieve local tumor destruction. It has been reported to trigger innate immune signaling pathways, impair T_H2 cells, activate CD8⁺ T cells, stimulate chemokine expression, and promote immune infiltration into tumors (158, 159, 162–164). However, radiation-induced inflammation (including IFN signaling) can enhance tumor PD-L1 expression (159, 163), which may reduce radiation-induced protective tumor immunity. Increased PD-L1 expression may provide a window of opportunity for PD pathway blockade. In line with this notion, radiation therapy combined with anti-PD therapy can synergistically promote antitumor immunity in several tumor-bearing mouse models (159, 163, 165, 166). Although there is a solid scientific rationale to support the combination of radiation and PD pathway blockade, radiation parameters including dose, site, and time may be critical to the success of such a combination and need to be further explored. Given the challenges in recapitulating human radiation fractionation regimens in animal models, clinical trials will be essential to carefully sort out the feasibility of this approach. Such clinical trials will also provide an opportunity to examine whether radiation induces immunogenic mutation-associated neoantigens and whether the induced mutations are associated with treatment response.

(ii) Chemotherapy. Mouse studies suggest that therapeutic responses to some chemotherapy agents including anthracyclines and oxaliplatin may partially depend on immune responses, particularly type I IFN signaling-mediated immunity (156, 167). PD-L1 can be induced by the IFN signaling pathway. Oxaliplatin treatment promotes PD-L1⁺ plasmacytoid tumor infiltration in a mouse prostate cancer model (168). These preclinical studies suggest that PD pathway blockade may enhance the efficacy of chemotherapy. As chemotherapy induces genetic mutations, it is also reasonable to hypothesize that such combinations may induce mutation-specific neoantigen-specific T cell responses and affect clinical outcomes. Nonetheless, chemotherapy also modulates the immune system, and PD pathway blockade depends on ongoing immune responses, especially those in the tumor microenvironment (15). Future clinical studies will determine which therapeutic modality, including agents, doses, and timing, will increase clinical responses in combination with PD pathway blockade.

(iii) Targeting oncogenic signals. Anti-HER2/neu antibody therapy and multiple receptor tyrosine kinase (RTK) inhibitors (sunitinib and imatinib) interrupt oncogenic signals and mediate tumor regression. Recent studies indicate that anti-HER2/neu therapy (160) and RTK inhibitors (169) can promote T cell activation and trafficking. PD pathway blockade in combination with anti-HER2/neu antibody or RTK inhibitors may be considered in the treatment of certain cancers. Careful attention is needed for proper dose and scheduling of targeted therapies with immunotherapies to avoid potential suppression of T cell or APC activity, given the physiologic role for some of the targeted pathways.

Other potential combinations

In addition to T cell and APC subsets and tumor cells, vascular endothelial cells, stromal fibroblasts, cancer stem cells, and microbiota may be targeted in combination with PD pathway blockade. Vascular endothelial growth factor A (VEGF-A) and CXCL12 (154, 170, 171) are highly expressed in the tumor microenvironment and mediate tumor angiogenesis. Targeting tumor stromal fibroblasts (172), CXCL12 and CXCR4 blockade (172), and VEGF-targeted therapy (173) have been tested as combinatorial partners for immunotherapy in the literature. As human cancer microenvironmental immune cells including macrophages (128), MDSCs (7), T_H22 cells (12), and inflammatory T_H17 cells (174) and their associated cytokines IL-6 (128, 175, 176), IL-8 (177), and IL-22 (12) can promote and maintain the cancer stem cell pool, targeting cancer stem cell pathway with PD blockade may be an important option. The gut microbiota influence the host responsiveness to immunotherapy (100, 101). Beneficial bacteria inoculation or detrimental bacteria inhibition may be combined with PD pathway blockade in a specific type of cancer. Additional potential combinatory strategies have been discussed in the literature (111).

CONCLUDING REMARKS

PD pathway blockade has elicited durable clinical responses in patients with a broad spectrum of cancers with a reasonable toxicity profile (Table 1). This therapy largely relies on efficient T cell infiltration into tumor and effector T cell function in the tumor microenvironment. The human cancer immune microenvironment thus holds the key to understanding the nature of immunity in response to tumor progression and tumor immunotherapy (1, 15, 33). PD blockade may potentially induce and/or expand T cells specific to mutated neoantigen in patients with cancer. Accumulating evidence points towards mechanism-based combination of various treatment regimens with PD pathway blockade to establish new standard of care for patients with cancer. Dynamic immunologic studies along with genetics and epigenetics in the human cancer microenvironment will guide the development of different combination therapies and generate novel insight into how the human immune system responds to and is shaped by a variety of tumor types.

REFERENCES AND NOTES

1. W. Zou, Immunosuppressive networks in the tumour environment and their therapeutic relevance. *Nat. Rev. Cancer* **5**, 263–274 (2005).
2. D. M. Pardoll, The blockade of immune checkpoints in cancer immunotherapy. *Nat. Rev. Cancer* **12**, 252–264 (2012).
3. T. F. Gajewski, H. Schreiber, Y.-X. Fu, Innate and adaptive immune cells in the tumor microenvironment. *Nat. Immunol.* **14**, 1014–1022 (2013).
4. T. J. Curiel, G. Coukos, L. Zou, X. Alvarez, P. Cheng, P. Mottram, M. Evdemon-Hogan, J. R. Conejo-Garcia, L. Zhang, M. Burow, Y. Zhu, S. Wei, I. Kryczek, B. Daniel, A. Gordon, L. Myers, A. Lackner, M. L. Disis, K. L. Knutson, L. Chen, W. Zou, Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival. *Nat. Med.* **10**, 942–949 (2004).
5. T. J. Curiel, S. Wei, H. Dong, X. Alvarez, P. Cheng, P. Mottram, R. Krzyziak, K. L. Knutson, B. Daniel, M. C. Zimmermann, O. David, M. Burow, A. Gordon, N. Dhurandhar, L. Myers, R. Berggren, A. Hemminki, R. D. Alvarez, D. Emilie, D. T. Curiel, L. Chen, W. Zou, Blockade of B7-H1 improves myeloid dendritic cell-mediated antitumor immunity. *Nat. Med.* **9**, 562–567 (2003).
6. I. Kryczek, L. Zou, P. Rodriguez, G. Zhu, S. Wei, P. Mottram, M. Brumlik, P. Cheng, T. Curiel, L. Myers, A. Lackner, X. Alvarez, A. Ochoa, L. Chen, W. Zou, B7-H4 expression identifies a

- novel suppressive macrophage population in human ovarian carcinoma. *J. Exp. Med.* **203**, 871–881 (2006).
7. T. X. Cui, I. Kryczek, L. Zhao, E. Zhao, R. Kuick, M. H. Roh, L. Vatan, W. Szeliga, Y. Mao, D. G. Thomas, J. Kotarski, R. Tarkowski, M. Wicha, K. Cho, T. Giordano, R. Liu, W. Zou, Myeloid-derived suppressor cells enhance stemness of cancer cells by inducing microRNA101 and suppressing the corepressor CtBP2. *Immunity* **39**, 611–621 (2013).
 8. L. Zhang, J. R. Conejo-García, D. Katsaros, P. A. Gimotty, M. Massobrio, G. Regnani, A. Makrigiannakis, H. Gray, K. Schlienger, M. N. Liebman, S. C. Rubin, G. Coukos, Intratumoral T cells, recurrence, and survival in epithelial ovarian cancer. *N. Engl. J. Med.* **348**, 203–213 (2003).
 9. F. Pagès, A. Berger, M. Camus, F. Sanchez-Cabo, A. Costes, R. Molitor, B. Mlecnik, A. Kirilovsky, M. Nilsson, D. Damotte, T. Meatchi, P. Bruneval, P.-H. Cugnenc, Z. Trajanoski, W.-H. Fridman, J. Galon, Effector memory T cells, early metastasis, and survival in colorectal cancer. *N. Engl. J. Med.* **353**, 2654–2666 (2005).
 10. J. Galon, A. Costes, F. Sanchez-Cabo, A. Kirilovsky, B. Mlecnik, C. Lagorce-Pagès, M. Tosolini, M. Camus, A. Berger, P. Wind, F. Zinzindohoué, P. Bruneval, P.-H. Cugnenc, Z. Trajanoski, W.-H. Fridman, F. Pagès, Type, density, and location of immune cells within human colorectal tumors predict clinical outcome. *Science* **313**, 1960–1964 (2006).
 11. I. Kryczek, E. Zhao, Y. Liu, Y. Wang, L. Vatan, W. Szeliga, J. Moyer, A. Klimczak, A. Lange, W. Zou, Human T_H17 cells are long-lived effector memory cells. *Sci. Transl. Med.* **3**, 104ra100 (2011).
 12. I. Kryczek, Y. Lin, N. Nagarsheth, D. Peng, L. Zhao, E. Zhao, L. Vatan, W. Szeliga, Y. Dou, S. Owens, W. Zgodzinski, M. Majewski, G. Wallner, J. Fang, E. Huang, W. Zou, IL-22⁺CD4⁺ T cells promote colorectal cancer stemness via STAT3 transcription factor activation and induction of the methyltransferase DOT1L. *Immunity* **40**, 772–784 (2014).
 13. E. Zhao, T. Maji, I. Kryczek, W. Li, K. Wu, L. Zhao, S. Wei, J. Crespo, S. Wan, L. Vatan, W. Szeliga, I. Shao, Y. Wang, Y. Liu, S. Varambally, A. M. Chinnaiyan, T. H. Welling, V. Marquez, J. Kotarski, H. Wang, Z. Wang, Y. Zhang, R. Liu, G. Wang, W. Zou, Cancer mediates effector T cell dysfunction by targeting microRNAs and EZH2 via glycolysis restriction. *Nat. Immunol.* **17**, 95–103 (2016).
 14. W. Zou, L. Chen, Inhibitory B7-family molecules in the tumour microenvironment. *Nat. Rev. Immunol.* **8**, 467–477 (2008).
 15. L. Chen, X. Han, Anti-PD-1/PD-L1 therapy of human cancer: Past, present, and future. *J. Clin. Invest.* **125**, 3384–3391 (2015).
 16. H. Dong, G. Zhu, K. Tamada, L. Chen, B7-H1, a third member of the B7 family, co-stimulates T-cell proliferation and interleukin-10 secretion. *Nat. Med.* **5**, 1365–1369 (1999).
 17. H. Dong, S. E. Strome, D. R. Salomao, H. Tamura, F. Hirano, D. B. Flies, P. C. Roche, J. Lu, G. Zhu, K. Tamada, V. A. Lennon, E. Celis, L. Chen, Tumor-associated B7-H1 promotes T-cell apoptosis: A potential mechanism of immune evasion. *Nat. Med.* **8**, 793–800 (2002).
 18. G. J. Freeman, A. J. Long, Y. Iwai, K. Bourque, T. Chernova, H. Nishimura, L. J. Fitz, N. Malenkovich, T. Okazaki, M. C. Byrne, H. F. Horton, L. Fouser, L. Carter, V. Ling, M. R. Bowman, B. M. Carreno, M. Collins, C. R. Wood, T. Honjo, Engagement of the Pd-1 immunoinhibitory receptor by a novel B7 family member leads to negative regulation of lymphocyte activation. *J. Exp. Med.* **192**, 1027–1034 (2000).
 19. R. M. Wong, R. R. Scotland, R. L. Lau, C. Wang, A. J. Korman, W. M. Kast, J. S. Weber, Programmed death-1 blockade enhances expansion and functional capacity of human melanoma antigen-specific CTLs. *Int. Immunol.* **19**, 1223–1234 (2007).
 20. K. Wu, I. Kryczek, L. Chen, W. Zou, T. H. Welling, Kupffer cell suppression of CD8⁺ T cells in human hepatocellular carcinoma is mediated by B7-H1/programmed death-1 interactions. *Cancer Res.* **69**, 8067–8075 (2009).
 21. M. Ahmadzadeh, L. A. Johnson, B. Heemskerk, J. R. Wunderlich, M. E. Dudley, D. E. White, S. A. Rosenberg, Tumor antigen-specific CD8 T cells infiltrating the tumor express high levels of PD-1 and are functionally impaired. *Blood* **114**, 1537–1544 (2009).
 22. J. Fourcade, Z. Sun, M. Benallaoua, P. Guillaume, I. F. Luescher, C. Sander, J. M. Kirkwood, V. Kuchroo, H. M. Zarour, Upregulation of Tim-3 and PD-1 expression is associated with tumor antigen-specific CD8⁺ T cell dysfunction in melanoma patients. *J. Exp. Med.* **207**, 2175–2186 (2010).
 23. J. Matsuzaki, S. Gnjatic, P. Mhawech-Fauceglia, A. Beck, A. Miller, T. Tsuji, C. Eppolito, F. Qian, S. Lele, P. Shrikant, L. J. Old, K. Odunsi, Tumor-infiltrating NY-ESO-1-specific CD8⁺ T cells are negatively regulated by LAG-3 and PD-1 in human ovarian cancer. *Proc. Natl. Acad. Sci. U.S.A.* **107**, 7875–7880 (2010).
 24. I. Perrot, D. Blanchard, N. Freymond, S. Isaac, B. Guibert, Y. Pacheco, S. Lebecque, Dendritic cells infiltrating human non-small cell lung cancer are blocked at immature stage. *J. Immunol.* **178**, 2763–2769 (2007).
 25. D.-M. Kuang, Q. Zhao, C. Peng, J. Xu, J.-P. Zhang, C. Wu, L. Zheng, Activated monocytes in peritumoral stroma of hepatocellular carcinoma foster immune privilege and disease progression through PD-L1. *J. Exp. Med.* **206**, 1327–1337 (2009).
 26. M. R. Nazareth, L. Broderick, M. R. Simpson-Abelson, R. J. Kelleher Jr., S. J. Yokota, R. B. Bankert, Characterization of human lung tumor-associated fibroblasts and their ability to modulate the activation of tumor-associated T cells. *J. Immunol.* **178**, 5552–5562 (2007).
 27. R. H. Thompson, M. D. Gillett, J. C. Chevillet, C. M. Lohse, H. Dong, W. S. Webster, K. G. Krejci, J. R. Lobo, S. Sengupta, L. Chen, H. Zincke, M. L. Blute, S. E. Strome, B. C. Leibovich, E. D. Kwon, Costimulatory B7-H1 in renal cell carcinoma patients: Indicator of tumor aggressiveness and potential therapeutic target. *Proc. Natl. Acad. Sci. U.S.A.* **101**, 17174–17179 (2004).
 28. H. Ghebeh, S. Mohammed, A. Al-Omair, A. Qattant, C. Lehe, G. Al-Qudaihi, N. Elkum, M. Alshabanah, S. Bin Amer, A. Tulbah, D. Ajarim, T. Al-Tweigeri, S. Dermime, The B7-H1 (PD-L1) T lymphocyte-inhibitory molecule is expressed in breast cancer patients with infiltrating ductal carcinoma: Correlation with important high-risk prognostic factors. *Neoplasia* **8**, 190–198 (2006).
 29. J. A. Brown, D. M. Dorfman, F.-R. Ma, E. L. Sullivan, O. Munoz, C. R. Wood, E. A. Greenfield, G. J. Freeman, Blockade of programmed death-1 ligands on dendritic cells enhances T cell activation and cytokine production. *J. Immunol.* **170**, 1257–1266 (2003).
 30. I. Kryczek, S. Wei, W. Gong, X. Shu, W. Szeliga, L. Vatan, L. Chen, G. Wang, W. Zou, Cutting edge: IFN- γ enables APC to promote memory Th17 and abate Th1 cell development. *J. Immunol.* **181**, 5842–5846 (2008).
 31. J. M. Taube, R. A. Anders, G. D. Young, H. Xu, R. Sharma, T. L. McMiller, S. Chen, A. P. Klein, D. M. Pardoll, S. L. Topalian, L. Chen, Colocalization of inflammatory response with B7-H1 expression in human melanocytic lesions supports an adaptive resistance mechanism of immune escape. *Sci. Transl. Med.* **4**, 127ra37 (2012).
 32. S. Spranger, R. M. Spaapan, Y. Zha, J. Williams, Y. Meng, T. T. Ha, T. F. Gajewski, Up-regulation of PD-L1, IDO, and T_Hregs in the melanoma tumor microenvironment is driven by CD8⁺ T cells. *Sci. Transl. Med.* **5**, 200ra116 (2013).
 33. S. L. Topalian, C. G. Drake, D. M. Pardoll, Immune checkpoint blockade: A common denominator approach to cancer therapy. *Cancer Cell* **27**, 450–461 (2015).
 34. A. T. Parsa, J. S. Waldron, A. Panner, C. A. Crane, I. F. Parney, J. J. Barry, K. E. Cachola, J. C. Murray, T. Tihan, M. C. Jensen, P. S. Mischel, D. Stokoe, R. O. Pieper, Loss of tumor suppressor PTEN function increases B7-H1 expression and immunoresistance in glioma. *Nat. Med.* **13**, 84–88 (2007).
 35. E. A. Mittendorf, A. V. Philips, F. Meric-Bernstam, N. Qiao, Y. Wu, S. Harrington, X. Su, Y. Wang, A. M. Gonzalez-Angulo, A. Akcakanat, A. Chawla, M. Curran, P. Hwu, P. Sharma, J. K. Litton, J. J. Mordrem, G. Alatrash, PD-L1 expression in triple-negative breast cancer. *Cancer Immunol. Res.* **2**, 361–370 (2014).
 36. M. Marzec, Q. Zhang, A. Goradia, P. N. Raghunath, X. Liu, M. Paessler, H. Y. Wang, M. Wysocka, M. Cheng, B. A. Ruggeri, M. A. Wasik, Oncogenic kinase NPM/ALK induces through STAT3 expression of immunosuppressive protein CD274 (PD-L1, B7-H1). *Proc. Natl. Acad. Sci. U.S.A.* **105**, 20852–20857 (2008).
 37. Y. Agata, A. Kawasaki, H. Nishimura, Y. Ishida, T. Tsubata, H. Yagita, T. Honjo, Expression of the PD-1 antigen on the surface of stimulated mouse T and B lymphocytes. *Int. Immunol.* **8**, 765–772 (1996).
 38. D. L. Barber, E. J. Wherry, D. Masopust, B. Zhu, J. P. Allison, A. H. Sharpe, G. J. Freeman, R. Ahmed, Restoring function in exhausted CD8 T cells during chronic viral infection. *Nature* **439**, 682–687 (2006).
 39. C. L. Day, D. E. Kaufmann, P. Kiepiela, J. A. Brown, E. S. Moodley, S. Reddy, E. W. Mackey, J. D. Miller, A. J. Leslie, C. DePierres, Z. Mncube, J. Duraiswamy, B. Zhu, Q. Eichbaum, M. Altfeld, E. J. Wherry, H. M. Coovadia, P. J. R. Goulder, P. Klennerman, R. Ahmed, G. J. Freeman, B. D. Walker, PD-1 expression on HIV-specific T cells is associated with T-cell exhaustion and disease progression. *Nature* **443**, 350–354 (2006).
 40. S. Kleffel, C. Posch, S. R. Barthel, H. Mueller, C. Schlapbach, E. Guenova, C. P. Elco, N. Lee, V. R. Juneja, Q. Zhan, C. G. Lian, R. Thomi, W. Hoetzenecker, A. Cozzio, R. Dummer, M. C. Mihm Jr, K. T. Flaherty, M. H. Frank, G. F. Murphy, A. H. Sharpe, T. S. Kupper, T. Schatton, Melanoma cell-intrinsic PD-1 receptor functions promote tumor growth. *Cell* **162**, 1242–1256 (2015).
 41. J. M. Taube, A. Klein, J. R. Brahmer, H. Xu, X. Pan, J. H. Kim, L. Chen, D. M. Pardoll, S. L. Topalian, R. A. Anders, Association of PD-1, PD-1 ligands, and other features of the tumor immune microenvironment with response to anti-PD-1 therapy. *Clin. Cancer Res.* **20**, 5064–5074 (2014).
 42. M. J. Butte, M. E. Keir, T. B. Phamduy, A. H. Sharpe, G. J. Freeman, Programmed death-1 ligand 1 interacts specifically with the B7-1 costimulatory molecule to inhibit T cell responses. *Immunity* **27**, 111–122 (2007).
 43. J.-J. Park, R. Omiya, Y. Matsumura, Y. Sakoda, A. Kuramasu, M. M. Augustine, S. Yao, F. Tushima, H. Narazaki, S. Anand, Y. Liu, S. E. Strome, L. Chen, K. Tamada, B7-H1/CD80 interaction is required for the induction and maintenance of peripheral T-cell tolerance. *Blood* **116**, 1291–1298 (2010).
 44. H. Dong, S. E. Strome, E. L. Matteson, K. G. Moder, D. B. Flies, G. Zhu, H. Tamura, C. L. W. Driscoll, L. Chen, Costimulating aberrant T cell responses by B7-H1 autoantibodies in rheumatoid arthritis. *J. Clin. Invest.* **111**, 363–370 (2003).
 45. T. Azuma, S. Yao, G. Zhu, A. S. Flies, S. J. Flies, L. Chen, B7-H1 is a ubiquitous antiapoptotic receptor on cancer cells. *Blood* **111**, 3635–3643 (2008).
 46. Y. Xiao, S. Yu, B. Zhu, D. Bedoret, X. Bu, L. M. Francisco, P. Hua, J. S. Duke-Cohan, D. T. Umetsu, A. H. Sharpe, R. H. DeKruyff, G. J. Freeman, RGMb is a novel binding partner for PD-L2 and its engagement with PD-L2 promotes respiratory tolerance. *J. Exp. Med.* **211**, 943–959 (2014).
 47. T. Powles, J. P. Eder, G. D. Fine, F. S. Braiteh, Y. Liorot, C. Cruz, J. Bellmunt, H. A. Burris, D. P. Petrylak, S.-I. Teng, X. Shen, Z. Boyd, P. S. Hegde, D. S. Chen, N. J. Vogelzang, MPDL3280A

- (anti-PD-L1) treatment leads to clinical activity in metastatic bladder cancer. *Nature* **515**, 558–562 (2014).
48. J. R. Brahmer, S. S. Tykodi, L. Q. M. Chow, W.-J. Hwu, S. L. Topalian, P. Hwu, C. G. Drake, L. H. Camacho, J. Kauh, K. Odunsi, H. C. Pitot, O. Hamid, S. Bhatia, R. Martins, K. Eaton, S. Chen, T. M. Salay, S. Alaparthy, J. F. Grosso, A. J. Korman, S. M. Parker, S. Agrawal, S. M. Goldberg, D. M. Pardoll, A. Gupta, J. M. Wigginton, Safety and activity of anti-PD-L1 antibody in patients with advanced cancer. *N. Engl. J. Med.* **366**, 2455–2465 (2012).
 49. R. S. Herbst, J.-C. Soria, M. Kowanetz, G. D. Fine, O. Hamid, M. S. Gordon, J. A. Sosman, D. F. McDermott, J. D. Powderly, S. N. Gettinger, H. E. K. Kohrt, L. Horn, D. P. Lawrence, S. Rost, M. Leabman, Y. Xiao, A. Mokatri, H. Koeppen, P. S. Hegde, I. Mellman, D. S. Chen, F. S. Hodi, Predictive correlates of response to the anti-PD-L1 antibody MPDL3280A in cancer patients. *Nature* **515**, 563–567 (2014).
 50. J. R. Brahmer, C. G. Drake, I. Wollner, J. D. Powderly, J. Picus, W. H. Sharfman, E. Stankevich, A. Pons, T. M. Salay, T. L. McMiller, M. M. Gilson, C. Wang, M. Selby, J. M. Taube, R. Anders, L. Chen, A. J. Korman, D. M. Pardoll, I. Lowy, S. L. Topalian, Phase I study of single-agent anti-programmed death-1 (MDX-1106) in refractory solid tumors: Safety, clinical activity, pharmacodynamics, and immunologic correlates. *J. Clin. Oncol.* **28**, 3167–3175 (2010).
 51. S. L. Topalian, F. S. Hodi, J. R. Brahmer, S. N. Gettinger, D. C. Smith, D. F. McDermott, J. D. Powderly, R. D. Carvajal, J. A. Sosman, M. B. Atkins, P. D. Leming, D. R. Spigel, S. J. Antonia, L. Horn, C. G. Drake, D. M. Pardoll, L. Chen, W. H. Sharfman, R. A. Anders, J. M. Taube, T. L. McMiller, H. Xu, A. J. Korman, M. Jure-Kunkel, S. Agrawal, D. McDonald, G. D. Kollia, A. Gupta, J. M. Wigginton, M. Sznol, Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. *N. Engl. J. Med.* **366**, 2443–2454 (2012).
 52. E. J. Lipson, W. H. Sharfman, C. G. Drake, I. Wollner, J. M. Taube, R. A. Anders, H. Xu, S. Yao, A. Pons, L. Chen, D. M. Pardoll, J. R. Brahmer, S. L. Topalian, Durable cancer regression off-treatment and effective reinduction therapy with an anti-PD-1 antibody. *Clin. Cancer Res.* **19**, 462–468 (2013).
 53. D. T. Le, J. N. Uram, H. Wang, B. R. Bartlett, H. Kemberling, A. D. Eyring, A. D. Skora, B. S. Luber, N. S. Azad, D. Laheru, B. Biedrzycki, R. C. Donehower, A. Zaheer, G. A. Fisher, T. S. Crocenzi, J. J. Lee, S. M. Duffy, R. M. Goldberg, A. de la Chapelle, M. Koshiji, F. Bhajee, T. Huebner, R. H. Hruban, L. D. Wood, N. Cuka, D. M. Pardoll, N. Papadopoulos, K. W. Kinzler, S. Zhou, T. C. Comish, J. M. Taube, R. A. Anders, J. R. Eshleman, B. Vogelstein, L. A. Diaz Jr, PD-1 blockade in tumors with mismatch-repair deficiency. *N. Engl. J. Med.* **372**, 2509–2520 (2015).
 54. P. Armand, A. Nagler, E. A. Weller, S. M. Devine, D. E. Avigan, Y.-B. Chen, M. S. Kaminski, H. K. Holland, J. N. Winter, J. R. Mason, J. W. Fay, D. A. Rizzieri, C. M. Hosing, E. D. Ball, J. P. Uberti, H. M. Lazarus, M. Y. Mapara, S. A. Gregory, J. M. Timmerman, D. Andorsky, R. Or, E. K. Waller, R. Rotem-Yehudar, L. I. Gordon, Disabling immune tolerance by programmed death-1 blockade with pidlizumab after autologous hematopoietic stem-cell transplantation for diffuse large B-cell lymphoma: Results of an international phase II trial. *J. Clin. Oncol.* **31**, 4199–4206 (2013).
 55. J. R. Westin, F. Chu, M. Zhang, L. E. Fayad, L. W. Kwak, N. Fowler, J. Romaguera, F. Hagemeister, M. Fanale, F. Samaniego, L. Feng, V. Baladandayuthapani, Z. Wang, W. Ma, Y. Gao, M. Wallace, L. M. Vence, L. Radvanyi, T. Muzzafar, R. Rotem-Yehudar, R. E. Davis, S. S. Neelapu, Safety and activity of PD1 blockade by pidlizumab in combination with rituximab in patients with relapsed follicular lymphoma: A single group, open-label, phase 2 trial. *Lancet Oncol.* **15**, 69–77 (2014).
 56. S. M. Ansell, A. M. Lesokhin, I. Borrello, A. Halwani, E. C. Scott, M. Gutierrez, S. J. Schuster, M. M. Millenson, D. Cattray, G. J. Freeman, S. J. Rodig, B. Chapuy, A. H. Ligon, L. Zhu, J. F. Grosso, S. Y. Kim, J. M. Timmerman, M. A. Shipp, P. Armand, PD-1 blockade with nivolumab in relapsed or refractory Hodgkin's lymphoma. *N. Engl. J. Med.* **372**, 311–319 (2015).
 57. O. Hamid, C. Robert, A. Daud, F. S. Hodi, W.-J. Hwu, R. Kefford, J. D. Wolchok, P. Hersey, R. W. Joseph, J. S. Weber, R. Dronca, T. C. Gangadhar, A. Patnaik, H. Zarour, A. M. Joshua, K. Gergich, J. Elassaiss-Schaap, A. Algazi, C. Mateus, P. Boasberg, P. C. Tume, B. Chmielowski, S. W. Ebbinghaus, X. N. Li, S. P. Kang, A. Ribas, Safety and tumor responses with lambrolizumab (anti-PD-1) in melanoma. *N. Engl. J. Med.* **369**, 134–144 (2013).
 58. J. D. Wolchok, H. Kluger, M. K. Callahan, M. A. Postow, N. A. Rizvi, A. M. Lesokhin, N. H. Segal, C. E. Ariyan, R.-A. Gordon, K. Reed, M. M. Burke, A. Caldwell, S. A. Kronenberg, B. U. Agunwamba, X. Zhang, I. Lowy, H. D. Inzunza, W. Feely, C. E. Horak, Q. Hong, A. J. Korman, J. M. Wigginton, A. Gupta, M. Sznol, Nivolumab plus ipilimumab in advanced melanoma. *N. Engl. J. Med.* **369**, 122–133 (2013).
 59. J. S. Weber, R. R. Kudchadkar, B. Yu, D. Gallenstein, C. E. Horak, H. D. Inzunza, X. Zhao, A. J. Martinez, W. Wang, G. Gibney, J. Kroeger, C. Eysmans, A. A. Sarnaik, Y. A. Chen, Safety, efficacy, and biomarkers of nivolumab with vaccine in ipilimumab-refractory or -naive melanoma. *J. Clin. Oncol.* **31**, 4311–4318 (2013).
 60. S. L. Topalian, M. Sznol, D. F. McDermott, H. M. Kluger, R. D. Carvajal, W. H. Sharfman, J. R. Brahmer, D. P. Lawrence, M. B. Atkins, J. D. Powderly, P. D. Leming, E. J. Lipson, I. Puzanov, D. C. Smith, J. M. Taube, J. M. Wigginton, G. D. Kollia, A. Gupta, D. M. Pardoll, J. A. Sosman, F. S. Hodi, Survival, durable tumor remission, and long-term safety in patients with advanced melanoma receiving nivolumab. *J. Clin. Oncol.* **32**, 1020–1030 (2014).
 61. C. Robert, A. Ribas, J. D. Wolchok, F. S. Hodi, O. Hamid, R. Kefford, J. S. Weber, A. M. Joshua, W.-J. Hwu, T. C. Gangadhar, A. Patnaik, R. Dronca, H. Zarour, R. W. Joseph, P. Boasberg, B. Chmielowski, C. Mateus, M. A. Postow, K. Gergich, J. Elassaiss-Schaap, X. N. Li, R. Iannone, S. W. Ebbinghaus, S. P. Kang, A. Daud, Anti-programmed-death-receptor-1 treatment with pembrolizumab in ipilimumab-refractory advanced melanoma: A randomised dose-comparison cohort of a phase 1 trial. *Lancet* **384**, 1109–1117 (2014).
 62. C. Robert, J. Schachter, G. V. Long, A. Arance, J. J. Grob, L. Mortier, A. Daud, M. S. Carlino, C. McNeil, M. Lotem, J. Larkin, P. Lorigan, B. Neyns, C. U. Blank, O. Hamid, C. Mateus, R. Shapira-Frommer, M. Kosh, H. Zhou, N. Ibrahim, S. Ebbinghaus, A. Ribas; KEYNOTE-006 investigators, Pembrolizumab versus ipilimumab in advanced melanoma. *N. Engl. J. Med.* **372**, 2521–2532 (2015).
 63. J. Larkin, V. Chiarion-Sileni, R. Gonzalez, J. J. Grob, C. L. Cowey, C. D. Lao, D. Schadendorf, R. Dummer, M. Smylie, P. Rutkowski, P. F. Ferrucci, A. Hill, J. Wagstaff, M. S. Carlino, J. B. Haanen, M. Maio, I. Marquez-Rodas, G. A. McArthur, P. A. Ascierto, G. V. Long, M. K. Callahan, M. A. Postow, K. Grossmann, M. Sznol, B. Dreno, L. Bastholt, A. Yang, L. M. Rollin, C. Horak, F. S. Hodi, J. D. Wolchok, Combined nivolumab and ipilimumab or monotherapy in untreated melanoma. *N. Engl. J. Med.* **373**, 23–34 (2015).
 64. C. Robert, G. V. Long, B. Brady, C. Dutriaux, M. Maio, L. Mortier, J. C. Hassel, P. Rutkowski, C. McNeil, E. Kalinka-Warchoła, K. J. Savage, M. M. Hemberg, C. Lebbé, J. Charles, C. Mihalcioiu, V. Chiarion-Sileni, C. Mauch, F. Cognetti, A. Arance, H. Schmidt, D. Schadendorf, H. Gogas, L. Lundgren-Eriksson, C. Horak, B. Sharkey, I. M. Waxman, V. Atkinson, P. A. Ascierto, Nivolumab in previously untreated melanoma without BRAF mutation. *N. Engl. J. Med.* **372**, 320–330 (2015).
 65. J. S. Weber, S. P. D'Angelo, D. Minor, F. S. Hodi, R. Gutzmer, B. Neyns, C. Hoeller, N. I. Khusalani, W. H. Miller Jr, C. D. Lao, G. P. Linette, L. Thomas, P. Lorigan, K. F. Grossmann, J. C. Hassel, M. Maio, M. Sznol, P. A. Ascierto, P. Mohr, B. Chmielowski, A. Bryce, I. M. Svane, J.-J. Grob, A. M. Krackhardt, C. Horak, A. Lambert, A. S. Yang, J. Larkin, Nivolumab versus chemotherapy in patients with advanced melanoma who progressed after anti-CTLA-4 treatment (CheckMate 037): A randomised, controlled, open-label, phase 3 trial. *Lancet Oncol.* **16**, 375–384 (2015).
 66. E. B. Garon, N. A. Rizvi, R. Hui, N. Leighl, A. S. Balmanoukian, J. P. Eder, A. Patnaik, C. Aggarwal, M. Gubens, L. Horn, E. Carcereny, M.-J. Ahn, E. Felip, J.-S. Lee, M. D. Hellmann, O. Hamid, J. W. Goldman, J.-C. Soria, M. Dolled-Filhart, R. Z. Rutledge, J. Zhang, J. K. Luceford, R. Rangwala, G. M. Lubiniecki, C. Roach, K. Emancipator, L. Gandhi; KEYNOTE-001 Investigators, Pembrolizumab for the treatment of non-small-cell lung cancer. *N. Engl. J. Med.* **372**, 2018–2028 (2015).
 67. N. A. Rizvi, M. D. Hellmann, A. Snyder, P. Kvistner, V. Makarov, J. J. Havel, W. Lee, J. Yuan, P. Wong, T. S. Ho, M. L. Miller, N. Rekhtman, A. L. Moreira, F. Ibrahim, C. Bruggeman, B. Gasmir, R. Zappasodi, Y. Maeda, C. Sander, E. B. Garon, T. Merghoub, J. D. Wolchok, T. N. Schumacher, T. A. Chan, Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. *Science* **348**, 124–128 (2015).
 68. J. Brahmer, K. L. Reckamp, P. Baas, L. Crinó, W. E. Eberhardt, E. Poddubska, S. Antonia, A. Pluzanski, E. E. Vokes, E. Holgado, D. Waterhouse, N. Ready, J. Gainor, O. Arén Frontera, L. Havel, M. Steins, M. C. Garassino, J. G. Aerts, M. Domine, L. Paz-Ares, M. Reck, C. Baudelet, C. T. Harbison, B. Lestini, D. R. Spigel, Nivolumab versus docetaxel in advanced squamous-cell non-small-cell lung cancer. *N. Engl. J. Med.* **373**, 123–135 (2015).
 69. S. N. Gettinger, L. Horn, L. Gandhi, D. R. Spigel, S. J. Antonia, N. A. Rizvi, J. D. Powderly, R. S. Heist, R. D. Carvajal, D. M. Jackman, L. V. Sequist, D. C. Smith, P. Leming, D. P. Carbone, M. C. Pinder-Schenck, S. L. Topalian, F. S. Hodi, J. A. Sosman, M. Sznol, D. F. McDermott, D. M. Pardoll, V. Sankar, C. M. Ahlers, M. Salvati, J. M. Wigginton, M. D. Hellmann, G. D. Kollia, A. K. Gupta, J. R. Brahmer, Overall survival and long-term safety of nivolumab (anti-programmed death 1 antibody, BMS-936558, ONO-4538) in patients with previously treated advanced non-small-cell lung cancer. *J. Clin. Oncol.* **33**, 2004–2012 (2015).
 70. N. A. Rizvi, J. Mazières, D. Planchard, T. E. Stinchcombe, G. K. Dy, S. J. Antonia, L. Horn, H. Lena, E. Minenza, B. Mennezier, G. A. Otterson, L. T. Campos, D. R. Gandara, B. P. Levy, S. G. Nair, G. Zalcman, J. Wolf, P.-J. Souquet, E. Baldini, F. Cappuzzo, C. Chouaid, A. Dowlati, R. Sanborn, A. Lopez-Chavez, C. Grohe, R. M. Huber, C. T. Harbison, C. Baudelet, B. J. Lestini, S. S. Ramalingam, Activity and safety of nivolumab, an anti-PD-1 immune checkpoint inhibitor, for patients with advanced, refractory squamous non-small-cell lung cancer (CheckMate 063): A phase 2, single-arm trial. *Lancet Oncol.* **16**, 257–265 (2015).
 71. D. F. McDermott, C. G. Drake, M. Sznol, T. K. Choueiri, J. D. Powderly, D. C. Smith, J. R. Brahmer, R. D. Carvajal, H. J. Hammers, I. Puzanov, F. S. Hodi, H. M. Kluger, S. L. Topalian, D. M. Pardoll, J. M. Wigginton, G. D. Kollia, A. Gupta, D. McDonald, V. Sankar, J. A. Sosman, M. B. Atkins, Survival, durable response, and long-term safety in patients with previously treated advanced renal cell carcinoma receiving nivolumab. *J. Clin. Oncol.* **33**, 2013–2020 (2015).
 72. R. H. Thompson, S. M. Kuntz, B. C. Leibovich, H. Dong, C. M. Lohse, W. S. Webster, S. Sengupta, I. Frank, A. S. Parker, H. Zincke, M. L. Blute, T. J. Sebo, J. C. Cheville, E. D. Kwon, Tumor B7-H1 is associated with poor prognosis in renal cell carcinoma patients with long-term follow-up. *Cancer Res.* **66**, 3381–3385 (2006).
 73. Y. Ohgashi, M. Sho, Y. Yamada, Y. Tsurui, K. Hamada, N. Ikeda, T. Mizuno, R. Yoriki, H. Kashizuka, K. Yane, F. Tsushima, N. Otsuki, H. Yagita, M. Azuma, Y. Nakajima, Clinical significance of programmed death-1 ligand-1 and programmed death-1 ligand-2 expression in human esophageal cancer. *Clin. Cancer Res.* **11**, 2947–2953 (2005).
 74. C. Wu, Y. Zhu, J. Jiang, J. Zhao, X.-G. Zhang, N. Xu, Immunohistochemical localization of programmed death-1 ligand-1 (PD-L1) in gastric carcinoma and its clinical significance. *Acta Histochem.* **108**, 19–24 (2006).

75. J. Hamanishi, M. Mandai, M. Iwasaki, T. Okazaki, Y. Tanaka, K. Yamaguchi, T. Higuchi, H. Yagi, K. Takakura, N. Minato, T. Horjo, S. Fujii, Programmed cell death 1 ligand 1 and tumor-infiltrating CD8⁺ T lymphocytes are prognostic factors of human ovarian cancer. *Proc. Natl. Acad. Sci. U.S.A.* **104**, 3360–3365 (2007).
76. M. A. Postow, J. Chesney, A. C. Pavlick, C. Robert, K. Grossmann, D. McDermott, G. P. Linette, N. Meyer, J. K. Giguere, S. S. Agarwala, M. Shaheen, M. S. Ernstoff, D. Minor, A. K. Salama, M. Taylor, P. A. Ott, L. M. Rollin, C. Horak, P. Gagnier, J. D. Wolchok, F. S. Hodi, Nivolumab and ipilimumab versus ipilimumab in untreated melanoma. *N. Engl. J. Med.* **372**, 2006–2017 (2015).
77. R. L. Maute, S. R. Gordon, A. T. Mayer, M. N. McCracken, A. Natarajan, N. G. Ring, R. Kimura, J. M. Tsai, A. Manglik, A. C. Kruse, S. S. Gambhir, I. L. Weissman, A. M. Ring, Engineering high-affinity PD-1 variants for optimized immunotherapy and immuno-PET imaging. *Proc. Natl. Acad. Sci. U.S.A.* **112**, E6506–E6514 (2015).
78. H. M. Kluger, C. R. Zito, M. L. Barr, M. K. Baine, V. L. S. Chiang, M. Sznol, D. L. Rimm, L. Chen, L. B. Jilaveanu, Characterization of PD-L1 expression and associated T-cell infiltrates in metastatic melanoma samples from variable anatomic sites. *Clin. Cancer Res.* **21**, 3052–3060 (2015).
79. M. DuPage, C. Mazumdar, L. M. Schmidt, A. F. Cheung, T. Jacks, Expression of tumour-specific antigens underlies cancer immunoeediting. *Nature* **482**, 405–409 (2012).
80. H. Matsushita, M. D. Vesely, D. C. Koboldt, C. G. Rickert, R. Uppaluri, V. J. Magrini, C. D. Arthur, J. M. White, Y.-S. Chen, L. K. Shea, J. Hundal, M. C. Wendt, R. Demeter, T. Wylie, J. P. Allison, M. J. Smyth, L. J. Old, E. R. Mardis, R. D. Schreiber, Cancer exome analysis reveals a T-cell-dependent mechanism of cancer immunoeediting. *Nature* **482**, 400–404 (2012).
81. T. Schumacher, L. Bunse, S. Pusch, F. Sahm, B. Wiestler, J. Quandt, O. Menn, M. Osswald, I. Oezen, M. Ott, M. Keil, J. Balb, K. Rauschenbach, A. K. Grabowska, I. Vogler, J. Diekmann, N. Trautwein, S. B. Eichmüller, J. Okun, S. Stevanović, A. B. Riemer, U. Sahin, M. A. Friese, P. Beckhove, A. von Deimling, W. Wick, M. Platten, A vaccine targeting mutant IDH1 induces antitumour immunity. *Nature* **512**, 324–327 (2014).
82. S. Kreiter, M. Vormehr, N. van de Roemer, M. Diken, M. Löwer, J. Diekmann, S. Boegel, B. Schrörs, F. Vascotto, J. C. Castle, A. D. Tadmor, S. P. Schoenberger, C. Huber, Ö. Türeci, U. Sahin, Mutant MHC class II epitopes drive therapeutic immune responses to cancer. *Nature* **520**, 692–696 (2015).
83. E. Tran, S. Turcotte, A. Gros, P. F. Robbins, Y.-C. Lu, M. E. Dudley, J. R. Wunderlich, R. P. Somerville, K. Hogan, C. S. Hinrichs, M. R. Parkhurst, J. C. Yang, S. A. Rosenberg, Cancer immunotherapy based on mutation-specific CD4⁺ T cells in a patient with epithelial cancer. *Science* **344**, 641–645 (2014).
84. C. Linnemann, M. M. van Buuren, L. Bies, E. M. E. Verdegaal, R. Schotte, J. J. A. Calis, S. Behjati, A. Velds, H. Hilkmann, D. e. Atmioui, M. Visser, M. R. Stratton, J. B. A. G. Haanen, H. Spits, S. H. van der Burg, T. N. M. Schumacher, High-throughput epitope discovery reveals frequent recognition of neo-antigens by CD4⁺ T cells in human melanoma. *Nat. Med.* **21**, 81–85 (2015).
85. P. F. Robbins, Y.-C. Lu, M. El-Gamil, Y. F. Li, C. Gross, J. Gartner, J. C. Lin, J. K. Teer, P. Cliften, E. Tycksen, Y. Samuels, S. A. Rosenberg, Mining exomic sequencing data to identify mutated antigens recognized by adoptively transferred tumor-reactive T cells. *Nat. Med.* **19**, 747–752 (2013).
86. B. M. Carreno, V. Magrini, M. Becker-Hapak, S. Kaabinejad, J. Hundal, A. A. Petti, A. Ly, W.-R. Lie, W. H. Hildebrand, E. R. Mardis, G. P. Linette, A dendritic cell vaccine increases the breadth and diversity of melanoma neoantigen-specific T cells. *Science* **348**, 803–808 (2015).
87. N. J. Llosa, M. Cruise, A. Tam, E. C. Wicks, E. M. Hechenbleikner, J. M. Taube, R. L. Blosser, H. Fan, H. Wang, B. S. Lubner, M. Zhang, N. Papadopoulos, K. W. Kinzler, B. Vogelstein, C. L. Sears, R. A. Anders, D. M. Pardoll, F. Housseau, The vigorous immune microenvironment of microsatellite instable colon cancer is balanced by multiple counter-inhibitory checkpoints. *Cancer Discov.* **5**, 43–51 (2015).
88. M. S. Rooney, S. A. Shukla, C. J. Wu, G. Getz, N. Hacohen, Molecular and genetic properties of tumors associated with local immune cytolytic activity. *Cell* **160**, 48–61 (2015).
89. M. M. Gubin, X. Zhang, H. Schuster, E. Caron, J. P. Ward, T. Noguchi, Y. Ivanova, J. Hundal, C. D. Arthur, W.-J. Kribber, G. E. Mulder, M. Toebes, M. D. Vesely, S. S. K. Lam, A. J. Korman, J. P. Allison, G. J. Freeman, A. H. Sharpe, E. L. Pearce, T. N. Schumacher, R. Aebbersold, H.-G. Rammensee, C. J. M. Melief, E. R. Mardis, W. E. Gillanders, M. N. Artyomov, R. D. Schreiber, Checkpoint blockade cancer immunotherapy targets tumour-specific mutant antigens. *Nature* **515**, 577–581 (2014).
90. L. B. Alexandrov, S. Nik-Zainal, D. C. Wedge, S. A. J. R. Aparicio, S. Behjati, A. V. Biankin, G. R. Bignell, N. Bolli, A. Borg, A.-L. Börresen-Dale, S. Boyault, B. Burkhardt, A. P. Butler, C. Caldas, H. R. Davies, C. Desmedt, R. Eils, J. E. Eyfjörð, J. A. Foekens, M. Greaves, F. Hosoda, B. Hutter, T. Ilicic, S. Imbeaud, M. Imielinski, N. Jäger, D. T. W. Jones, D. Jones, S. Knappskog, M. Kool, S. R. Lakhani, C. López-Otin, S. Martin, N. C. Munshi, H. Nakamura, P. A. Northcott, M. Pajic, E. Papaemmanuil, A. Paradiso, J. V. Pearson, X. S. Puente, K. Raine, M. Ramakrishna, A. L. Richardson, J. Richter, P. Rosenstiel, M. Schlesner, T. N. Schumacher, P. N. Span, J. W. Teague, Y. Totoki, A. N. J. Tutt, R. Valdés-Mas, M. M. van Buuren, L. van 't Veer, A. Vincent-Salomon, N. Waddell, L. R. Yates; Australian Pancreatic Cancer Genome Initiative;
- ICGC Breast Cancer Consortium; ICGC MML-Seq Consortium, ICGC PedBrain, J. Zucman-Rossi, P. A. Futreal, U. McDermott, P. Lichter, M. Meyerson, S. M. Grimmond, R. Siebert, E. Campo, T. Shibata, S. M. Pfister, P. J. Campbell, M. R. Stratton, Signatures of mutational processes in human cancer. *Nature* **500**, 415–421 (2013).
91. T. N. Schumacher, R. D. Schreiber, Neoantigens in cancer immunotherapy. *Science* **348**, 69–74 (2015).
92. I. Kryczek, M. Banerjee, P. Cheng, L. Vatan, W. Szeliga, S. Wei, E. Huang, E. Finlayson, D. Simeone, T. H. Welling, A. Chang, G. Coukos, R. Liu, W. Zou, Phenotype, distribution, generation, and functional and clinical relevance of Th17 cells in the human tumor environments. *Blood* **114**, 1141–1149 (2009).
93. S. Spranger, R. Bao, T. F. Gajewski, Melanoma-intrinsic β -catenin signalling prevents anti-tumour immunity. *Nature* **523**, 231–235 (2015).
94. D. Peng, I. Kryczek, N. Nagarsheth, L. Zhao, S. Wei, W. Wang, Y. Sun, E. Zhao, L. Vatan, W. Szeliga, J. Kotarski, R. Tarkowski, Y. Dou, K. Cho, S. Hensley-Alford, A. Munkarah, R. Liu, W. Zou, Epigenetic silencing of T_H1-type chemokines shapes tumour immunity and immunotherapy. *Nature* **527**, 249–253 (2015).
95. N. Nagarsheth, D. Peng, I. Kryczek, K. Wu, W. Li, E. Zhao, L. Zhao, S. Wei, T. Frankel, L. Vatan, W. Szeliga, Y. Dou, S. Owens, V. Marquez, K. Tao, E. Huang, G. Wang, W. Zou, PRC2 epigenetically silences Th1-type chemokines to suppress effector T cell trafficking in colon cancer. *Cancer Res.* **76**, 275–282 (2016).
96. W. Timp, A. P. Feinberg, Cancer as a dysregulated epigenome allowing cellular growth advantage at the expense of the host. *Nat. Rev. Cancer* **13**, 497–510 (2013).
97. P. C. Tumeq, C. L. Harview, J. H. Yearley, I. P. Shintaku, E. J. M. Taylor, L. Robert, B. Chmielowski, M. Spasic, G. Henry, V. Ciobanu, A. N. West, M. Carmona, C. Kivork, E. Seja, G. Cherry, A. J. Gutierrez, T. R. Grogan, C. Mateus, G. Tomasic, J. A. Glaspy, R. O. Emerson, H. Robins, R. H. Pierce, D. A. Elashoff, C. Robert, A. Ribas, PD-1 blockade induces responses by inhibiting adaptive immune resistance. *Nature* **515**, 568–571 (2014).
98. S. Viaud, F. Saccheri, G. Mignot, T. Yamazaki, R. Daillère, D. Hannani, D. P. Enot, C. Pfirschke, C. Engblom, M. J. Pittet, A. Schlitzer, F. Ginhoux, L. Apetoh, E. Chachaty, P.-L. Woerther, G. Eberl, M. Bérard, C. Ecobichon, D. Clermont, C. Bizet, V. Gaboriau-Routhiau, N. Cerf-Bensussan, P. Opolon, N. Yessaad, E. Vivier, B. Ryffel, C. O. Elson, J. Doré, G. Kroemer, P. Lepage, I. G. Boneca, F. Ghiringhelli, L. Zitvogel, The intestinal microbiota modulates the anticancer immune effects of cyclophosphamide. *Science* **342**, 971–976 (2013).
99. N. Iida, A. Dzutsev, C. A. Stewart, L. Smith, N. Bouladoux, R. A. Weingarten, D. A. Molina, R. Salcedo, T. Back, S. Cramer, R.-M. Dai, H. Kiu, M. Cardone, S. Naik, A. K. Patri, E. Wang, F. M. Marincola, K. M. Frank, Y. Belkaid, G. Trinchieri, R. S. Goldszmid, Commensal bacteria control cancer response to therapy by modulating the tumor microenvironment. *Science* **342**, 967–970 (2013).
100. A. Sivan, L. Corrales, N. Hubert, J. B. Williams, K. Aquino-Michaels, Z. M. Earley, F. W. Benyamin, Y. M. Lei, B. Jabri, M.-L. Alegre, E. B. Chang, T. F. Gajewski, Commensal *Bifidobacterium* promotes antitumor immunity and facilitates anti-PD-L1 efficacy. *Science* **350**, 1084–1089 (2015).
101. M. Vétizou, J. M. Pitt, R. Daillère, P. Lepage, N. Waldschmitt, C. Flament, S. Rusakiewicz, B. Routy, M. P. Roberti, C. P. M. Duong, V. Poirier-Colame, A. Roux, S. Becharef, S. Formenti, E. Golden, S. Cording, G. Eberl, A. Schlitzer, F. Ginhoux, S. Mani, T. Yamazaki, N. Jacquelot, D. P. Enot, M. Bérard, J. Nigou, P. Opolon, A. Eggemont, P.-L. Woerther, E. Chachaty, N. Chaput, C. Robert, C. Mateus, G. Kroemer, D. Raoult, I. G. Boneca, F. Carbonnel, M. Chamailard, L. Zitvogel, Anticancer immunotherapy by CTLA-4 blockade relies on the gut microbiota. *Science* **350**, 1079–1084 (2015).
102. V. Velcheti, K. A. Schalper, D. E. Carvajal, V. K. Anagnostou, K. N. Syrigos, M. Sznol, R. S. Herbst, S. N. Gettinger, L. Chen, D. L. Rimm, Programmed death ligand-1 expression in non-small cell lung cancer. *Lab. Invest.* **94**, 107–116 (2014).
103. J. Tan, X. Yang, L. Zhuang, X. Jiang, W. Chen, P. L. Lee, R. K. Karuturi, P. B. O. Tan, E. T. Liu, Q. Yu, Pharmacologic disruption of Polycomb-repressive complex 2-mediated gene repression selectively induces apoptosis in cancer cells. *Genes Dev.* **21**, 1050–1063 (2007).
104. M. T. McCabe, H. M. Ott, G. Ganji, S. Korenchuk, C. Thompson, G. S. Van Aller, Y. Liu, A. P. Graves, A. Della Pietra III, E. Diaz, L. V. LaFrance, M. Mellinger, C. Duquenne, X. Tian, R. G. Kruger, C. F. McHugh, M. Brandt, W. H. Miller, D. Dhanak, S. K. Verma, P. J. Tummino, C. L. Creasy, EZH2 inhibition as a therapeutic strategy for lymphoma with EZH2-activating mutations. *Nature* **492**, 108–112 (2012).
105. H. Li, K. B. Chiappinelli, A. A. Guzzetta, H. Easwaran, R.-W. C. Yen, R. Vatapalli, M. J. Topper, J. Luo, R. M. Connolly, N. S. Azad, V. Stearns, D. M. Pardoll, N. Davidson, P. A. Jones, D. J. Slamon, S. B. Baylin, C. A. Zahnow, N. Ahuja, Immune regulation by low doses of the DNA methyltransferase inhibitor 5-azacytidine in common human epithelial cancers. *Oncotarget* **5**, 587–598 (2014).
106. J. Wrangle, W. Wang, A. Koch, H. Easwaran, H. P. Mohammad, X. Pan, F. Vendetti, W. VanCrieke, T. DeMeyer, Z. Du, P. Parsana, K. Rodgers, R.-W. Yen, C. A. Zahnow, J. M. Taube, J. B. Brahmer, S. S. Tykodi, K. Easton, R. D. Carvajal, P. A. Jones, P. W. Laird, D. J. Weisenberger, S. Tsai, R. A. Juergens, S. L. Topalian, C. M. Rudin, M. V. Brock, D. Pardoll, S. B. Baylin, Alterations of immune response of non-small cell lung cancer with azacytidine. *Oncotarget* **4**, 2067–2079 (2013).

107. A. Woloszynska-Read, P. Mhaweche-Fauceglia, J. Yu, K. Odunsi, A. R. Karpf, Intertumor and intratumor NY-ESO-1 expression heterogeneity is associated with promoter-specific and global DNA methylation status in ovarian cancer. *Clin. Cancer Res.* **14**, 3283–3290 (2008).
108. L. Wang, Z. Amoozgar, J. Huang, M. H. Saleh, D. Xing, S. Orsulic, M. S. Goldberg, Decitabine enhances lymphocyte migration and function and synergizes with CTLA-4 blockade in a murine ovarian cancer model. *Cancer Immunol. Res.* **3**, 1030–1041 (2015).
109. A. Gros, P. F. Robbins, X. Yao, Y. F. Li, S. Turcotte, E. Tran, J. R. Wunderlich, A. Mixon, S. Farid, M. E. Dudley, K.-i. Hanada, J. R. Almeida, S. Darko, D. C. Douek, J. C. Yang, S. A. Rosenberg, PD-1 identifies the patient-specific CD8⁺ tumor-reactive repertoire infiltrating human tumors. *J. Clin. Invest.* **124**, 2246–2259 (2014).
110. J. D. Beane, G. Lee, Z. Zheng, M. Mendel, D. Abate-Daga, M. Bharathan, M. Black, N. Gandhi, Z. Yu, S. Chandran, M. Giedlin, D. Ando, J. Miller, D. Paschon, D. Guschin, E. J. Rebar, A. Reik, M. C. Holmes, P. D. Gregory, N. P. Restifo, S. A. Rosenberg, R. A. Morgan, S. A. Feldman, Clinical scale zinc finger nuclease-mediated gene editing of PD-1 in tumor infiltrating lymphocytes for the treatment of metastatic melanoma. *Mol. Ther.* **23**, 1380–1390 (2015).
111. K. M. Mahoney, P. D. Rennett, G. J. Freeman, Combination cancer immunotherapy and new immunomodulatory targets. *Nat. Rev. Drug Discov.* **14**, 561–584 (2015).
112. G. L. Beatty, D. A. Torigan, E. G. Chiorean, B. Saboury, A. Brothers, A. Alavi, A. B. Troxel, W. Sun, U. R. Teitelbaum, R. H. Vonderheide, P. J. O'Dwyer, A phase I study of an agonist CD40 monoclonal antibody (CP-870,893) in combination with gemcitabine in patients with advanced pancreatic ductal adenocarcinoma. *Clin. Cancer Res.* **19**, 6286–6295 (2013).
113. J. C. Byrd, T. J. Kipps, I. W. Flinn, M. Cooper, O. Odenike, J. Bendiske, J. Rediske, S. Bilic, J. Dey, J. Baeck, S. O'Brien, Phase I study of the anti-CD40 humanized monoclonal antibody lucatumumab (HCD122) in relapsed chronic lymphocytic leukemia. *Leuk. Lymphoma* **53**, 2136–2142 (2012).
114. P. Bansal-Pakala, A. G.-H. Jember, M. Croft, Signaling through OX40 (CD134) breaks peripheral T-cell tolerance. *Nat. Med.* **7**, 907–912 (2001).
115. A. D. Weinberg, N. P. Morris, M. Kovacsovics-Bankowski, W. J. Urba, B. D. Curti, Science gone translational: The OX40 agonist story. *Immunol. Rev.* **244**, 218–231 (2011).
116. I. Melero, W. W. Shuford, S. A. Newby, A. Aruffo, J. A. Ledbetter, K. E. Hellström, R. S. Mittler, L. Chen, Monoclonal antibodies against the 4-1BB T-cell activation molecule eradicate established tumors. *Nat. Med.* **3**, 682–685 (1997).
117. C.-H. Chang, J. Qiu, D. O'Sullivan, M. D. Buck, T. Noguchi, J. D. Curtis, Q. Chen, M. Gindin, M. M. Gubin, G. J. W. van der Windt, E. Tonc, R. D. Schreiber, E. J. Pearce, E. L. Pearce, Metabolic competition in the tumor microenvironment is a driver of cancer progression. *Cell* **162**, 1229–1241 (2015).
118. P.-C. Ho, J. D. Bihuniak, A. N. Macintyre, M. Staron, X. Liu, R. Amezcua, Y.-C. Tsui, G. Cui, G. Micevic, J. C. Perales, S. H. Kleinstein, E. D. Abel, K. L. Insogna, S. Feske, J. W. Locasale, M. W. Bosenberg, J. C. Rathmell, S. M. Kaech, Phosphoenolpyruvate is a metabolic checkpoint of anti-tumor T cell responses. *Cell* **162**, 1217–1228 (2015).
119. S. Wei, I. Kryczek, W. Zou, Regulatory T-cell compartmentalization and trafficking. *Blood* **108**, 426–431 (2006).
120. W. Zou, Regulatory T cells, tumour immunity and immunotherapy. *Nat. Rev. Immunol.* **6**, 295–307 (2006).
121. C. Ménétrier-Caux, T. Curiel, J. Faget, M. Manuel, C. Caux, W. Zou, Targeting regulatory T cells. *Target Oncol.* **7**, 15–28 (2012).
122. T. R. Simpson, F. Li, W. Montalvo-Ortiz, M. A. Sepulveda, K. Bergerhoff, F. Arce, C. Roddie, J. Y. Henry, H. Yagita, J. D. Wolchok, K. S. Peggs, J. V. Ravetch, J. P. Allison, S. A. Quezada, Fc-dependent depletion of tumor-infiltrating regulatory T cells co-defines the efficacy of anti-CTLA-4 therapy against melanoma. *J. Exp. Med.* **210**, 1695–1710 (2013).
123. F. S. Hodi, S. J. O'Day, D. F. McDermott, R. W. Weber, J. A. Sosman, J. B. Haanen, R. Gonzalez, C. Robert, D. Schadendorf, J. C. Hassel, W. Akerley, A. J. M. van den Eertwegh, J. Lutzky, P. Lorigan, J. M. Vaubel, G. P. Linette, D. Hogg, C. H. Ottensmeier, C. Lebbé, C. Peschel, I. Quidt, J. I. Clark, J. D. Wolchok, J. S. Weber, J. Tian, M. J. Yellin, G. M. Nichol, A. Hoos, W. J. Urba, Improved survival with ipilimumab in patients with metastatic melanoma. *N. Engl. J. Med.* **363**, 711–723 (2010).
124. D. Schadendorf, F. S. Hodi, C. Robert, J. S. Weber, K. Margolin, O. Hamid, D. Patt, T.-T. Chen, D. M. Berman, J. D. Wolchok, Pooled analysis of long-term survival data from phase II and phase III trials of ipilimumab in unresectable or metastatic melanoma. *J. Clin. Oncol.* **33**, 1889–1894 (2015).
125. S. Wei, A. B. Shreiner, N. Takeshita, L. Chen, W. Zou, A. E. Chang, Tumor-induced immune suppression of *in vivo* effector T-cell priming is mediated by the B7-H1/PD-1 axis and transforming growth factor β . *Cancer Res.* **68**, 5432–5438 (2008).
126. M. Ogura, T. Ishida, K. Hatake, M. Taniwaki, K. Ando, K. Tobinai, K. Fujimoto, K. Yamamoto, T. Miyamoto, N. Uike, M. Tanimoto, K. Tsukasaki, K. Ishizawa, J. Suzumiya, H. Inagaki, K. Tamura, S. Akinaga, M. Tomonaga, R. Ueda, Multicenter phase II study of mogamulizumab (KW-0761), a defucosylated anti-CC chemokine receptor 4 antibody, in patients with relapsed peripheral T-cell lymphoma and cutaneous T-cell lymphoma. *J. Clin. Oncol.* **32**, 1157–1163 (2014).
127. B. Allard, S. Pommey, M. J. Smyth, J. Stagg, Targeting CD73 enhances the antitumor activity of anti-PD-1 and anti-CTLA-4 mAbs. *Clin. Cancer Res.* **19**, 5626–5635 (2013).
128. S. Wan, E. Zhao, I. Kryczek, L. Vatan, A. Sadovskaya, G. Ludema, D. M. Simeone, W. Zou, T. H. Welling, Tumor-associated macrophages produce interleukin 6 and signal via STAT3 to promote expansion of human hepatocellular carcinoma stem cells. *Gastroenterology* **147**, 1393–1404 (2014).
129. A. L. Mellor, D. H. Munn, IDO expression by dendritic cells: Tolerance and tryptophan catabolism. *Nat. Rev. Immunol.* **4**, 762–774 (2004).
130. T. Maj, S. Wei, T. Welling, W. Zou, T cells and costimulation in cancer. *Cancer J.* **19**, 473–482 (2013).
131. I. Kryczek, S. Wei, G. Zhu, L. Myers, P. Mottram, P. Cheng, L. Chen, G. Coukos, W. Zou, Relationship between B7-H4, regulatory T cells, and patient outcome in human ovarian carcinoma. *Cancer Res.* **67**, 8900–8905 (2007).
132. T.-W. Sun, Q. Gao, S.-J. Qiu, J. Zhou, X.-Y. Wang, Y. Yi, J.-Y. Shi, Y.-F. Xu, Y.-H. Shi, K. Song, Y.-S. Xiao, J. Fan, B7-H3 is expressed in human hepatocellular carcinoma and is associated with tumor aggressiveness and postoperative recurrence. *Cancer Immunol. Immunother.* **61**, 2171–2182 (2012).
133. I. Yamato, M. Sho, T. Nomi, T. Akahori, K. Shimada, K. Hotta, H. Kanehiro, N. Konishi, H. Yagita, Y. Nakajima, Clinical importance of B7-H3 expression in human pancreatic cancer. *Br. J. Cancer* **101**, 1709–1716 (2009).
134. L. Luo, A. I. Chapoval, D. B. Flies, G. Zhu, F. Hirano, S. Wang, J. S. Lau, H. Dong, K. Tamada, A. S. Flies, Y. Liu, L. Chen, B7-H3 enhances tumor immunity *in vivo* by costimulating rapid clonal expansion of antigen-specific CD8⁺ cytolytic T cells. *J. Immunol.* **173**, 5445–5450 (2004).
135. R. Rahbar, A. Lin, M. Ghazarian, H.-L. Yau, S. Paramathas, P. A. Lang, A. Schildknecht, A. R. Elford, C. Garcia-Batres, B. Martin, H. K. Berman, W. L. Leong, D. R. McCready, M. Reedijk, S. J. Done, N. Miller, B. Youngson, W.-K. Suh, T. W. Mak, P. S. Ohashi, B7-H4 expression by nonhematopoietic cells in the tumor microenvironment promotes antitumor immunity. *Cancer Immunol. Res.* **3**, 184–195 (2015).
136. L. Wang, R. Rubinstein, J. L. Lines, A. Wasiuk, C. Ahonen, Y. Guo, L.-F. Lu, D. Gondek, Y. Wang, R. A. Fava, A. Fiser, S. Almo, R. J. Noelle, VISTA, a novel mouse Ig superfamily ligand that negatively regulates T cell responses. *J. Exp. Med.* **208**, 577–592 (2011).
137. D. B. Flies, X. Han, T. Higuchi, L. Zheng, J. Sun, J. J. Ye, L. Chen, Coinhibitory receptor PD-1H preferentially suppresses CD4⁺ T cell-mediated immunity. *J. Clin. Invest.* **124**, 1966–1975 (2014).
138. D. B. Flies, T. Higuchi, L. Chen, Mechanistic assessment of PD-1H coinhibitory receptor-induced T cell tolerance to allogeneic antigens. *J. Immunol.* **194**, 5294–5304 (2015).
139. K. Sakuishi, L. Apetoh, J. M. Sullivan, B. R. Blazar, V. K. Kuchroo, A. C. Anderson, Targeting Tim-3 and PD-1 pathways to reverse T cell exhaustion and restore anti-tumor immunity. *J. Exp. Med.* **207**, 2187–2194 (2010).
140. H. Li, K. Wu, K. Tao, L. Chen, Q. Zheng, X. Lu, J. Liu, L. Shi, C. Liu, G. Wang, W. Zou, Tim-3/galectin-9 signaling pathway mediates T-cell dysfunction and predicts poor prognosis in patients with hepatitis B virus-associated hepatocellular carcinoma. *Hepatology* **56**, 1342–1351 (2012).
141. R. J. Johnston, L. Comps-Agrar, J. Hackney, X. Yu, M. Huseni, Y. Yang, S. Park, V. Javinal, H. Chiu, B. Irving, D. L. Eaton, J. L. Grogan, The immunoreceptor TIGIT regulates antitumor and antiviral CD8⁺ T cell effector function. *Cancer Cell* **26**, 923–937 (2014).
142. J.-M. Chauvin, O. Pagliano, J. Fourcade, Z. Sun, H. Wang, C. Sander, J. M. Kirkwood, T.-h. T. Chen, M. Maurer, A. J. Korman, H. M. Zarour, TIGIT and PD-1 impair tumor antigen-specific CD8⁺ T cells in melanoma patients. *J. Clin. Invest.* **125**, 2046–2058 (2015).
143. S.-R. Woo, M. E. Turnis, M. V. Goldberg, J. Bankoti, M. Selby, C. J. Nirschl, M. L. Bettini, D. M. Gravano, P. Vogel, C. L. Liu, S. Tangsombatvisit, J. F. Grosso, G. Netto, M. P. Smeltzer, A. Chau, P. J. Utz, C. J. Workman, D. M. Pardoll, A. J. Korman, C. G. Drake, D. A. A. Vignali, Immune inhibitory molecules LAG-3 and PD-1 synergistically regulate T-cell function to promote tumoral immune escape. *Cancer Res.* **72**, 917–927 (2012).
144. S. A. Rosenberg, J. C. Yang, N. P. Restifo, Cancer immunotherapy: Moving beyond current vaccines. *Nat. Med.* **10**, 909–915 (2004).
145. S. Pilon-Thomas, A. Mackay, N. Vohra, J. J. Mulé, Blockade of programmed death ligand 1 enhances the therapeutic efficacy of combination immunotherapy against melanoma. *J. Immunol.* **184**, 3442–3449 (2010).
146. J. Duraiswamy, G. J. Freeman, G. Coukos, Therapeutic PD-1 pathway blockade augments with other modalities of immunotherapy T-cell function to prevent immune decline in ovarian cancer. *Cancer Res.* **73**, 6900–6912 (2013).
147. L. Karyampudi, P. Lamichhane, A. D. Scheid, K. R. Kalli, B. Shreeder, J. W. Krempski, M. D. Behrens, K. L. Knutson, Accumulation of memory precursor CD8 T cells in regressing tumors following combination therapy with vaccine and anti-PD-1 antibody. *Cancer Res.* **74**, 2974–2985 (2014).
148. S. A. Rosenberg, Decade in review—Cancer immunotherapy: Entering the mainstream of cancer treatment. *Nat. Rev. Clin. Oncol.* **11**, 630–632 (2014).
149. R. H. I. Andtbacka, H. L. Kaufman, F. Collichio, T. Amatruda, N. Senzer, J. Chesney, K. A. Delman, L. E. Spitzer, I. Puzanov, S. S. Agarwala, M. Milhem, L. Cranmer, B. Curti, K. Lewis, M. Ross, T. Guthrie, G. P. Linette, G. A. Daniels, K. Harrington, M. R. Middleton, W. H. Miller Jr, J. S. Zager, Y. Ye, B. Yao, A. Li, S. Doleman, A. VanderWalde, J. Gansert, R. S. Coffin, Talimogene laherparepvec improves durable response rate in patients with advanced melanoma. *J. Clin. Oncol.* **33**, 2780–2788 (2015).

150. M. Nakanishi, D. W. Rosenberg, Multifaceted roles of PGE₂ in inflammation and cancer. *Semin. Immunopathol.* **35**, 123–137 (2013).
151. Y. Li, M. Fang, J. Zhang, J. Wang, Y. Song, J. Shi, W. Li, G. Wu, J. Ren, Z. Wang, W. Zou, L. Wang, Hydrogel dual delivered celecoxib and anti-PD-1 synergistically improve antitumor immunity. *Oncotarget* 10.1080/2162402X.2015.1074374 (2015).
152. S. Zelenay, A. G. van der Veer, J. P. Böttcher, K. J. Snellgrove, N. Rogers, S. E. Acton, P. Chakravarty, M. R. Girotti, R. Marais, S. A. Quezada, E. Sahai, C. Reis e Sousa, Cyclooxygenase-dependent tumor Growth through evasion of immunity. *Cell* **162**, 1257–1270 (2015).
153. M. Cheng, Y. Chen, W. Xiao, R. Sun, Z. Tian, NK cell-based immunotherapy for malignant diseases. *Cell. Mol. Immunol.* **10**, 230–252 (2013).
154. W. Zou, V. Machelon, A. Coulomb-L'Hermin, J. Borvak, F. Nome, T. Isaeva, S. Wei, R. Krzysiek, I. Durand-Gasselain, A. Gordon, T. Pustilnik, D. T. Curiel, P. Galanaud, F. Capron, D. Emilie, T. J. Curiel, Stromal-derived factor-1 in human tumors recruits and alters the function of plasmacytoid precursor dendritic cells. *Nat. Med.* **7**, 1339–1346 (2001).
155. S. Wei, I. Kryczek, L. Zou, B. Daniel, P. Cheng, P. Mottram, T. Curiel, A. Lange, W. Zou, Plasmacytoid dendritic cells induce CD8⁺ regulatory T cells in human ovarian carcinoma. *Cancer Res.* **65**, 5020–5026 (2005).
156. A. Sistigu, T. Yamazaki, E. Vacchelli, K. Chaba, D. P. Enot, J. Adam, I. Vitale, A. Goubar, E. E. Baracco, C. Remédios, L. Fend, D. Hannani, L. Aymeric, Y. Ma, M. Niso-Santano, O. Kepp, J. L. Schultze, T. Tüting, F. Belardelli, L. Bracci, V. La Sorsa, G. Ziccheddu, P. Sestili, F. Urbani, M. Delorenzi, M. Lacroix-Triki, V. Quidville, R. Conforti, J.-P. Spano, L. Pusztai, V. Poirier-Colame, S. Delalogue, F. Penault-Llorca, S. Ladoire, L. Arnould, J. Cyra, M.-C. Dessoliers, A. Eggermont, M. E. Bianchi, M. Pittet, C. Engblom, C. Pfirschke, X. Prévigne, G. Uzè, R. D. Schreiber, M. T. Chow, M. J. Smyth, E. Proietti, F. André, G. Kroemer, L. Zitvogel, Cancer cell–autonomous contribution of type I interferon signaling to the efficacy of chemotherapy. *Nat. Med.* **20**, 1301–1309 (2014).
157. S. Viaud, C. Flament, M. Zoubir, P. Pautier, A. LeCesne, V. Ribrag, J.-C. Soria, V. Marty, P. Vielh, C. Robert, N. Chaput, L. Zitvogel, Cyclophosphamide induces differentiation of Th17 cells in cancer patients. *Cancer Res.* **71**, 661–665 (2011).
158. Y. Lee, S. L. Auh, Y. Wang, B. Burnette, Y. Wang, Y. Meng, M. Beckett, R. Sharma, R. Chin, T. Tu, R. R. Weichselbaum, Y.-X. Fu, Therapeutic effects of ablative radiation on local tumor require CD8⁺ T cells: Changing strategies for cancer treatment. *Blood* **114**, 589–595 (2009).
159. L. Deng, H. Liang, B. Burnette, M. Beckett, T. Darga, R. R. Weichselbaum, Y.-X. Fu, Irradiation and anti-PD-L1 treatment synergistically promote antitumor immunity in mice. *J. Clin. Invest.* **124**, 687–695 (2014).
160. S. Park, Z. Jiang, E. D. Mortenson, L. Deng, O. Radkevich-Brown, X. Yang, H. Sattar, Y. Wang, N. K. Brown, M. Greene, Y. Liu, J. Tang, S. Wang, Y.-X. Fu, The therapeutic effect of anti-HER2/neu antibody depends on both innate and adaptive immunity. *Cancer Cell* **18**, 160–170 (2010).
161. A. Beano, E. Signorino, A. Evangelista, D. Brusa, M. Mistrangelo, M. A. Polimeni, R. Spadi, M. Donadio, L. Ciuffreda, L. Matera, Correlation between NK function and response to trastuzumab in metastatic breast cancer patients. *J. Transl. Med.* **6**, 25 (2008).
162. S. Wei, M. U. Egenti, S. Teitz-Tennenbaum, W. Zou, A. E. Chang, Effects of tumor irradiation on host T-regulatory cells and systemic immunity in the context of adoptive T-cell therapy in mice. *J. Immunother.* **36**, 124–132 (2013).
163. C. Twyman-Saint Victor, A. J. Rech, A. Maity, R. Rengan, K. E. Pauken, E. Stelekati, J. L. Benci, B. Xu, H. Dada, P. M. Odorizzi, R. S. Herati, K. D. Mansfield, D. Patsch, R. K. Amaravadi, L. M. Schuchter, H. Ishwaran, R. Mick, D. A. Pryma, X. Xu, M. D. Feldman, T. C. Gangadhar, S. M. Hahn, E. J. Wherry, R. H. Vonderheide, A. J. Minn, Radiation and dual checkpoint blockade activate non-redundant immune mechanisms in cancer. *Nature* **520**, 373–377 (2015).
164. F. Klug, H. Prakash, P. E. Huber, T. Seibel, N. Bender, N. Halama, C. Pfirschke, R. H. Voss, C. Timke, L. Umansky, K. Klapproth, K. Schäkel, N. Garbi, D. Jäger, J. Weitz, H. Schmitz-Winnenthal, G. J. Hämmerling, P. Beckhove, Low-dose irradiation programs macrophage differentiation to an iNOS⁺/M1 phenotype that orchestrates effective T cell immunotherapy. *Cancer Cell* **24**, 589–602 (2013).
165. S. J. Dovedi, A. L. Adlard, G. Lipowska-Bhalla, C. McKenna, S. Jones, E. J. Cheadle, I. J. Stratford, E. Poon, M. Morrow, R. Stewart, H. Jones, R. W. Wilkinson, J. Honeychurch, T. M. Illidge, Acquired resistance to fractionated radiotherapy can be overcome by concurrent PD-L1 blockade. *Cancer Res.* **74**, 5458–5468 (2014).
166. A. B. Sharabi, C. J. Nirschl, C. M. Kochel, T. R. Nirschl, B. J. Francica, E. Velarde, T. L. Dewese, C. G. Drake, Stereotactic radiation therapy augments antigen-specific PD-1–mediated antitumor immune responses via cross-presentation of tumor antigen. *Cancer Immunol. Res.* **3**, 345–355 (2015).
167. L. Zitvogel, O. Kepp, G. Kroemer, Decoding cell death signals in inflammation and immunity. *Cell* **140**, 798–804 (2010).
168. S. Shalappour, J. Font-Burgada, G. Di Caro, Z. Zhong, E. Sanchez-Lopez, D. Dhar, G. Willmsky, M. Ammirante, A. Strasser, D. E. Hansel, C. Jamieson, C. J. Kane, T. Klatte, P. Birner, L. Kenner, M. Karin, Immunosuppressive plasma cells impede T-cell-dependent immunogenic chemotherapy. *Nature* **521**, 94–98 (2015).
169. J. Ozao-Choy, G. Ma, J. Kao, G. X. Wang, M. Meseck, M. Sung, M. Schwartz, C. M. Divino, P.-Y. Pan, S.-H. Chen, The novel role of tyrosine kinase inhibitor in the reversal of immune suppression and modulation of tumor microenvironment for immune-based cancer therapies. *Cancer Res.* **69**, 2514–2522 (2009).
170. C. J. Scotton, J. L. Wilson, D. Milliken, G. Stamp, F. R. Balkwill, Epithelial cancer cell migration: A role for chemokine receptors? *Cancer Res.* **61**, 4961–4965 (2001).
171. I. Kryczek, A. Lange, P. Mottram, X. Alvarez, P. Cheng, M. Hogan, L. Moons, S. Wei, L. Zou, V. Machelon, D. Emilie, M. Terrassa, A. Lackner, T. J. Curiel, P. Carmeliet, W. Zou, CXCL12 and vascular endothelial growth factor synergistically induce neoangiogenesis in human ovarian cancers. *Cancer Res.* **65**, 465–472 (2005).
172. C. Feig, J. O. Jones, M. Kraman, R. J. B. Wells, A. Deonaraine, D. S. Chan, C. M. Connell, E. W. Roberts, Q. Zhao, O. L. Caballero, S. A. Teichmann, T. Janowitz, D. I. Jodrell, D. A. Tuveson, D. T. Fearon, Targeting CXCL12 from FAP-expressing carcinoma-associated fibroblasts synergizes with anti-PD-L1 immunotherapy in pancreatic cancer. *Proc. Natl. Acad. Sci. U.S.A.* **110**, 20212–20217 (2013).
173. T. K. Choueiri, D. J. Figueroa, A. P. Fay, S. Signoretti, Y. Liu, R. Gagnon, K. Deen, C. Carpenter, P. Benson, T. H. Ho, L. Pandite, P. de Souza, T. Powles, R. J. Motzer, Correlation of PD-L1 tumor expression and treatment outcomes in patients with renal cell carcinoma receiving sunitinib or pazopanib: Results from COMPARE, a randomized controlled trial. *Clin. Cancer Res.* **21**, 1071–1077 (2015).
174. I. Kryczek, K. Wu, E. Zhao, S. Wei, L. Vatan, W. Szeliga, E. Huang, J. Greenson, A. Chang, J. Roliński, P. Radwan, J. Fang, G. Wang, W. Zou, IL-17⁺ regulatory T cells in the micro-environments of chronic inflammation and cancer. *J. Immunol.* **186**, 4388–4395 (2011).
175. P. Sansone, G. Storci, S. Tavolari, T. Guarnieri, C. Giovannini, M. Taffurelli, C. Ceccarelli, D. Santini, P. Paterini, K. B. Marcu, P. Chieco, M. Bonafè, IL-6 triggers malignant features in mammospheres from human ductal breast carcinoma and normal mammary gland. *J. Clin. Invest.* **117**, 3988–4002 (2007).
176. L. L. C. Marotta, V. Almendro, A. Marusyk, M. Shipitsin, J. Schemme, S. R. Walker, N. Bloushtain-Qimron, J. J. Kim, S. A. Choudhury, R. Maruyama, Z. Wu, M. Gönen, L. A. Mulvey, M. O. Bessarabova, S. J. Huh, S. J. Silver, S. Y. Kim, S. Y. Park, H. E. Lee, K. S. Anderson, A. L. Richardson, T. Nikolskaya, Y. Nikolsky, X. S. Liu, D. E. Root, W. C. Hahn, D. A. Frank, K. Polyak, The JAK2/STAT3 signaling pathway is required for growth of CD44+CD24⁻ stem cell-like breast cancer cells in human tumors. *J. Clin. Invest.* **121**, 2723–2735 (2011).
177. C. Ginestier, S. Liu, M. E. Diebel, H. Korkaya, M. Luo, M. Brown, J. Wicinski, O. Cabaud, E. Charafe-Jauffret, D. Birnbaum, J.-L. Guan, G. Dontu, M. S. Wicha, CXCR1 blockade selectively targets human breast cancer stem cells in vitro and in xenografts. *J. Clin. Invest.* **120**, 485–497 (2010).

Acknowledgments: We would like to thank our former and current collaborators and trainees for their intellectual input and hard work. PD-L1 and PD-1 (PD) pathway blockade is a highly promising therapy and has elicited durable antitumor responses and long-term remissions in a subset of patients with a broad spectrum of cancers. The review largely focuses on the human cancer immune microenvironment and cancer patient-oriented studies. Because of the plethora of literature related to the topic described in this review, it makes a complete and extensive review extremely challenging. We apologize in advance for any inadvertent omission. **Funding:** The work described in this Review was supported by grants from the U.S. NIH (CA193136, CA190176, CA171306, CA152470, CA099985, CA156685, CA123088, CA133620, CA092562, and CA100227) to W.Z.; grants P30 CA008748 and R01 CA056821, the Ludwig Trust, the Lloyd Charitable Trust, and the CRI/SU2C Immunotherapy Dream Team to J.D.W.; grants CA121974, CA177444, and CA016359 to L.C.; the Ovarian Cancer Research Fund and the U.S. Department of Defense to W.Z.; and a Stand Up to Cancer fund to L.C. **Competing interests:** W.Z. is a consultant to NGM and Lycera and has sponsored research grants from MedImmune and Lycera. J.D.W. is a consultant to Bristol-Myers Squibb, Merck, Genentech, and MedImmune. L.C. is the scientific founder of NextCure Inc. and receives loyalty payment from Bristol-Myers Squibb, Ventana, and ImmuNext. He is a consultant to Pfizer, MedImmune, and NextCure and has sponsored research grants from Boehringer Ingelheim, Pfizer, and NextCure.

Submitted 23 October 2015
Accepted 20 January 2016
Published 2 March 2016
10.1126/scitranslmed.aad7118

Citation: W. Zou, J. D. Wolchok, L. Chen, PD-L1 (B7-H1) and PD-1 pathway blockade for cancer therapy: Mechanisms, response biomarkers, and combinations. *Sci. Transl. Med.* **8**, 328rv4 (2016).

PD-L1 (B7-H1) and PD-1 pathway blockade for cancer therapy: Mechanisms, response biomarkers, and combinations

Weiping Zou, Jedd D. Wolchok and Lieping Chen

Sci Transl Med 8, 328rv4328rv4.
DOI: 10.1126/scitranslmed.aad7118

ARTICLE TOOLS

<http://stm.sciencemag.org/content/8/328/328rv4>

RELATED CONTENT

<http://stm.sciencemag.org/content/scitransmed/3/111/111ra120.full>
<http://stm.sciencemag.org/content/scitransmed/6/237/237ra67.full>
<http://science.sciencemag.org/content/sci/352/6282/227.full>
<http://science.sciencemag.org/content/sci/351/6280/1463.full>
<http://stm.sciencemag.org/content/scitransmed/8/343/343fs10.full>
<http://science.sciencemag.org/content/sci/354/6316/1104.full>
<http://science.sciencemag.org/content/sci/354/6316/1165.full>
<http://science.sciencemag.org/content/sci/354/6316/1160.full>
<http://stm.sciencemag.org/content/scitransmed/8/369/369ra177.full>
<http://stm.sciencemag.org/content/scitransmed/8/370/370ra180.full>
<http://stm.sciencemag.org/content/scitransmed/9/379/eaah3560.full>
<http://science.sciencemag.org/content/sci/355/6332/1373.full>
<http://science.sciencemag.org/content/sci/355/6332/1428.full>
<http://science.sciencemag.org/content/sci/355/6332/1423.full>
<http://stm.sciencemag.org/content/scitransmed/9/385/eaak9670.full>
<http://stm.sciencemag.org/content/scitransmed/9/385/eaak9679.full>
<http://science.sciencemag.org/content/sci/356/6334/200.full>
<http://science.sciencemag.org/content/sci/358/6363/573.full>
<http://science.sciencemag.org/content/sci/358/6365/852.full>
<http://science.sciencemag.org/content/sci/359/6371/91.full>
<http://science.sciencemag.org/content/sci/359/6371/97.full>
<http://stke.sciencemag.org/content/sigtrans/9/419/ec58.abstract>
<http://stke.sciencemag.org/content/sigtrans/10/494/eaak9702.full>
<http://science.sciencemag.org/content/sci/359/6371/32.full>
<http://science.sciencemag.org/content/sci/359/6371/104.full>
<http://science.sciencemag.org/content/sci/353/6297/399.full>
<http://stm.sciencemag.org/content/scitransmed/10/429/eaam7729.full>
<http://stke.sciencemag.org/content/sigtrans/11/541/eaao4874.full>
<http://science.sciencemag.org/content/sci/362/6410/13.full>
<http://science.sciencemag.org/content/sci/362/6411/eaar3593.full>
<http://stm.sciencemag.org/content/scitransmed/11/478/eaav4810.full>
<http://science.sciencemag.org/content/sci/364/6439/485.full>
<http://science.sciencemag.org/content/sci/364/6440/558.full>
<http://science.sciencemag.org/content/sci/369/6510/1427.full>
<http://science.sciencemag.org/content/sci/369/6510/1481.full>

Use of this article is subject to the [Terms of Service](#)

Science Translational Medicine (ISSN 1946-6242) is published by the American Association for the Advancement of Science, 1200 New York Avenue NW, Washington, DC 20005. The title *Science Translational Medicine* is a registered trademark of AAAS.

Copyright © 2016, American Association for the Advancement of Science

REFERENCES

This article cites 177 articles, 77 of which you can access for free
<http://stm.sciencemag.org/content/8/328/328rv4#BIBL>

PERMISSIONS

<http://www.sciencemag.org/help/reprints-and-permissions>

Use of this article is subject to the [Terms of Service](#)

Science Translational Medicine (ISSN 1946-6242) is published by the American Association for the Advancement of Science, 1200 New York Avenue NW, Washington, DC 20005. The title *Science Translational Medicine* is a registered trademark of AAAS.

Copyright © 2016, American Association for the Advancement of Science