

HIV

Sources of HIV infection among men having sex with men and implications for prevention

Oliver Ratmann,^{1*} Ard van Sighem,² Daniela Bezemer,² Alexandra Gavryushkina,³ Suzanne Jurriaans,⁴ Annemarie Wensing,⁵ Frank de Wolf,¹ Peter Reiss,^{2,6} Christophe Fraser,¹ ATHENA observational cohort

New HIV diagnoses among men having sex with men (MSM) have not decreased appreciably in most countries, even though care and prevention services have been scaled up substantially in the past 20 years. To maximize the impact of prevention strategies, it is crucial to quantify the sources of transmission at the population level. We used viral sequence and clinical patient data from one of Europe's nationwide cohort studies to estimate probable sources of transmission for 617 recently infected MSM. Seventy-one percent of transmissions were from undiagnosed men, 6% from men who had initiated antiretroviral therapy (ART), 1% from men with no contact to care for at least 18 months, and 43% from those in their first year of infection. The lack of substantial reductions in incidence among Dutch MSM is not a result of ineffective ART provision or inadequate retention in care. In counterfactual modeling scenarios, 19% of these past cases could have been averted with current annual testing coverage and immediate ART to those testing positive. Sixty-six percent of these cases could have been averted with available antiretrovirals (immediate ART provided to all MSM testing positive, and preexposure antiretroviral prophylaxis taken by half of all who test negative for HIV), but only if half of all men at risk of transmission had tested annually. With increasing sequence coverage, molecular epidemiological analyses can be a key tool to direct HIV prevention strategies to the predominant sources of infection, and help send HIV epidemics among MSM into a decisive decline.

INTRODUCTION

Combination antiretroviral therapy (ART) transformed HIV from a deadly to a lifelong disease and is also one of the most effective strategies for preventing onward infections (1, 2). However, among men having sex with men (MSM), the substantial scale-up of ART in the past 20 years has not resulted in appreciable reductions of new HIV infections and diagnoses (Table 1) (3). Building on successful behavioral and biomedical HIV prevention strategies (4), further interventions exist that could be used to reduce the number of HIV infections among MSM. The 2016 World Health Organization (WHO) guidelines now recommend ART initiation regardless of CD4 cell count after diagnosis (immediate ART), as well as provision of antiretrovirals as preexposure prophylaxis (PrEP) to those at substantial risk of infection (5). Future prevention programs could focus on one or both recommended interventions, as well as on increased routine HIV testing and diagnosis (6); RNA testing to detect MSM with acute infection, at which time they are thought to be most infectious (7); and improved adherence and linkage support to assist patients with attaining and sustaining undetectable viral loads while on ART (8). The potential impact of any of these interventions, and specifically those recommended by the WHO, relies crucially on how many HIV transmissions originate from different stages in the entire HIV infection and care continuum, ranging from undiagnosed acute infection through treated infection and loss to follow-up. This has been challenging to measure directly through classical epidemiological approaches.

Here, we use the viral phylogenetic relationship between partial HIV-1 subtype B polymerase sequences to reconstruct past, probable transmission events in the Netherlands (Fig. 1). These sequences were routinely collected for drug resistance testing of HIV-infected patients who are in care (9). Among sampled MSM, 94% were of subtype B. Then, we use clinical records to determine the staging of probable transmission events within the infection and care continuum (Fig. 2A and Table 2). This enabled us to estimate the population-level proportion of transmissions among the reconstructed transmission events that are attributable to the 14 stages of the infection and care continuum in Fig. 2A. Transmissions could be attributed to stages before diagnosis because HIV sequences, always collected after diagnosis, diverge fast enough to indicate past transmission events (10). Similarly, transmissions could also be attributed to men with no contact to care for at least 18 months. Finally, using these estimates, we quantified the potential impact of currently not implemented prevention programs in the Dutch MSM population, had they been used in the last 3 years. In particular, we evaluate if the revised 2016 WHO guidelines on immediate ART and PrEP could have substantially altered the course of the Dutch HIV epidemic among MSM.

Understanding which interventions should be prioritized for the Dutch MSM epidemic is an important case study. First, the number of new MSM infections in the Netherlands has not decreased appreciably (9) despite comprehensive linkage and retention in care, substantial ART scale-up free of charge, and frequent follow-up to maintain viral control of the vast majority of those on ART (Table 1). Second, similar epidemic trends are reported from other countries with an overall equally comprehensive cascade of care (Table 1), casting more general doubts on the population-level impact of current prevention strategies targeting MSM epidemics (11). Third, nearly all HIV-infected MSM in care are enrolled in the nationwide Aids Therapy Evaluation in the Netherlands cohort (ATHENA) since early 1996 (9). HIV care is monitored comprehensively at high frequency (clinic visits, treatment histories, comorbidities

¹Department of Infectious Disease Epidemiology, School of Public Health, Imperial College London, London W21PG, UK. ²Stichting HIV Monitoring, 1105 BD Amsterdam, the Netherlands. ³Department of Computer Science, University of Auckland, Auckland 1142, New Zealand. ⁴Department of Medical Microbiology, Academic Medical Center, 1105 AZ Amsterdam, the Netherlands. ⁵Department of Medical Microbiology, University Medical Center Utrecht, 3584 CX Utrecht, the Netherlands. ⁶Department of Global Health, Academic Medical Center, 1105 BM Amsterdam, the Netherlands.

*Corresponding author. E-mail: oliver.ratmann@imperial.ac.uk

recorded; ~3 viral loads/CD4 measurements per year per individual) (9), which allowed us to characterize phylogenetically reconstructed transmission events in detail.

RESULTS

Potential transmissions to MSM that were confirmed to have recent infection at time of diagnosis

By 2013, 11,863 HIV-infected MSM were registered and still in care in the Netherlands. To estimate their sources of transmission and then the impact of prevention programs, we focused on transmissions to MSM that were recently infected at time of diagnosis (stage A in Fig. 1). Between July 1996 and December 2010, 1794 MSM were confirmed to have been infected at most 12 months before diagnosis. Types of evidence were a previous negative HIV test (76%), laboratory diagnosis (7%), or clinical diagnosis of acute infection (17%). For 1045 (58%) of these, a sequence was available. To these recipient MSM, we considered as potential transmitters all HIV-infected men whose course of infection overlapped with the infection window of the recipient (stage A in Fig. 1). Using this approach, we could resolve the timing and direction of potential transmission events (12). Of all 12,207 potential transmitters, 5593 (46%) had a viral sequence and formed ~4.4 million potential transmission pairs with sequences available for both individuals (stage B in Fig. 1).

Phylogenetically probable transmission events

Genetic sequences of the virus alone cannot prove epidemiological linkage (13). However, most of the potential transmission pairs could be ruled out as implausible based on the phylogenetic relationship of the viral sequences. The viral phylogeny among the Dutch sequences and their closest matches in the Los Alamos HIV sequence database (www.hiv.lanl.gov/) were reconstructed with maximum-likelihood methods, and reliable subtrees were identified (see Material and Methods). Potential transmitters whose sequences did not occur in the same reliable subtree as those of the recipient MSM were excluded (stage C in Fig. 1) (10), as were potential transmitters whose sequences were incompatible with a direct HIV transmission event (stage D in Fig. 1) (14). Direct transmission could be excluded in 99.96% of all potential transmission pairs. We identified 903 phylogenetically probable transmitters to 617 recipient MSM in 2343 pairs. Our analyses are based on this open observational cohort of past, phylogenetically reconstructed transmission events.

To guide and interpret this exclusion analysis, we evaluated patterns of viral divergence between sequences isolated from epidemiologically confirmed transmission pairs (14, 15) and pairings of Dutch MSM that could not have infected each other (see Material and Methods). On the basis of these pairs, the above exclusion criteria were highly specific (true transmitters to recipients are not excluded, >90%), whereas sensitivity was low (incorrect transmission

Table 1. HIV incidence trends and care for infected MSM in the Netherlands and other countries.

Country	Uninfected MSM testing annually		Diagnosed MSM receiving ART			Treated MSM with suppressed viral load			MSM retained in care		HIV incidence among MSM	
	Year	%	Year	%	Median CD4 count at ART initiation (cells/ml)	Year	%	Viral load threshold (cps/ml)	Year	%		Trend
The Netherlands	2003	??	2003	79	202	2003	80	<100	2003	92	2003	Increasing*
	2013	38.4 [†]	2013	90	382	2013	91	<100	2012	95	2013	Stable*
Australia	2013	61.1 [‡]	2013	75 [‡]	379 ^{§¶}	2013	88 ^{¶¶}	<50	2013	96 ^{§¶}	2013	Stable to increasing**
British Columbia	2009	51 ^{††}	2014	85 ^{‡‡}	411 ^{‡‡}	2014	84 ^{‡‡}	<50	2011	86 ^{¶¶§§}	2013	Stable ^{¶¶}
Switzerland	2010	39.3 [‡]	2014	86 ^{¶¶}	402 ^{¶¶}	2012	96 ^{¶¶¶¶}	<200	2012	97 ^{¶¶†††}	2014	Decreasing new diagnoses ^{¶¶¶}
UK	2010	36.4 [‡]	2013	86 ^{§§§}	420 ^{¶¶¶}	2013	91 ^{¶¶}	<200	2013	95 ^{¶¶}	2013	Stable ^{¶¶¶¶}

*From (47). †From The EMIS Network. EMIS 2010: The European Men-Who-Have-Sex-With-Men Internet Survey. Findings from 38 countries. Stockholm: European Centre for Disease Prevention and Control, 2013. ‡From Gay Community Periodic Surveys, https://kirby.unsw.edu.au/projects/gay-community-periodic-surveys, reported in HIV, hepatitis, and sexually transmissible infections in Australia Annual surveillance report 2014. §From Australian HIV Observational Database Annual Report 2014, reporting care indicators in a closed observational cohort. ¶Estimate not specific to MSM. ¶¶From the Australian HIV Observational Database, reported in HIV, hepatitis, and sexually transmissible infections in Australia Annual surveillance report 2014. ¶¶¶From fact sheet HIV and AIDS in Australia, 20th International AIDS conference. ¶¶¶¶From Mancount, prospective cross-sectional survey in Vancouver, www.mancount.ca/files/ManCount_Report2010.pdf. ¶¶¶¶¶From HIV monitoring quarterly report for British Columbia, fourth quarter 2014. ¶¶¶¶¶From B. Nosyk, J. S. Montaner, G. Colley, V. D. Lima, K. Chan, K. Heath, B. Yip, H. Samji, M. Gilbert, R. Barrios, R. Gustafson, R. S. Hogg, The cascade of HIV care in British Columbia, Canada, 1996–2011: A population-based retrospective cohort study. *Lancet Infect. Dis.* **14**, 40–49 (2014). ¶¶¶¶¶¶From www.phac-aspc.gc.ca/aids-sida/publication/epi/2010/index-eng.php. ¶¶¶¶¶¶¶2621 of 3081 MSM on ART and registered in the Swiss HIV Cohort Study, personal communication with the Datacenter of the Swiss HIV Cohort Study. ¶¶¶¶¶¶¶¶From P. Kohler, A. J. Schmidt, B. Ledergerber, P. Vernazza, CROI2015, www.croi-conference.org/sites/default/files/posters-2015/1008.pdf. ¶¶¶¶¶¶¶¶¶From www.shcs.ch/155-shcs-key-data-figures, update June 2014. ¶¶¶¶¶¶¶¶¶From HIV- und STI-Fallzahlen 2014: Berichterstattung, Analysen und Trends, in comparison with numbers for 2008 in the 2012 report, www.bag.admin.ch/hiv_aids/12472/12480/12481/12484/index.html?lang=de. ¶¶¶¶¶¶¶¶¶¶From www.gov.uk/government/statistics/hiv-data-tables. ¶¶¶¶¶¶¶¶¶¶¶Within 9 months before ART initiation, personal communication PHE. ¶¶¶¶¶¶¶¶¶¶¶From HIV in the UK: 2014 Report. ¶¶¶¶¶¶¶¶¶¶¶¶From P. J. Birrell, O. N. Gill, V. C. Delpech, A. E. Brown, S. Desai, T. R. Chadborn, B. D. Rice, D. De Angelis, HIV incidence in men who have sex with men in England and Wales 2001–10: A nationwide population study. *Lancet Infect. Dis.* **13**, 313–318 (2013).

Downloaded from http://stm.sciencemag.org/ by guest on September 19, 2019

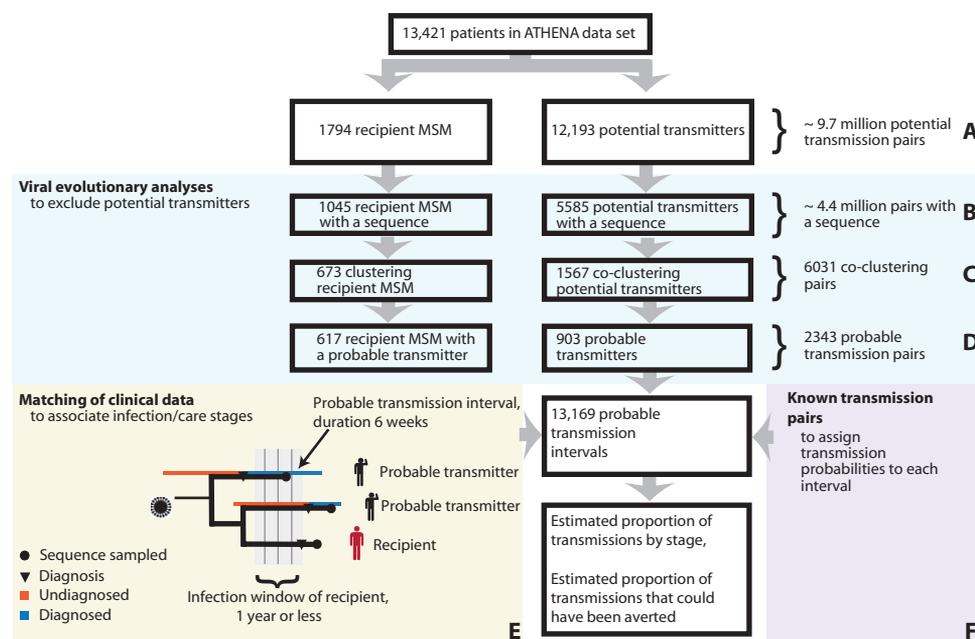


Fig. 1. Study design. Nationwide sources of transmission were identified of MSM with evidence of recent infection in the first year before diagnosis (recipient MSM). **(A)** Out of all patients in the ATHENA cohort, men whose course of infection overlapped with the infection window were considered as potential transmitters. **(B)** Only those pairs with sequences from both individuals were considered for further analysis. **(C and D)** With viral phylogenetic analyses, the vast majority of pairs could be ruled out. All remaining pairs were considered phylogenetically probable. **(E)** On the basis of detailed clinical records, probable transmission events were characterized by stage in the HIV infection and care continuum. Because transmitters progressed in stage over time, we considered time-resolved transmission intervals. **(F)** Independent viral phylogenetic data from epidemiologically confirmed pairs were used to determine the phylogenetic probability of direct transmission during each interval. Statistical analyses were adjusted for extensive sampling and censoring biases.

pairs could not always be excluded, ~60%). This indicates that the actual transmitter is almost certainly among the phylogenetically reconstructed, probable transmitters, provided he was sequenced. From the known sequence coverage alone, we expected that about half of all 1045 recipient MSM with a sequence had their actual transmitter sampled, further suggesting that the actual transmitter is among the phylogenetically reconstructed, probable transmitters for the large majority of the reconstructed 617 transmission events.

The clinical and demographic characteristics of the selected 617 recipient MSM were typical of all 1794 MSM that were confirmed to have recent infection at time of diagnosis (Table 3).

Characterization of individual transmission events by stage in the HIV infection and care continuum

Using clinical records, we then enumerated all stages in the HIV infection and care continuum during which the 617 transmission events could have occurred. Probable transmitters progressed in stage over time and overlapped with infection windows in 13,169 time-resolved, 6-week-long transmission intervals (Fig. 2B). Censoring and sequence sampling biases were identified for each stage by comparing men with and without a sequence, and were adjusted in line with previous work (16). Reflecting targeted sequence collection, intervals were not missing at random (Fig. 2C and fig. S9). Each interval

was associated with a phylogenetic transmission probability on the basis of the genetic distance between sequences from the transmitter and recipient and the time elapsed since the putative transmission interval and the sampling dates of both individuals (see Materials and Methods and fig. S10). For each recipient, the probability that transmission occurred from one of the 14 stages then depends on the number of his probable transmitters in that stage and the transmission probabilities associated with each of the corresponding transmission intervals (see Materials and Methods).

Sources of HIV transmission

The population-level proportions of HIV transmissions attributable to the 14 infection/care stages were obtained by summing individual-level transmission probabilities by stage across all recipients and are shown in Table 4. Figure 3 compares the proportion of transmissions from each stage to the population-level proportion of infected men in these stages. Between July 1996 and December 2010, an estimated 71% (66 to 73%) of all 617 transmission events originated from undiagnosed men, 22% (21 to 26%) from diagnosed but not yet treated men, 6% (5 to 8%) from men who initiated ART, and 1%

(0.7 to 1.6%) from men with no contact to care for at least 18 months. An estimated 43% (37 to 46%) of the 617 recipient MSM were infected by men undergoing their first year of infection.

Impact of prevention strategies

Figure 4 describes the counterfactual prevention scenarios for which we calculated the proportion of transmissions in the cohort that could have been averted between mid-2008 to December 2010, had we intervened to redistribute the identified, probable transmitters to less infectious infection/care stages. Young MSM are at particularly high risk of infection (17, 18). We therefore considered—along the revised 2016 WHO guidelines (5)—rollout of immediate ART to all infected MSM and PrEP to half of all MSM age 30 years or younger who test negative: at most, 30% (22 to 39%) of infections could have been averted without increased annual testing. Immediate ART alone could have averted 19% (13 to 26%) of these cases at current testing levels. In practice, low adherence is associated with decreasing effectiveness of PrEP (19). We assumed an 86% efficacy of PrEP as reported in the recent Ipergay and PROUD trials (20, 21). Figure S12 reports the impact of lower efficacy values. Figure S13 reports the impact of lower or higher PrEP coverage. Next, we considered increased annual testing. Only 17% of identified probable transmitters had a last negative test in the year before diagnosis compared to 27% of diagnosed MSM between mid-2008 and December 2010 and 38% of uninfected MSM in 2013 (Table 1). If half of all transmitters

Table 2. Stages in the HIV infection and care continuum.

Infection/care stage of transmitter	Definition
Undiagnosed	Transmission intervals whose midpoint is before diagnosis:
Confirmed to have recent infection at diagnosis	All transmission intervals of transmitters that were confirmed to have recent infection at time of diagnosis.
Estimated to have recent infection	Considering transmitters that had no evidence of recent infection at time of diagnosis, all transmission intervals whose midpoint is less than 12 months after the estimated infection date.
Estimated to have chronic infection	Considering transmitters that had no evidence of recent infection at time of diagnosis, all transmission intervals whose midpoint is more than 12 months after the estimated infection date.
Diagnosed	Transmission intervals whose midpoint is after diagnosis and before ART start (only of transmitters that are in contact with care services):
Diagnosed <3 months, confirmed recent infection at diagnosis	Considering transmitters who had confirmed recent infection at time of diagnosis, all transmission intervals whose midpoint is within the first 3 months after diagnosis.
No CD4 measured	No available CD4 count since diagnosis up to the midpoint of the interval.
CD4 >500	CD4 counts remained above 500 cells/ml between the first CD4 count and the midpoint of the interval.
CD4 350–500	CD4 counts decreased to 350–500 cells/ml between the first CD4 count and the midpoint of the interval.
CD4 <350	CD4 counts decreased to below 350 cells/ml between the first CD4 count and the midpoint of the interval.
ART initiated	Transmission intervals whose midpoint is after ART start (only of transmitters that are in contact with care services):
Before first viral suppression	No first viral load measurement below 100 copies/ml in any transmission interval of the transmitter after ART start
After first viral suppression*	
No viral load measured*	No viral load measurement in any transmission interval of the transmitter after ART start
No viral suppression*	At least one viral load measurement at or above 100 copies/ml in any transmission interval of the transmitter after ART start
Viral suppression, one observation*	One viral load measurement in any transmission interval of the transmitter after ART start, which is below 100 copies/ml.
Viral suppression, ≥ 2 observations*	Several viral load measurements in any transmission interval of the transmitter after ART start, all of which are below 100 copies/ml.
Not in contact	No patient record (last contact, clinic visit, CD4 measurement, viral load measurement) in the past and future 9 months from the midpoint of the transmission interval.

*Although flow through the stages is typically unidirectional, men could move freely between these stages.

had tested annually, immediate ART and PrEP to half of all MSM age 30 years or younger who test negative could have averted 45% (34 to 56%) of infections. Comprehensive rollout of PrEP to half of all men testing negative irrespective of their age would have substantially boosted the combination intervention: 66% (50 to 78%) of infections could have been averted.

DISCUSSION

HIV epidemics among MSM have, unlike other settings (22), not declined appreciably with substantial improvements to care and ART scale-up (Table 1). We characterized 617 past transmission events

among MSM in the Netherlands based on phylogenetic and clinical data, estimated their sources throughout the infection and care continuum, and quantified the impact that biomedical prevention programs could have had in averting the reconstructed transmission events. Analyzing this transmission cohort, we aim to inform the design of future prevention interventions beyond high levels of ART coverage and the numerous successful behavioral interventions that are already in place (9).

A potential limitation of this study is that transmitters to MSM with recent infection at diagnosis may differ from typical transmitters. On average, fewer men diagnosed late with a CD4 count below 350 cells/ml occurred in phylogenetic transmission clusters with a recipient MSM compared to those without (fig. S23). This may imply that, overall,

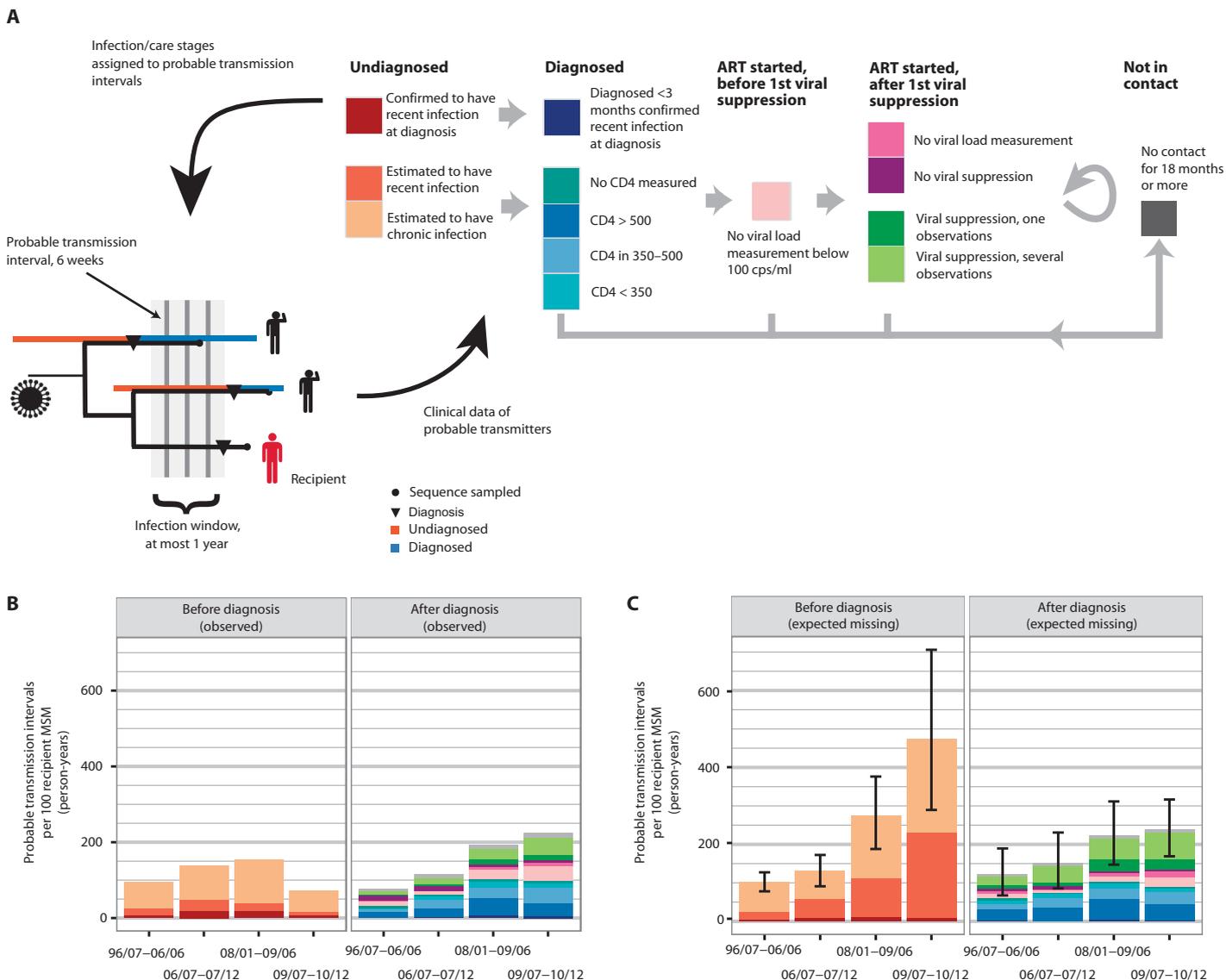


Fig. 2. Phylogenetically probable transmission intervals, linked to stages in the infection and care continuum. (A) Left: Each recipient could have been infected during his infection window from multiple probable transmitters. For each transmitter, the transmission window was split into 6-week-long probable transmission intervals. Infection/care stages were assigned to these intervals on the basis of clinical data to reflect progression of the transmitters through the infection/care continuum. Right: Relationship between the 14 infection/care stages as defined in Table 2. Transmitters progress unidirectionally, except for stages after first viral suppression, or when individuals reenter care (as indicated by arrows). (B) For each stage, the total number of observed transmission intervals to recipient MSM during their infection windows is shown by date of diagnosis of the recipients.

the proportion of transmissions from undiagnosed men with chronic infection is higher and, consequently, that the impact that immediate ART could have had is lower than our estimates. Conversely, the impact of increased annual testing and PrEP could be larger than reported, if men diagnosed late are not more difficult to reach than the average transmitter in our cohort. Further, this study focuses on the sources and

Overall, the number of probable transmission intervals per recipient increases with time, reflecting the increasing number of infected men in care. Transmitters are increasingly less likely to have been diagnosed by 2013, resulting in a decreasing number of undiagnosed transmission intervals toward the present. (C) In addition to censoring, diagnosed transmitters may not have a sequence sampled. Comparing men with and without a sequence in the near-complete population cohort, we could adjust for these biases. The total number of expected missing transmission intervals to recipients is shown, along with 95% bootstrap confidence intervals. Observed and expected missing transmission intervals were associated with phylogenetic transmission probabilities, which sum to 1 per recipient.

prevention of in-country transmissions: 97% of the recipient MSM reported that infection was likely acquired in the Netherlands, compared to 86% of diagnosed MSM. The contribution of cross-border transmissions may increase as the response is strengthened (23), an effect that we did not consider. Phylogenetic uncertainty and the phylogenetic exclusion criteria had little impact on our findings (figs. S14 to S22). A further

Table 3. Characteristics of the recipient MSM with identified sources of transmission. IQR, interquartile range.

Characteristic	Recipient MSM with a phylogenetically probable transmitter (n = 617)	Recipient MSM with or without a sequence (n = 1794)	Diagnosed MSM (n = 7978)
Evidence for infection in the past year			
Previous negative test in the past year (%)	77	76	17
Laboratory diagnosis (%)	8	7	2
Clinical diagnosis of acute infection (%)	15	17	4
Age at diagnosis (years; mean and IQR)	36.8 (29.5–42.9)	37.2 (29.9–43.5)	38.7 (31.3–45.1)
First CD4 count within 12 months of diagnosis and before ART start (cells/ml; mean and IQR)	505 (350–630)	534 (360–670)	402 (200–560)
Viral load count within 12 months of diagnosis (log ₁₀ RNA; mean and IQR)	4.9 (4.4–5.5)	4.8 (4.3–5.4)	4.7 (4.3–5.3)
In care in the Amsterdam metropolitan area (%)	45.1	43.5	43.6
Last negative test within 12 months before diagnosis (%)	77.0	76.1	17.1
Self-reported in country infection* (%)	96.9	91.9	88.5

*Of those self-reporting a country of origin.

Table 4. Proportion of transmissions by stage in the HIV infection and care continuum.

Infection/care stage of transmitter	% of transmissions by time of diagnosis of recipient MSM (95% confidence interval)				
	Overall (n = 617)	Jul. 1996 to Apr. 2006 (n = 165)	May 2006 to Dec. 2007 (n = 145)	Jan. 2008 to Jun. 2009 (n = 151)	Jul. 2009 to Dec. 2010 (n = 156)
Undiagnosed (total)	70.9 (65.8–72.5)	67.6 (59.3–72.7)	72.3 (64.2–76.9)	71.8 (63.4–76.3)	72.2 (63.3–76.3)
Confirmed recent infection at diagnosis	15.5 (11.9–17.4)	15 (7.6–19.4)	21.7 (15–26.5)	16.4 (11–20.8)	9.4 (5.6–14.1)
Estimated to have recent infection	25.1 (19.4–28.1)	17.3 (11.7–22.7)	23 (15.1–30.1)	25.9 (15.4–33.6)	34.6 (19.4–43.4)
Estimated to have chronic infection	30.3 (28–34)	35.2 (30.2–42)	27.6 (22.4–34)	29.5 (24.2–36.1)	28.2 (23–35.7)
Diagnosed (total)	22.4 (20.7–26.2)	23.6 (18.5–29.7)	22.9 (18.6–29.1)	22.8 (18.3–29.4)	20.7 (17.4–27.3)
Diagnosed <3 months, recent infection at diagnosis	2.9 (2.2–4.1)	2.5 (1–4.9)	3.2 (1.7–5.5)	3 (1.9–5.4)	2.8 (1.8–4.4)
No CD4 measured	1.6 (1.2–2.4)	2.9 (1.6–4.8)	0.8 (0.4–1.8)	1.5 (0.6–3)	1 (0.6–2.1)
CD4 >500	8.3 (7–10.3)	10.2 (6.7–14.2)	7 (4.5–10.8)	8.7 (5.9–12.5)	7.1 (5.4–10.1)
CD4 in 350–500	6.4 (5.4–7.9)	4.8 (2.6–7.8)	7.3 (5.1–10.5)	5.9 (4.2–8.3)	7.7 (5.7–11)
CD4 <350	3.4 (2.5–4.3)	3.2 (1.2–5.5)	4.6 (2.6–6.6)	3.7 (2.2–5.6)	2.1 (1.3–3.3)
ART initiated (total)	5.7 (5.2–7.8)	7 (4.8–11.7)	3.7 (2.2–6.5)	4.9 (3.7–8.1)	6.7 (5.4–10.2)
Before first viral suppression	1.8 (1.6–2.7)	2.2 (1.2–4.4)	0.7 (0.4–1.5)	1.3 (0.9–2.6)	2.8 (2.1–4.6)
After first viral suppression					
No viral load measured	0.5 (0.3–1)	0.9 (0.1–2.4)	0.1 (0–0.3)	0.3 (0.1–0.9)	0.8 (0.4–1.8)
No viral suppression	1.4 (0.9–2.1)	2.8 (1.2–5.2)	1.2 (0.4–2.6)	0.9 (0.4–1.9)	0.5 (0.1–1)
Viral suppression, one observation	0.4 (0.3–0.8)	0.1 (0–0.5)	0.2 (0–0.8)	0.5 (0.2–1.7)	0.6 (0.3–1.5)
Viral suppression, ≥2 observations	1.6 (1.1–2.5)	1 (0.1–2.6)	1.5 (0.6–3.1)	1.9 (0.9–3.6)	2 (1.1–3.6)
Not in contact	1 (0.7–1.6)	1.8 (0.8–3.4)	1.1 (0.4–2.3)	0.5 (0.2–1.4)	0.4 (0.2–0.8)
Recent infection (total)	43.5 (36.6–46)	34.9 (25.4–40.6)	47.9 (36.9–54.8)	45.3 (33.3–54.1)	47.7 (32.8–53.8)

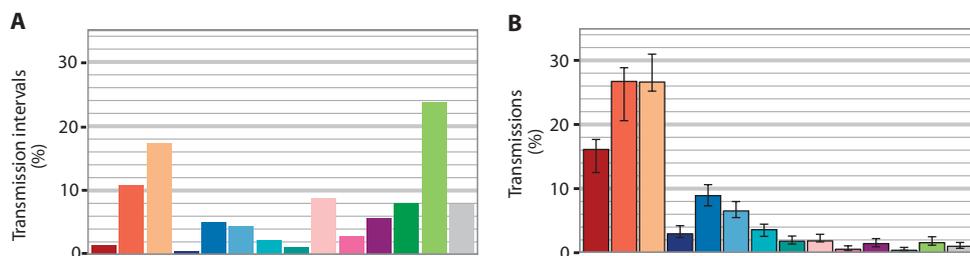


Fig. 3. Proportion of transmissions by stage in the infection and care continuum versus proportion of these stages among infected men. (A) Relative frequency of infection/care stages in the population, among potential transmitters that overlap with the infection windows of recipient MSM and could have, in principle, transmitted to one of the recipient MSM (stage A in Fig. 1; color codes as in Fig. 2). (B) Proportion of the 617 transmission events attributable to each infection/care stage (bar: 95% bootstrap confidence interval).

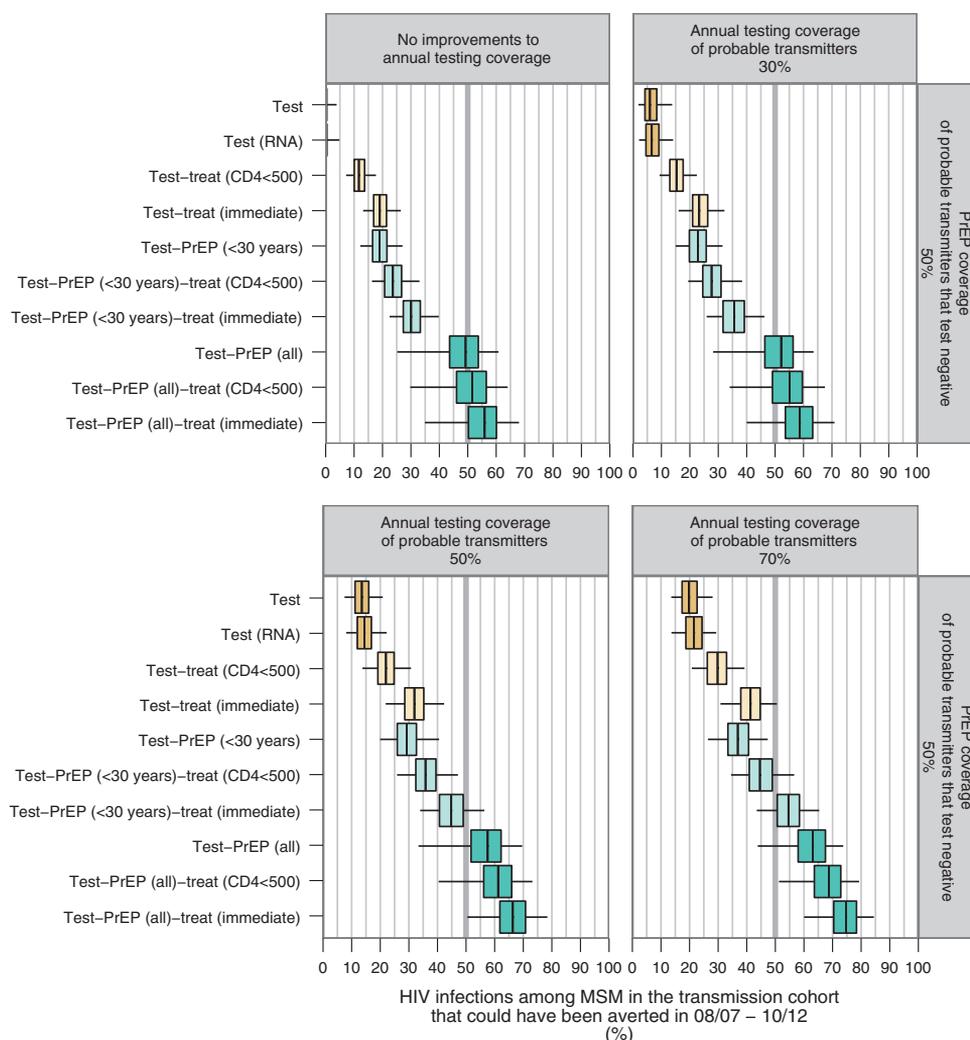


Fig. 4. Impact of biomedical interventions among MSM in the Netherlands. Estimated proportion of transmissions that could have been averted in the period July 2008 to December 2010 if the corresponding additional prevention strategies had been implemented by July 2008 (line, median; box, bootstrap interquartile range; whiskers, 95% bootstrap confidence interval). Scenarios were varied by annual testing coverage of phylogenetically identified, probable transmitters. Current testing coverage was 17%, corresponding to the proportion of probable transmitters that had a negative test in the 12 months before diagnosis.

potential caveat to the robustness of our findings is that only half of all potential transmitters had a viral sequence sampled. Although population-level sampling biases were adjusted, we must acknowledge that the actual transmitter may not have been sampled for all recipients. Improving sequence sampling coverage at time of diagnosis is needed to facilitate phylogenetic prevention analyses (24).

The identified sources of transmission imply, first, that viral suppression induced by ART is highly effective in preventing transmissions in this population (Fig. 3). The relative risk of HIV transmission from men after ART initiation varies by stage but is always estimated well below 1 when compared to diagnosed, untreated men with a CD4 count above 500 cells/ml, and is in particular 0.04 (0.02 to 0.1) for men with viral suppression (fig. S11).

Second, very few transmissions are attributable to temporary or permanent loss to follow-up, which must be considered in the context of high linkage and retention to care in the Netherlands: few diagnosed MSM had subsequently no contact to care for at least 18 months (8.2%) and most reentered care within 5 years (69%) (9). In contrast, several studies indicate that more than half of all transmissions among MSM in the United States originate from men that were not retained in care (25–27). The estimated impact of particular prevention strategies in Fig. 4 is limited to settings with a similar epidemic profile and care cascade as the Netherlands (Table 1).

Third, not more than an estimated 20% of infections in the cohort could have been averted between mid-2008 and December 2010 with immediate ART after diagnosis. Given the remarkable expansion of ART coverage in the Netherlands in the past (9), the prevention potential of immediate ART is now limited. Nonetheless, starting ART at a cell count above 500 cells/ml leads to improved clinical outcomes and remains a priority (28).

Fourth, and similar to other locations (24, 29), almost half of all infections in our transmission cohort originated from men in their first year of infection. Frequent early transmission limits the overall impact of annual

testing plus immediate ART to those testing positive (Fig. 4), thus implying that prevention services to uninfected MSM must be strengthened. The substantial, estimated impact that PrEP would have had in averting transmissions in our cohort (Fig. 4) supports making PrEP available to MSM testing negative as in the United States (30). Recent PrEP demonstration projects (31, 32) indicate that existing barriers such as low awareness (33) and a lack of experience among providers (34) can be addressed. Concerns regarding the toxicity of PrEP, increasing sexual risk behavior, and emerging drug resistance have to date not been substantiated since PrEP was made available in the United States (35). In the context of PrEP-experienced prevention services, high discontinuation rates after PrEP initiation appear to be the greatest challenge to maintain protection from infection (31).

Fifth, without substantial increases in current annual testing coverage, ART and PrEP offered along the revised 2016 WHO guidelines could not have prevented more than a third of all infections in our transmission cohort. Because phylogenetically probable transmitters tend to test much less frequently than the average diagnosed MSM, substantial barriers likely exist in reaching men at high risk of onward transmission, and further work is needed to characterize these (36). Strategies such as self-testing (37), community-based testing (38), and more provider-initiated routine testing in general practices and at medical admissions raised annual testing coverage in pilot projects (39) and need to be expanded alongside biomedical interventions.

Sixth, this study indicates that substantial reductions in HIV incidence among MSM could have been realized with a combination approach that includes—critically—increased annual testing, with uptake of PrEP by young MSM testing negative and provision of immediate ART to those testing positive. This finding is primarily based on the impact of increased annual testing and the higher efficacy of PrEP reported in two recent randomized controlled trials (20, 21), and updates previous studies that estimate more limited benefits (4, 40, 41). Beyond age at testing, other characteristics not available to this study may also indicate high infection risk (42) and thereby identify groups of MSM to which PrEP should be made available as a priority too. Provision of PrEP to all men testing negative is not affordable at current drug prices in high-income countries (40). The magnitude of the predicted impact of test-and-PrEP-and-treat for all (Fig. 4) could set an aspirational target for the fight against HIV among MSM.

The lack of substantial reductions in incidence among Dutch MSM is not a result of ineffective ART provision or inadequate retention in care. New HIV infections among MSM are challenging to prevent because of frequent early transmission and continued low testing uptake of men at risk of transmission. Counterfactual prevention scenarios on phylogenetically reconstructed, past transmission events to MSM with recent infection at diagnosis predict that increased annual testing and uptake of PrEP by men at high risk of infection have a key role to send the HIV epidemic among MSM into a decisive decline.

MATERIALS AND METHODS

Study design

We conducted a retrospective viral phylogenetic transmission and prevention study that focuses on transmissions to MSM that were confirmed to have recent HIV infection at time of diagnosis in the Netherlands (Fig. 1). The prespecified objectives were to, first, reconstruct past, phylogeneti-

cally probable transmission events to these recipient MSM; second, to estimate the proportion of transmissions originating throughout the infection and care continuum based on the reconstructed transmission events; and, third, to estimate the proportion of infections that could have been averted through reallocating past, probable transmitters to less infectious stages in counterfactual modeling scenarios.

The ATHENA national observational HIV cohort includes anonymized data of all HIV-infected patients followed longitudinally in the 27 HIV treatment centers in the Netherlands since 1996, except 1.5% who opt-out (9). ATHENA patients are informed of data collection by their treating physician and can refuse further collection of clinical data according to an opt-out procedure. Patients who were diagnosed between 1981 and 1995 were included in the cohort when they were still alive in 1996 (9). Demographic, clinical, and viral sequence data were collected at entry and follow-up visits as described previously (9). By March 2013, viral sequence data had been systematically entered until December 2010. Therefore, recipients were enrolled between early 1996 and December 2010. Potential transmitters were enrolled until database closure in March 2013. Table S1 characterizes the demographic, clinical, and viral sequence data that were used in this study. The resolution of the infection/care stages in Table 2 was adjusted to ensure adequate sample sizes. The number of probable transmission intervals after first viral suppression was too small to enable further stratification by treatment class. This study was reviewed and approved by the HIV Monitoring Institutional Data Access and Ethics Committee, and reported along STROME-ID guidelines.

Viral sequences of different subtypes ($n = 355$ from MSM), with less than 250 nucleotides ($n = 368$) or indication for intra-subtype recombination ($n = 52$), were removed before analysis. Primary drug resistance mutations were masked in each sequence (43). Demographic and clinical data were checked for consistency along patient timelines and to lie within appropriate ranges. Outliers were reported to the ATHENA quality control team and manually updated.

Recently infected, recipient MSM and infection windows

We enrolled as recipients all MSM for whom a narrow infection window could be identified. MSM had evidence of infection within 12 months before diagnosis if either a last negative HIV-1 antibody test in the 12 months preceding diagnosis, an indeterminate HIV-1 Western blot, or clinical diagnosis of acute infection was reported. Figure S1 shows enrollment progress over time. Infection windows were at most 12 months or shorter if indicated by a last negative HIV antibody test (fig. S2).

Potential transmitters to recipient MSM

We enrolled as potential transmitters all registered infected men that overlapped with infection windows of recipients and thus could have in principle infected a recipient. This definition required estimation of putative infection times. Calculations are based on a method by Rice and colleagues (44) (see the Supplementary Materials). Estimated infection times are associated with substantial uncertainty, and sensitivity analyses were conducted for lower and upper 95% estimates. Table S2 characterizes the potential transmitters to all recipients. Further analysis was restricted to potential transmission pairs with sequences from both individuals (stage B in Fig. 1).

Viral phylogenetic exclusion analysis to construct the transmission cohort

The viral phylogeny was reconstructed under the GTR nucleotide substitution model with maximum-likelihood methods (45) and is

shown in fig. S3. Five hundred bootstrap trees were created to quantify uncertainty in tree reconstruction (10). Genetic distances between sequences from transmitter-recipient pairs were highly variable (fig. S4), which was accounted for in all analyses. To guide our choice of exclusion criteria, we considered, first, epidemiologically confirmed transmission pairs from previously published transmission chains in Belgium and Sweden (15, 46). The Belgium transmission chain was subsequently oversampled (14), providing 2807 sequence pairs from confirmed transmitters and recipients without multidrug resistance. Further, we considered 4117 pairs of sequences from the same Dutch patient and 201,605 pairs between Dutch patients who died before the last negative antibody test of another patient. These pairs were used to quantify patterns of viral evolutionary diversification that can be expected among confirmed linked and unlinked pairs, and to develop exclusion criteria with high specificity (see the Supplementary Materials). The Swedish pairs were used for validation purposes. All potential transmitters that were not excluded were considered phylogenetically probable and are characterized in table S4.

Relative pairwise transmission probabilities

Among the 2807 confirmed transmission pairs (14), the genetic distance between sequences from the transmitter and the recipient was strongly associated with the time elapsed between both sampling dates and the midpoint of the established infection window (fig. S5). We fitted a probabilistic molecular clock model to these data to describe the relative probability of observing a given genetic distance between sequences from a transmission pair that diverged for a specified amount of time from each other. The fitted model was then used to express the relative probability that a phylogenetically identified transmitter was the actual transmitter to a recipient (fig. S5).

Matching of clinical data to associate infection/care stages with transmission intervals

Sources of transmission were not defined in terms of individuals but by the 14 stages in the infection and care continuum in Table 2 (stage E in Fig. 1). Stages were allocated to transmission intervals on the basis of available clinical data (table S1). The duration of transmission intervals was set to 6 weeks to accommodate abrupt changes in infection/care stages.

Adjusting for censoring and sequence sampling biases

Toward the present, an increasing fraction of potential transmitters may not have been diagnosed by the time of database closure. Potential transmitters with recent infection at time of diagnosis must, by definition, have been diagnosed within 12 months after the putative transmission interval. Therefore, the extent of right censoring differs between stages. To adjust for right censoring, we counted when potential transmitters in a particular infection/care stage became diagnosed in relation to the time of diagnosis of their recipient (fig. S6). This enabled us to estimate the proportion of censored intervals for a hypothetical database closure time in the past (fig. S6). We then extrapolated these estimates to the actual database closure time with a bootstrap algorithm (see the Supplementary Materials). To quantify sequence sampling biases, we compared men with and without a sequence in the near-complete population cohort (fig. S7). A negative binomial missing data model was then used to adjust for the number of missing transmission intervals. Adjustments accounted for censoring; increasing sampling frequency with duration in care; high sampling frequency of men returning to care, men participating in particular substudies, and men

with indication of drug resistance; as well as increasing sampling frequency with calendar time (fig. S7).

Epidemiological transmission analysis

Each interval was associated with a phylogenetic transmission probability (stage F in Fig. 1). The relative pairwise transmission probabilities (fig. S5) were equally apportioned to all observed intervals of the same transmitter-recipient pair. Stage-specific data such as viral load were not used to determine these probabilities to avoid circularity in the attribution of transmissions to infection/care stages. Then, the transmission probability in an observed interval τ from transmitter i to recipient j was calculated by

$$p_{ij\tau} = \omega_{ij\tau} / \left(\sum_{k,s} \omega_{kjs} + \sum_z m_j(z) \omega(z) \right),$$

where $\omega_{ij\tau}$ is the relative transmission probability in interval τ , and the denominator sums over all observed, competing intervals as well as expected missing intervals $m_j(z)$ in stage z to recipient j . For missing intervals, relative transmission probabilities were imputed and set to the median ω_{ijs} of all observed intervals s in stage z , denoted by $\omega(z)$. For a missing transmission interval v in stage x to recipient j , we calculated

$$p_{jv} = \omega^{(x)} / \left(\sum_{k,s} \omega_{kjs} + \sum_z m_j(z) \omega(z) \right).$$

In 24 cases, two recipients were each other's phylogenetically probable transmitter. We considered transmission in each direction equally likely. The relative transmission probabilities $\omega_{ij\tau}$ were calculated by

$$\omega_{ij\tau} = \omega_{ij} \phi_{ij} / \tau_{ij},$$

where ϕ_{ij} is equal to 0.5 if i and j are each other's phylogenetically probable transmitters, otherwise 1; ω_{ij} are the relative pairwise probabilities shown in fig. S5; and τ_{ij} is the number of transmission intervals between transmitter i and recipient j .

These probabilities sum to 1 per recipient. If all transmitters are sampled, we obtain $p_{ij\tau} = \omega_{ij\tau} / \sum_{k,s} \omega_{kjs}$. If some transmitters are not sampled, the first part of the denominator, $\sum_{k,s} \omega_{kjs}$, is smaller and adjusted by the second part of the denominator. The number of expected missing intervals $m_j(z)$ differs by stage and adjusts for stage-specific censoring and sampling biases.

The proportion of transmissions originating from the 14 infection/care stages was obtained by summing the corresponding individual-level transmission probabilities (fig. S8). Precisely, the proportion of transmissions from stage x to recipients diagnosed in $[t_1, t_2]$ was calculated by

$$P^T(x, t_1, t_2) = \frac{\sum_{j \in R(t_1, t_2)} p_j(x)}{\sum_z \sum_{j \in R(t_1, t_2)} p_j(z)} = \frac{1}{J} \sum_{j \in R(t_1, t_2)} p_j(x),$$

where $R(t_1, t_2)$ is the set of recipients with date of diagnosis in $[t_1, t_2]$, J is the number of recipients with date of diagnosis in $[t_1, t_2]$, and $p_j(x)$ is the probability that recipient j was infected by a transmitter in stage x . The probability $p_j(x)$ is the sum

$$p_j(x) = \sum_{i \in I_j} \sum_{\tau \in V_{ij(x)}} p_{ij\tau} + \sum_{v=1}^{m_j(x)} p_{jv},$$

where I_j are the observed, phylogenetically probable transmitters to recipient j ; $V_{ij(x)}$ is the set of observed transmission intervals between i and j in stage x ; and all other quantities are as defined above. The formula for $P^T(x, t_1, t_2)$ can be intuitively interpreted as the average

probability that a recipient was infected by a transmitter in stage x . Thus, the precision in the estimated $P^T(x, t_1, t_2)$ depends primarily on the number of recipients. We identified substantial individual-level variation in the transmission probabilities $p_j(x)$ (fig. S8), suggesting that a relatively large number of past transmission events are needed to reliably quantify sources of transmission.

Epidemiological prevention analysis

With the sources of transmission estimated, we compared the impact of prevention strategies in counterfactual scenarios that modeled the redistribution of phylogenetically identified transmitters to less infectious stages in the HIV infection and care continuum. This reduced the overall probability that any of the recipients would have been infected to less than 1. The proportion of infections that could have been averted in the period $[t_1, t_2]$ with a counterfactual prevention scenario H is

$$a(H) = 1 - \sum_{j \in R(t_1, t_2)} \sum_x p_j^H(x)$$

where $p_j^H(x)$ is the probability that recipient j is infected by someone in stage x under the counterfactual prevention scenario H . The individual-level prevention models are described in the Supplementary Materials.

Statistical uncertainty

Central estimates of $P^T(x, t_1, t_2)$ and $a(H)$ were obtained under central estimates of the genetic distances in fig. S4, the resulting phylogenetic transmission probabilities ω_{ij} , and the expected number of missing transmission intervals (Fig. 2C). Bootstrap sampling of the recipients, the empirical distribution of genetic distances, the number of missing transmission intervals under a negative binomial missing data model, and the counterfactual reallocation procedure of probable transmitters to less infectious infection/care stages were conducted to obtain nonparametric 95% confidence intervals. Confidence intervals were based on 1000 bootstrap replicates.

SUPPLEMENTARY MATERIALS

www.sciencetranslationalmedicine.org/cgi/content/full/8/320/320ra2/DC1

Extended acknowledgments

Materials and Methods

Fig. S1. Number of identified recipient MSM by 3-month intervals.

Fig. S2. Duration of infection windows of recipient MSM.

Fig. S3. Snapshot of the reconstructed viral phylogeny.

Fig. S4. Uncertainty in the estimated genetic distance between sequences from the transmitter and recipient of potential transmission pairs.

Fig. S5. Genetic distance between sequence pairs from previously published, epidemiologically confirmed transmitter-recipient pairs, and sequence pairs from the phylogenetically probable transmission pairs in this study.

Fig. S6. Right censoring at past, hypothetical database closure times.

Fig. S7. Sequence sampling probabilities by stage in the infection and care continuum.

Fig. S8. Individual-level variation in phylogenetically derived transmission probabilities by infection/care stages.

Fig. S9. Frequency of infection/care stages among phylogenetically probable transmitters.

Fig. S10. Phylogenetically derived transmission probabilities of observed transmission intervals.

Fig. S11. Transmission risk ratio from men after ART start compared to diagnosed untreated men with CD4 >500 cells/ml.

Fig. S12. Sensitivity analysis on the impact of PrEP with lower efficacy.

Fig. S13. Sensitivity analysis on the impact of lower or higher PrEP coverage.

Fig. S14. Impact of sampling and censoring adjustments on the estimated proportion of transmissions from stages in the infection and care continuum.

Fig. S15. Impact of phylogenetic transmission probabilities on the estimated proportion of transmissions from stages in the infection and care continuum.

Fig. S16. Impact of infection time estimates on the estimated proportion of transmissions from stages in the infection and care continuum.

Fig. S17. Impact of phylogenetic clustering criteria on the estimated proportion of transmissions from stages in the infection and care continuum.

Fig. S18. Impact of additional genetic distance criteria on the estimated proportion of transmissions from stages in the infection and care continuum.

Fig. S19. Impact of sequence sampling and censoring adjustments on the estimated proportion of averted infections.

Fig. S20. Impact of phylogenetic transmission probabilities on the estimated proportion of averted infections.

Fig. S21. Impact of infection time estimates and phylogenetic exclusion criteria on the estimated proportion of averted infections.

Fig. S22. Impact of additional genetic distance criteria on the estimated proportion of averted infections per biomedical intervention.

Fig. S23. Differences in transmission networks with and without a recipient MSM.

Fig. S24. Exploratory local polynomial regression fits to the time to diagnosis of MSM with a last negative test in the ATHENA cohort.

Fig. S25. Multivariable gamma regression model fitted to the time between the midpoint of the seroconversion interval and diagnosis of MSM with a last negative test in the ATHENA cohort.

Fig. S26. Estimated probability that the time between the midpoint of the seroconversion interval and diagnosis among MSM with a last negative test is larger than t years.

Fig. S27. Time to diagnosis estimates.

Fig. S28. Genetic distance among sequence pairs from transmitter-recipient pairs in the Belgium and Swedish transmission chains.

Fig. S29. Approximate type I error of the phylogenetic clustering criterion as a function of the clade frequency threshold.

Fig. S30. Type I error and power of the coalescence compatibility test.

Fig. S31. Estimated fraction of noncensored potential transmission intervals.

Fig. S32. Time between last negative test and diagnosis among MSM diagnosed in July 2009 to December 2010 and probable transmitters of recipients diagnosed in July 2009 to December 2010.

Table S1. Clinical and viral sequence data used in this study.

Table S2. Potential transmitters and potential transmission pairs to the recipient MSM.

Table S3. Identified phylogenetically probable transmitters and phylogenetically probable transmission pairs to the recipient MSM.

Table S4. Demographic and clinic characteristics of the 3025 MSM with a last negative test, which were used to fit the multivariable regression model.

References (48–52)

REFERENCES AND NOTES

1. M. S. Cohen, Y. Q. Chen, M. McCauley, T. Gamble, M. C. Hosseini, N. Kumarasamy, J. G. Hakim, J. Kumwenda, B. Grinsztejn, J. H. S. Pilotto, S. V. Godbole, S. Mehendale, S. Chariyalertsak, B. R. Santos, K. H. Mayer, I. F. Hoffman, S. H. Eshleman, E. Piwowar-Manning, L. Wang, J. Makhema, L. A. Mills, G. de Bruyn, I. Sanne, J. Eron, J. Gallant, D. Havlir, S. Swindells, H. Ribaud, V. Elharrar, D. Burns, T. E. Taha, K. Nielsen-Saines, D. Celentano, M. Essex, T. R. Fleming, Prevention of HIV-1 infection with early antiretroviral therapy. *N. Engl. J. Med.* **365**, 493–505 (2011).
2. A. Rodger, T. Bruun, V. Cambiano, P. Vernazza, V. Estrada, J. Van Lunzen, S. Collins, A. M. Geretti, A. Phillips, J. Lundgren, HIV transmission risk through condomless sex if HIV+ partner on suppressive ART: PARTNER Study, 21st Conference on Retroviruses and Opportunistic Infections (CROI'14), Boston, MA, 3 to 6 March 2014.
3. C. Beyrer, S. D. Baral, F. van Griensven, S. M. Goodreau, S. Chariyalertsak, A. L. Wirtz, R. Brookmeyer, Global epidemiology of HIV infection in men who have sex with men. *Lancet* **380**, 367–377 (2012).
4. P. S. Sullivan, A. Carballo-Diéguez, T. Coates, S. M. Goodreau, I. McGowan, E. J. Sanders, A. Smith, P. Goswami, J. Sanchez, Successes and challenges of HIV prevention in men who have sex with men. *Lancet* **380**, 388–399 (2012).
5. World Health Organization, *Guideline on When to Start Antiretroviral Therapy and on Pre-Exposure Prophylaxis for HIV* (WHO, Geneva, 2015).
6. A. Fogarty, L. Mao, I. Zablotska Manos, H. R. Santana, G. Prestage, J. Rule, P. Canavan, D. Murphy, D. McGuigan, *The Health in Men and Positive Health Cohorts: A Comparison of Trends in the Health and Sexual Behaviour of HIV-Negative and HIV-Positive Gay Men, 2002–2005* (National Centre in HIV Social Research, Sydney, 2006).
7. C. D. Pilcher, S. A. Fiscus, T. Q. Nguyen, E. Foust, L. Wolf, D. Williams, R. Ashby, J. O. O'Dowd, J. T. McPherson, B. Stalzer, L. Hightow, W. C. Miller, J. J. Eron Jr., M. S. Cohen, P. A. Leone, Detection of acute infections during HIV testing in North Carolina. *N. Engl. J. Med.* **352**, 1873–1883 (2005).

8. H. A. Weiss, J. N. Wasserheit, R. V. Barnabas, R. J. Hayes, L. J. Abu-Raddad, Persisting with prevention: The importance of adherence for HIV prevention. *Emerg. Themes Epidemiol.* **5**, 8 (2008).
9. A. van Sighem, L. Gras, A. Kesselring, C. Smit, I. Engelhard, I. Stolte, P. Reiss, *Monitoring Report 2013: Of Human Immunodeficiency Virus Infection in the Netherlands* (Stichting HIV Monitoring, Amsterdam, 2013).
10. T. T.-Y. Lam, C.-C. Hon, J. W. Tang, Use of phylogenetics in the molecular epidemiology and evolutionary studies of viral infections. *Crit. Rev. Clin. Lab. Sci.* **47**, 5–49 (2010).
11. D. P. Wilson, HIV treatment as prevention: Natural experiments highlight limits of antiretroviral treatment as HIV prevention. *PLOS Med.* **9**, e1001231 (2012).
12. E. Romero-Severson, H. Skar, I. Bulla, J. Albert, T. Leitner, Timing and order of transmission events is not directly reflected in a pathogen phylogeny. *Mol. Biol. Evol.* **31**, 2472–2482 (2014).
13. D. Pillay, A. Rambaut, A. M. Geretti, A. J. L. Brown, HIV phylogenetics. *BMJ* **335**, 460–461 (2007).
14. B. Vrancken, A. Rambaut, M. A. Suchard, A. Drummond, G. Baele, I. Derdelinckx, E. Van Wijngaerden, A.-M. Vandamme, K. Van Laethem, P. Lemey, The genealogical population dynamics of HIV-1 in a large transmission chain: Bridging within and among host evolutionary rates. *PLOS Comput. Biol.* **10**, e1003505 (2014).
15. P. Lemey, I. Derdelinckx, A. Rambaut, K. Van Laethem, S. Dumont, S. Vermeulen, E. Van Wijngaerden, A.-M. Vandamme, Molecular footprint of drug-selective pressure in a human immunodeficiency virus transmission chain. *J. Virol.* **79**, 11981–11989 (2005).
16. N. B. Carnegie, R. Wang, V. Novitsky, V. De Gruttola, Linkage of viral sequences among HIV-infected village residents in Botswana: estimation of linkage rates in the presence of missing data. *PLOS Comput. Biol.* **10**, e1003430 (2014).
17. F. van Griensven, T. H. Holtz, W. Thienkrua, W. Chonwattana, W. Wimonasate, S. Chaikummao, A. Varangrat, T. Chemnasiri, M. Sukwicha, M. E. Curlin, T. Samandari, A. Chitwarakorn, P. A. Mock, Temporal trends in HIV-1 incidence and risk behaviours in men who have sex with men in Bangkok, Thailand, 2006–13: An observational study. *Lancet HIV* **2**, e64–e70 (2015).
18. F. D. H. Koedijk, B. H. B. van Benthem, E. M. D. C. Vrolings, W. Zuilhof, M. A. B. van der Sande, Increasing sexually transmitted infection rates in young men having sex with men in the Netherlands, 2006–2012. *Emerg. Themes Epidemiol.* **11**, 12 (2014).
19. R. M. Grant, J. R. Lama, P. L. Anderson, V. McMahan, A. Y. Liu, L. Vargas, P. Goicochea, M. Casapia, J. V. Guanira-Carranza, M. E. Ramirez-Cardich, O. Montoya-Herrera, T. Fernández, V. G. Veloso, S. P. Buchbinder, S. Charialertsak, M. Schechter, L.-G. Bekker, K. H. Mayer, E. G. Kallás, K. R. Amico, K. Mulligan, L. R. Bushman, R. J. Hance, C. Ganoza, P. Defechereux, B. Postle, F. Wang, J. J. McConnell, J.-H. Zheng, J. Lee, J. F. Rooney, H. S. Jaffe, A. I. Martinez, D. N. Burns, D. V. Glidden, iPrEx Study Team, Preexposure chemoprophylaxis for HIV prevention in men who have sex with men. *N. Engl. J. Med.* **363**, 2587–2599 (2010).
20. S. McCormack, D. T. Dunn, M. Desai, D. I. Dolling, M. Gafos, R. Gilson, A. K. Sullivan, A. Clarke, I. Reeves, G. Schembri, N. Mackie, C. Bowman, C. J. Lacey, V. Apea, M. Brady, J. Fox, S. Taylor, S. Antonucci, S. H. Khoo, J. Rooney, A. Nardone, M. Fisher, A. McOwan, A. N. Phillips, A. M. Johnson, B. Gazzard, O. N. Gill, Pre-exposure prophylaxis to prevent the acquisition of HIV-1 infection (PROUD): Effectiveness results from the pilot phase of a pragmatic open-label randomised trial. *Lancet* **381**, S0140-6736(15)00056-2 (2015).
21. J. M. Molina, C. Capitant, B. Spire, G. Pialoux, C. Chidiac, I. Charreau, C. Tremblay, L. Meyer, J. F. Delfraissy, On demand PrEP with oral TDF-FTC in MSM: Results of the ANRS Ipergay Trial, Conference on Retroviruses and Opportunistic Infections (CROI'15), Seattle, WA, 23 to 26 February 2015.
22. F. Tanser, T. Barnighausen, E. Grapsa, J. Zaidi, M.-L. Newell, High coverage of ART associated with decline in risk of HIV acquisition in rural KwaZulu-Natal, South Africa. *Science* **339**, 966–971 (2013).
23. D. Frentz, A. M. J. Wensing, J. Albert, D. Paraskevis, A. B. Abecasis, O. Hamouda, L. B. Jørgensen, C. Kücherer, D. Struck, J.-C. Schmit, B. Åsjö, C. Balotta, D. Beshkov, R. J. Camacho, B. Clotet, S. Coughlan, S. De Wit, A. Griskevicius, Z. Grossman, A. Horban, T. Kolupajeva, K. Korn, L. G. Kostrikis, K. Liitsola, M. Linka, C. Nielsen, D. Otelea, R. Paredes, M. Poljak, E. Puchhammer-Stöckl, A. Sönnnerborg, D. Stanekova, M. Stanojevic, A.-M. Vandamme, C. A. B. Boucher, D. A. M. C. Van de Vijver, SPREAD Programme, Limited cross-border infections in patients newly diagnosed with HIV in Europe. *Retrovirology* **10**, 36 (2013).
24. B. G. Brenner, M. A. Wainberg, Future of phylogeny in HIV prevention. *J. Acquir. Immune Defic. Syndr.* **63**, S248–S254 (2013).
25. A. B. Cope, K. A. Powers, J. D. Kuruc, P. A. Leone, J. A. Anderson, L.-H. Ping, L. P. Kincer, R. Swanstrom, V. L. Mobley, E. Foust, C. L. Gay, J. J. Eron, M. S. Cohen, W. C. Miller, Ongoing HIV transmission and the HIV care continuum in North Carolina. *PLOS One* **10**, e0127950 (2015).
26. J. Skarbinski, E. Rosenberg, G. Paz-Bailey, H. I. Hall, C. E. Rose, A. H. Viall, J. L. Fagan, A. Lansky, J. H. Mermin, Human immunodeficiency virus transmission by each step of the care continuum in the United States. *JAMA Intern. Med.* **175**, 588–596 (2015).
27. E. S. Rosenberg, G. A. Millet, P. S. Sullivan, C. del Rio, J. W. Curran, Understanding the HIV disparities between black and white men who have sex with men in the USA using the HIV care continuum: A modeling study. *Lancet HIV* **1**, e112–e118 (2014).
28. INSIGHT START Study Group, J. D. Lundgren, A. G. Babiker, F. Gordin, S. Emery, B. Grund, S. Sharma, A. Avihingsanon, D. A. Cooper, G. Fätkenheuer, J. M. Llibre, J. M. Molina, P. Munderi, M. Schechter, R. Wood, K. L. Klingman, S. Collins, H. C. Lane, A. N. Phillips, J. D. Neaton, Initiation of antiretroviral therapy in early asymptomatic HIV infection. *N. Engl. J. Med.* **373**, 795–807 (2015).
29. E. M. Volz, E. Ionides, E. O. Romero-Severson, M.-G. Brandt, E. Mokotoff, J. S. Koopman, HIV-1 transmission during early infection in men who have sex with men: A phylodynamic analysis. *PLOS Med.* **10**, e1001568 (2013).
30. *Truvada Approved to Reduce the Risk of Sexually Transmitted HIV in People Who Are Not Infected with the Virus* (U.S. Food and Drug Administration, Silver Spring, MD, 2012).
31. R. M. Grant, P. L. Anderson, V. McMahan, A. Liu, K. R. Amico, M. Mehrotra, S. Hosenk, C. Mosquera, M. Casapia, O. Montoya, S. Buchbinder, V. G. Veloso, K. Mayer, S. Charialertsak, L.-G. Bekker, E. G. Kallás, M. Schechter, J. Guanira, L. Bushman, D. N. Burns, J. F. Rooney, D. V. Glidden, iPrEx study team, Uptake of pre-exposure prophylaxis, sexual practices, and HIV incidence in men and transgender women who have sex with men: A cohort study. *Lancet Infect. Dis.* **14**, 820–829 (2014).
32. A. Liu, S. Cohen, S. Follansbee, D. Cohan, S. Weber, D. Sachdev, S. Buchbinder, Early experiences implementing pre-exposure prophylaxis (PrEP) for HIV prevention in San Francisco. *PLOS Med.* **11**, e1001613 (2014).
33. J. P. Bil, U. Davidovich, W. M. van der Veldt, M. Prins, H. J. C. de Vries, G. J. B. Sonder, I. G. Stolte, What do Dutch MSM think of preexposure prophylaxis to prevent HIV-infection? A cross-sectional study. *AIDS* **29**, 955–964 (2015).
34. M. J. Mimiaga, J. M. White, D. S. Krakower, K. B. Biello, K. H. Mayer, Suboptimal awareness and comprehension of published preexposure prophylaxis efficacy results among physicians in Massachusetts. *AIDS Care* **26**, 684–693 (2014).
35. K. H. Mayer, S. Hosenk, S. Cohen, A. Liu, J. Pickett, M. Warren, D. Krakower, R. Grant, Antiretroviral pre-exposure prophylaxis implementation in the United States: A work in progress. *J. Int. AIDS Soc.* **18**, 19980 (2015).
36. D. Pao, M. Fisher, S. Hué, G. Dean, G. Murphy, P. A. Cane, C. A. Sabin, D. Pillay, Transmission of HIV-1 during primary infection: Relationship to sexual risk and sexually transmitted infections. *AIDS* **19**, 85–90 (2005).
37. N. Pant Pai, J. Sharma, S. Shivkumar, S. Pillay, C. Vadnais, L. Joseph, K. Dheda, R. W. Peeling, Supervised and unsupervised self-testing for HIV in high- and low-risk populations: A systematic review. *PLOS Med.* **10**, e1001414 (2013).
38. N. Lorente, M. Preau, C. Vernay-Vaisse, M. Mora, J. Blanche, J. Otis, A. Passeron, J.-M. Le Gall, P. Dhotte, M. P. Carrieri, M. Suzan-Monti, B. Spire, ANRS-DRAG Study Group, Expanding access to non-medicalized community-based rapid testing to men who have sex with men: An urgent HIV prevention intervention (The ANRS-DRAG study). *PLOS One* **8**, e61225 (2013).
39. *Time to Test for HIV: Expanding HIV Testing in Healthcare and Community Services in England* (Health Protection Agency, London, 2011).
40. G. B. Gomez, A. Borquez, K. K. Case, A. Wheelock, A. Vassall, C. Hankins, The cost and impact of scaling up pre-exposure prophylaxis for HIV prevention: A systematic review of cost-effectiveness modelling studies. *PLOS Med.* **10**, e1001401 (2013).
41. R. B. Birger, T. B. Hallett, A. Sinha, B. T. Grenfell, S. L. Hodder, Modeling the impact of interventions along the HIV continuum of care in Newark, New Jersey. *Clin. Infect. Dis.* **58**, 274–284 (2014).
42. J. Heuker, G. J. B. Sonder, I. Stolte, R. Geskus, A. van den Hoek, High HIV incidence among MSM prescribed postexposure prophylaxis, 2000–2009: Indications for ongoing sexual risk behaviour. *AIDS* **26**, 505–512 (2012).
43. V. A. Johnson, V. Calvez, H. F. Günthard, R. Paredes, D. Pillay, R. W. Shafer, A. M. Wensing, D. D. Richman, Update of the drug resistance mutations in HIV-1: March 2013. *Top. Antivir. Med.* **21**, 6–14 (2013).
44. B. D. Rice, J. E. Elford, Z. Yin, V. C. Delpech, A new method to assign country of HIV infection among heterosexuals born abroad and diagnosed with HIV. *AIDS* **26**, 1961–1966 (2012).
45. A. M. Kozlov, A. J. Aberer, A. Stamatakis, ExaML version 3: A tool for phylogenomic analyses on supercomputers. *Bioinformatics* **31**, 2577–2579 (2015).
46. T. Leitner, D. Escanilla, C. Franzén, M. Uhlén, J. Albert, Accurate reconstruction of a known HIV-1 transmission history by phylogenetic tree analysis. *Proc. Natl. Acad. Sci. U.S.A.* **93**, 10864–10869 (1996).
47. A. van Sighem, F. Nakagawa, D. De Angelis, C. Quinten, D. Bezemer, E. O. de Coul, M. Egger, F. de Wolf, C. Fraser, A. Phillips, Estimating HIV incidence, time to diagnosis, and the undiagnosed HIV epidemic using routine surveillance data. *Epidemiology* **26**, 653–660 (2015).
48. *Longitudinal Analysis of the Trajectories of CD4 Cell Counts* (Health Protection Agency, London, 2011).
49. D. Bezemer, F. de Wolf, M. C. Boerlijst, A. van Sighem, T. D. Hollingsworth, C. Fraser, 27 years of the HIV epidemic amongst men having sex with men in the Netherlands: An in depth mathematical model-based analysis. *Epidemics* **2**, 66–79 (2010).
50. S. H. Eshleman, S. E. Hudelson, A. D. Redd, L. Wang, R. Debes, Y. Q. Chen, C. A. Martens, S. M. Ricklefs, E. J. Selig, S. F. Porcella, S. Munshaw, S. C. Ray, E. Piwovar-Manning, M. McCauley, M. C. Hosseinipour, J. Kumwenda, J. G. Hakim, S. Charialertsak, G. de Bruyn, B. Grinsztajn, N. Kumarasamy, J. Makhema, K. H. Mayer, J. Pilotto, B. R. Santos, T. C. Quinn, M. S. Cohen, J. P. Hughes, Analysis of genetic linkage of HIV from couples enrolled in the HIV Prevention Trials Network 052 trial. *J. Infect. Dis.* **204**, 1918–1926 (2011).

51. A. Gavryushkina, D. Welch, T. Stadler, A. J. Drummond, Bayesian inference of sampled ancestor trees for epidemiology and fossil calibration. *PLoS Comput. Biol.* **10**, e1003919 (2014).
52. S. B. McCombs, E. McCray, D. A. Wendell, P. A. Sweeney, I. M. Onorato, Epidemiology of HIV-1 infection in bisexual women. *J. Acquir. Immune Defic. Syndr.* **5**, 850–852 (1992).
53. B. Efron, R. J. Tibshirani, *An Introduction to the Bootstrap* (Chapman and Hall, Boca Raton, FL, 1998).

Acknowledgments: We thank the Imperial College High Performance Computing Service (www3.imperial.ac.uk/ict/services/hpc), three anonymous referees, the HIV treating physicians, and HIV nurse consultants and staff of the diagnostic laboratories and facilities in the HIV treatment centers, along with the data collecting and monitoring staff both within and outside the Stichting HIV Monitoring Foundation for their contributions to make this work possible (see Extended Acknowledgments in the Supplementary Materials). **Funding:** O.R. is supported by the Wellcome Trust (fellowship WR092311MF), and C.F. by the European Research Council (Advanced Grant PBDR-339251) and the Bill & Melinda Gates Foundation (PANGEA-HIV consortium). P.R. through his institution received independent scientific grant support from Bristol-Myers Squibb, ViiV Healthcare, Gilead Sciences, Janssen Pharmaceuticals Inc., and Merck & Co; served on a scientific advisory board for Gilead Sciences; and serves on a data safety monitoring committee for Janssen Pharmaceuticals Inc., for which his institution has received remuneration. The ATHENA observational cohort study is part of Stichting HIV Monitoring and supported by a grant from the Netherlands Ministry of Health, Welfare and Sport through its

Centre for Infectious Disease Control–National Institute for Public Health and the Environment. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. **Author contributions:** O.R., F.d.W., P.R., and C.F. conceived the study. O.R. and C.F. developed the methods, did the analysis, and reviewed all statistical aspects of the analysis. A.v.S., D.B., S.J., and A.W. provided the data used in the analysis. A.G. assisted in estimating the viral phylogeny. A.v.S., D.B., F.d.W., and P.R. gave advice on analysis and interpretation. O.R. and C.F. wrote the first draft. All authors reviewed and approved the final version. **Competing interests:** The authors declared that they have no competing interests. **Data and materials availability:** Data are available from the HIV Monitoring Institutional Data Access/Ethics Committee for researchers who meet the criteria for access to confidential data. Contact E-mail: secretariaat.shm@amc.uva.nl.

Submitted 5 August 2015

Accepted 3 December 2015

Published 6 January 2016

10.1126/scitranslmed.aad1863

Citation: O. Ratmann, A. van Sighem, D. Bezemer, A. Gavryushkina, S. Juriaans, A. Wensing, F. de Wolf, P. Reiss, C. Fraser, ATHENA observational cohort, Sources of HIV infection among men having sex with men and implications for prevention. *Sci. Transl. Med.* **8**, 320ra2 (2016).

Sources of HIV infection among men having sex with men and implications for prevention

Oliver Ratmann, Ard van Sighem, Daniela Bezemer, Alexandra Gavryushkina, Suzanne Jurriaans, Annemarie Wensing, Frank de Wolf, Peter Reiss, Christophe Fraser and ATHENA observational cohort

Sci Transl Med **8**, 320ra2320ra2.
DOI: 10.1126/scitranslmed.aad1863

The ART of HIV prevention

Despite the relative success of antiretroviral therapy (ART) for individuals infected with HIV, the rate of new diagnoses has remained fairly constant in vulnerable population groups, particularly men having sex with men (MSM). Now, ART is also available in the United States to uninfected individuals to directly prevent infection with the virus. Ratmann *et al.* were able to reconstruct ~600 past transmission events among men having sex with men in the Netherlands, and examined probable sources of transmission. They found that the large majority of new infections is neither attributable to ineffective ART nor inadequate retention in care. Rather, many of these cases could have been averted with more comprehensive HIV testing and a broader use of ART that includes provision to uninfected men as well as starting ART as soon as possible among newly diagnosed men. These findings support making ART for pre-exposure prophylaxis available worldwide, and especially in countries with high retention in care and high ART coverage among infected MSM.

ARTICLE TOOLS <http://stm.sciencemag.org/content/8/320/320ra2>

SUPPLEMENTARY MATERIALS <http://stm.sciencemag.org/content/suppl/2016/01/04/8.320.320ra2.DC1>

Use of this article is subject to the [Terms of Service](#)

**RELATED
CONTENT**

<http://stm.sciencemag.org/content/scitransmed/7/270/270ra5.full>
<http://stm.sciencemag.org/content/scitransmed/7/270/270ra4.full>
<http://stm.sciencemag.org/content/scitransmed/4/151/151ra125.full>
<http://science.sciencemag.org/content/sci/351/6272/434.full>
<http://stm.sciencemag.org/content/scitransmed/8/336/336ra62.full>
<http://science.sciencemag.org/content/sci/352/6288/1001.full>
<http://science.sciencemag.org/content/sci/352/6288/997.full>
<http://science.sciencemag.org/content/sci/353/6294/18.full>
<http://science.sciencemag.org/content/sci/353/6295/172.full>
<http://science.sciencemag.org/content/sci/353/6298/432.full>
<http://science.sciencemag.org/content/sci/353/6298/506.full>
<http://science.sciencemag.org/content/sci/353/6307/1557.full>
<http://science.sciencemag.org/content/sci/354/6309/157.full>
<http://science.sciencemag.org/content/sci/354/6309/177.full>
<http://science.sciencemag.org/content/sci/354/6317/1213.full>
<http://science.sciencemag.org/content/sci/354/6318/1434.full>
<http://science.sciencemag.org/content/sci/354/6318/1384.full>
<http://science.sciencemag.org/content/sci/355/6320/89.full>
<http://science.sciencemag.org/content/sci/355/6320/93.full>
<http://science.sciencemag.org/content/sci/354/6312/535.full>
<http://science.sciencemag.org/content/sci/352/6287/828.full>
<http://science.sciencemag.org/content/sci/353/6297/aaf6517.full>
<http://science.sciencemag.org/content/sci/354/6309/197.full>

REFERENCES

This article cites 42 articles, 4 of which you can access for free
<http://stm.sciencemag.org/content/8/320/320ra2#BIBL>

PERMISSIONS

<http://www.sciencemag.org/help/reprints-and-permissions>

Use of this article is subject to the [Terms of Service](#)

Science Translational Medicine (ISSN 1946-6242) is published by the American Association for the Advancement of Science, 1200 New York Avenue NW, Washington, DC 20005. 2017 © The Authors, some rights reserved; exclusive licensee American Association for the Advancement of Science. No claim to original U.S. Government Works. The title *Science Translational Medicine* is a registered trademark of AAAS.