A rapidly deployable individualized system for augmenting ventilator capacity

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Strategies to split ventilators to support multiple patients requiring ventilatory support have been proposed and used in emergency cases in which shortages of ventilators cannot otherwise be remedied by production or procurement strategies. However, the current approaches to ventilator sharing lack the ability to individualize ventilation to each patient, measure pulmonary mechanics, and accommodate rebalancing of the airflow when one patient improves or deteriorates, posing safety concerns to patients. Potential cross-contamination, lack of alarms, insufficient monitoring, and inability to adapt to sudden changes in patient status have prevented widespread acceptance of ventilator sharing. We have developed an individualized system for augmenting ventilator efficacy (iSAVE) as a rapidly deployable platform that uses a single ventilator to simultaneously and more safely support two individuals. The iSAVE enables individual-specific volume and pressure control and the rebalancing of ventilation in response to improvement or deterioration in an individual’s respiratory status. The iSAVE incorporates mechanisms to measure pulmonary mechanics, mitigate cross-contamination and backflow, and accommodate sudden flow changes due to individual interdependencies within the respiratory circuit. We demonstrate these capacities through validation using closed- and open-circuit ventilators on linear test lungs. We show that the iSAVE can temporarily ventilate two pigs on one ventilator as efficaciously as each pig on its own ventilator. By leveraging off-the-shelf medical components, the iSAVE could rapidly expand the ventilation capacity of health care facilities during emergency situations such as pandemics.

INTRODUCTION

Ventilators are vital equipment for critical-care conditions, surgical interventions, and procedures requiring anesthesia. Ventilators assist with respiration by moving air into and out of the lungs and are extensively used in the treatment of patients with acute respiratory distress syndrome (ARDS) and respiratory failure. Pandemics of infections that cause severe respiratory dysfunction can lead to a rapid increase in the volume of patients requiring ventilation, exceeding the number of ventilators available in health care facilities and necessitating difficult triage decisions (1, 2). The most recent publicly available data (2010) report that about 62,000 full-featured mechanical ventilators are available within the United States, with an additional 12,700 stockpiled in the Centers for Disease Control and Prevention strategic national stockpile (1) . On the basis of quantified disruptions in supply chain, recent coronavirus disease 2019 (COVID-19) hospitalization rates, and predictive models, estimates of the shortfall of ventilators within the United States range between 45,000 and 160,000 (1–4). Globally, the World Health Organization estimates that fewer than 2000 ventilators are available across the 41 African countries (5) to support a population of 1.2 billion people. Coupled with high population density and relative poverty, pandemics can make the procurement, building, or buying of new emergency ventilators challenging for some countries. Low-cost and emergency ventilators have been developed to address the cost barrier associated with the acquisition of new ventilators at scale. However, their manufacturing and deployment rely on supply, assembly, and distribution chains, which can be disrupted by public health crises involving lockdowns and import/export restrictions. The use of new ventilator designs has also raised safety concerns, as clinical staff would need to operate unfamiliar technology. Given both supply and implementation hurdles that may delay rapid deployment of low-cost ventilators, other strategies warrant consideration.

Ventilator sharing, or dividing the airflow from one ventilator among multiple patients, has been previously performed in a few emergency cases (6). By using readily available tubing and ventilator equipment, ventilator sharing can be immediately implemented to expand the capacity of existing ventilators with which clinicians are familiar. In previously proposed configurations, multiplexing ventilation involved connecting multiple outflow tracts to the ventilator to divide flow among patients (6–9), wherein the compliance (C) and resistance (R) of each patient’s pulmonary system became part of the same circuit and drove the balance of airflow. This patient interdependence poses various safety concerns: (i) Independent control of volume and pressure to each patient is not possible, which is important for lung-protective ventilation and the standard of care for ARDS; (ii) alarm monitoring becomes challenging because of the complex circuit configuration; (iii) sudden events, such as pneumothorax, tube occlusion, or disconnection of an endotracheal tube, cause potentially harmful rebalancing of ventilation; and
RESULTS

Design of the iSAVE

The iSAVE uses a series of valves and flow regulators in parallel limbs to effectively maintain the desired tidal volume ($V_T$) and positive end-expiratory pressure (PEEP) for each patient (Fig. 1 and tables S1 and S2) under volume control mode. In closed-circuit ventilators (Fig. 1), Y or T connectors are used to multiplex individual inspiratory channels for each patient. Each inspiratory channel consists of a filter, flow control valve, a one-way flow valve, and standard sensors (pressure, flow, and capnostat) in series. The expiratory limb consists of a filter, pressure release valve, and one-way valve before connection to Y or T connectors, which are routed back to the ventilator. The iSAVE can be configured to both open-circuit ventilators (which consist of only an inspiratory limb with passive expiration) and closed-circuit ventilators (which have inspiratory and expiratory limbs). Open-circuit ventilators can be used with the same circuit shown in Fig. 1, except that the expiratory limb would be connected to a Whisper Swivel (Philips Respironics) exhalation adaptor or equivalent valve for the expiratory flow port. We used positive expiratory pressure (PEP) threshold devices as one-way valves, set to their highest setting, because these are readily available in hospitals. See tables S1 and S2 for a full list of supplies.

The flow control valve is used to allocate the appropriate $V_T$ to each patient. Filters on each limb mitigate cross-contamination between individual patient circuits and filter expired gas before release into the room through the pressure release valve, thus limiting pathogen exposure to health care workers. The one-way valves prevent backflow and mitigate overdistention in cases of rapid flow change. PEEP valves ensure the individualized control of PEEP for each patient and function as pop-off valves to release excess pressure. Pressure, flow, and CO$_2$ sensors are positioned on the patient Y piece, and data, visualized on a separate patient monitor, are used for setup, titration, and monitoring. During initial setup, the respiratory rate (RR), PEEP, FiO$_2$ (fraction of inspired oxygen), inspiration:expiration (I:E) ratio, and the sum of the $V_T$ for each patient are set on the ventilator under volume control mode. Then, the flow control and PEEP valves can be titrated to individualize $V_T$ and PEEP. The exhaled volume and minute ventilation alarms on the ventilator are set according to the sum of the exhaled tidal volumes from both patients and enable the ventilator to alarm in response to sudden change in either patient’s status (shunt, occlusion, or disconnection of the endotracheal tube). Through this design, the iSAVE overcomes many of the aforementioned challenges of splitting ventilation (Table 1).

Benchtop testing of the iSAVE using linear test lungs

The iSAVE was connected to three common models of intensive care unit (ICU) ventilators (Hamilton, Puritan Bennett, and Philips) (fig. S1). Testing confirmed that the circuit was generalizable and could be used to ventilate two linear test lungs. Individualized management of ventilation and patient interdependence

We tested the ability of the iSAVE to meet the diverse $V_T$ needs that may be presented by patients of varying sizes and respiratory mechanics. We first delivered $V_T$ of 800 ml to two test lungs with healthy ($C = 50$ ml/cm H$_2$O, $R = 5$ cm H$_2$O/liter per second) and diseased ($C = 20$ ml/cm H$_2$O, $R = 5$ cm H$_2$O/liter per second) respiratory characteristics. By titrating the flow control valve, we were able to achieve differential $V_T$, spanning ratios from 50:50 to 15:85 (Fig. 2A). However, at ratios more disproportionate than 20:80, pressure exceeded 40 cm H$_2$O, which is supratherapeutic.

Accommodating lung compliance and resistance changes

Respiratory mechanics for patients with ARDS can vary considerably and evolve rapidly throughout the course of disease and recovery. Assuming that patients are initially matched, clinical improvement or deterioration will yield mismatches in airflow, requiring individualized management for optimized therapy. We tested the ability of the iSAVE to compensate for static changes in compliance and resistance of one test lung while minimizing effects on the other lung. For all tests, we (i) measured the baseline ventilation values, (ii) performed an intervention to change compliance or resistance, (iii) noted any safety features or alarms that were activated, and (iv) titrated valves to restore ventilation to baseline (within 5% error).

We decreased the compliance of one of the test lungs (from $C = 50$ ml/cm H$_2$O to $C = 15$ ml/cm H$_2$O), simulating the parameters characteristic of ARDS (Fig. 2B). Flow was quickly diverted, resulting in a disproportionate volume being delivered to the healthy lung ($C = 50$ ml/cm H$_2$O). The PEEP valve released excess volume during the period of titration, preventing overdistention of the healthy lung. The desired volume of 400 ml per lung was restored by titrating the flow control valve. We next began with two lungs simulating ARDS ($C = 20$ ml/cm H$_2$O, $R = 5$ cm H$_2$O/liter per second) and increased the compliance of one lung (from $C = 20$ ml/cm H$_2$O to $C = 40$ ml/cm H$_2$O), which created a shift in the volumes delivered. Adjustment of the flow control valve restored the desired flow to both lungs (Fig. 2C).

We then performed ventilation (300 ml per lung) under high resistances, simulating the physiology characteristic of the comorbidities commonly associated with ARDS, including bacterial pneumonia, asthma, chronic obstructive pulmonary disease/emphysema, and presence of viscous airway secretions. With both lungs simulating ARDS ($C = 20$ ml/cm H$_2$O, $R = 5$ cm H$_2$O/liter per second), the resistance of one lung was increased 10-fold ($R = 5$ cm H$_2$O/liter...
per second to $R = 50 \text{ cm H}_2\text{O/liter per second}$, causing a marked reduction in the flow to the lung. Titration of the valve along with an increase in the inspiratory time enabled the desired flow to be achieved while maintaining lower pressures (Fig. 2D). The aforementioned benchtop tests were also performed with an open-circuit ventilator, yielding similar results (figs. S2 to S4).

Managing abrupt changes in respiratory status and adjusting the number of individuals on the iSAVE

Whereas subacute changes in resistance and compliance can be accommodated, it is vital that the iSAVE enables alarms in response to acute changes for patient safety. The ventilator alarm was set to detect changes in the overall expiratory volume. We mechanically occluded tubing of one lung to simulate an instantaneous change in resistance, which created a reduction of flow in one channel and spike in pressures/volumes of the other (fig. S5). This successfully caused the ventilator alarm to activate instantaneously. We also simulated the loss of the endotracheal tube to one lung, resulting in a leak in the system and yielding minimized flow to the other lung. This activated the main ventilator leak alarm. We immediately closed the valve to the disconnected lung’s circuit, effectively closing the leak and removing the test lung from the circuit. We then titrated the other lung’s valve to deliver the desired volume (Fig. 2E).

This process of shutting airflow to one segment of the circuit could be used in cases such as cardiac arrest or weaning from the ventilator to remove a patient from shared ventilation without leaking air and thereby mitigating potential aerosolization of infectious agents.

A practical challenge to be managed by this system is the addition of patients without excessive disruption to other patients. We simulated the addition of a second patient to a shared ventilator while minimizing deleterious effects to the original patient with a series of protocolized steps using artificial lungs (fig. S6).

Cross-contamination

It is critical that the iSAVE prevent cross-contamination under potentially turbulent or unusual airflow patterns caused by shared ventilation, particularly for use in patients with highly airborne and infectious pathogens. Contamination patterns were tested by nebulizing trypan blue into the airflow tract of one artificial lung and testing for contaminants in each segment of the circuit using a closed-circuit ventilator (fig. S7A). Even at unrealistic conditions yielding turbulent flows at high pressures [nebulization at 40 cm H$_2$O, 10 min of continuous nebulization, 5 ml of nebulized particles, $V_T = 300$ ml, and RR = 30 breaths/min (bpm)], no cross contamination of filters was visually observed (fig. S7B) or detected in wipe tests of each segment of the circuit (fig. S7C).

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Fig. 1. Design of the individualized system for augmenting ventilation efficacy. (A) Schematic of iSAVE setup on a closed-circuit ventilator for simultaneous ventilation of two patients. (B) Circuit diagram of iSAVE for closed-circuit ventilation. (C) Photograph of iSAVE connected to a Puritan Bennett 840 ICU ventilator and two test lungs.
Measuring pulmonary mechanics

With shared ventilation, pulmonary mechanics used to optimize ventilation become challenging to evaluate for each patient. The measurement of plateau pressure ($P_{plat}$), the pressure maintained during inspiration when flow is equal to zero and approximates the alveolar pressure, can be used to calculate the lungs’ $R$ and $C$. On closed-circuit ventilators with iSAVE, the end-inspiratory hold feature can be used to yield the $P_{plat}$. On open-circuit ventilators with iSAVE, an additional flow valve can be added to the expiratory channel and briefly closed at the end of inspiration to yield $P_{plat}$. While connected to two artificial test lungs using a closed-circuit ventilator, we performed an inspiratory hold and visualized $P_{plat}$. Because of the circuits’ interdependence, $P_{plat}$ was uniform between both lungs; however, PEEP and tidal volumes differed. Thus, accurate compliances can be computed. In three separate trials, we simulated this procedure with artificial lungs of varying compliances. Computed compliances were $\leq 11\%$ error of the set compliances (table S3).

In vivo testing

**Individualized management of ventilation and patient interdependence**

To evaluate the practical management of rebalancing ventilation using the iSAVE when ventilating real lungs, which exhibit variable respiratory mechanics, we performed closed-circuit ventilation using a large animal (Yorkshire swine, 70 kg) alongside an artificial lung (fig. S8A). The pig has a lung capacity of 5 to 6 liters, similar to human lungs, and serves as a translational preclinical model for this testing. We varied the lung mechanics of either the porcine or artificial lung ($C = 20$, $50$, $60$, and $120$ ml/cm H$_2$O) to test the system performance, delivering $V_T = 500$ ml from the ventilator. The iSAVE delivered differential volumes to each channel. Ratios of 50:50, 40:60, and 30:70 are shown in fig. S8B.

**Accommodation to static compliance changes and altered respiratory status**

To evaluate the system’s response to changes in lung compliance, we initially distributed $V_T$ from the ventilator equally to each channel (600 ml total, 300 ml per channel). The compliance of the artificial lung was decreased (from $C = 120$ ml/cm H$_2$O to $C = 60$ ml/cm H$_2$O), resulting in a greater allocation of volume (~100 ml) to the porcine lung (fig. S8C). By titrating the flow valve, $V_T$ distribution was restored within three breaths.

During the course of recovery from ARDS, lung compliance increases; thus, we tested a range of such scenarios ($C = 20$, $50$, $60$, and $120$ ml/cm H$_2$O). To model an extreme case, we adjusted the artificial lung compliance from $C = 20$ ml/cm H$_2$O to $C = 120$ ml/cm H$_2$O: iSAVE immediately diverted flow away from the lower compliance in the animal’s lungs toward the more compliant artificial lung (fig. S8D). Valve adjustment restored the desired flow to the animal’s lungs and.

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**Table 1. Key challenges in splitting ventilation.** A comparison of the capabilities of existing splitting mechanisms and iSAVE. PEEP, positive end-expiratory pressure; $\text{FiO}_2$, fraction of inspired oxygen; $\Delta C$, change in compliance; $\Delta R$, change in resistance; $P_{plat}$, plateau pressure.

<table>
<thead>
<tr>
<th>Concern</th>
<th>Uniform splitting (pressure control mode)</th>
<th>iSAVE (volume control mode)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Individualized management of ventilation</td>
<td>x Shared between patients</td>
<td>o Individualized to each patient</td>
</tr>
<tr>
<td>-PEEP</td>
<td></td>
<td></td>
</tr>
<tr>
<td>-Tidal volume</td>
<td>x Shared between patients</td>
<td>o Individualized to each patient</td>
</tr>
<tr>
<td>-$\text{FiO}_2$ respiratory rate</td>
<td>x Shared between patients</td>
<td>x Shared between patients</td>
</tr>
<tr>
<td>-Alarms</td>
<td>x Changes to one patient's status may not result in main ventilator alarm.</td>
<td>o Changes to one patient's status will cause main ventilator to alarm. Mechanical components to provide auditory alarms can be incorporated.</td>
</tr>
<tr>
<td>Sudden changes to patient status can cause damaging rebalancing of airflow to other patient(s) toward most compliant lungs.</td>
<td>x Ventilation cannot be quickly adjusted.</td>
<td>o Can be managed by titrating flow control valves. One-way valves prevent backflow. Pressure release valves prevent excess pressure delivery.</td>
</tr>
<tr>
<td>Improvement or deterioration of one patient ($\Delta C$, $\Delta R$) will automatically rebalance airflow, potentially harming other patient(s).</td>
<td>x Ventilation cannot be individually rebalanced. Patients would need to be rematched as they improve/deteriorate.</td>
<td>o Desired ventilation for each patient can be achieved through valve adjustment, allowing patients to improve/deteriorate while remaining on the same system.</td>
</tr>
<tr>
<td>Abruptly removing patients requires breaking the circuit, causing aerosolization of the virus, exposing health care personnel.</td>
<td>x Individual patient circuits cannot be quickly removed from circuit.</td>
<td>o Individual patients can be quickly shunted/removed from the circuit. Inline filters limit aerosolization risk.</td>
</tr>
<tr>
<td>Monitoring</td>
<td>x Additional respiratory monitors and heightened clinical vigilance required</td>
<td>x Additional respiratory monitors and heightened clinical vigilance required</td>
</tr>
<tr>
<td>Measurement of pulmonary mechanics</td>
<td>x Shared between patients</td>
<td>o $P_{plat}$ can be measured using expiratory hold button. C and R can be computed for each patient.</td>
</tr>
<tr>
<td>Ventilator calibration/self-test</td>
<td>x Added circuit volume defeats the operational self-test.</td>
<td>o Can be executed with modifications to circuit*</td>
</tr>
<tr>
<td>Triggering</td>
<td>x Disabled. Patients will require sedation.</td>
<td>x Disabled. Patients will require sedation.</td>
</tr>
</tbody>
</table>

*See fig. S9 for details regarding the rerouting of standard sensing devices required for ventilator calibration and self-tests.
Fig. 2. Individualized ventilation and management of patient interdependence using artificial test lungs. (A) Pressure, flow, and tidal volume waveforms illustrating three settings of differential tidal volume (VT) for two test lungs (blue and black) using closed-circuit ventilation. The ratio (50:50, 35:65, and 15:85) refers to the VT of the black:blue lungs. Pressure, volume, and flow in both lungs upon (B) decreased compliance in one lung (black) and (C) increased compliance in the other lung (blue). The orange dotted line indicates decrease or increase in compliance. The green dotted line indicates return of baseline ventilation parameters upon titration of the valves. Pressure, volume, and flow in both lungs upon (D) increased resistance in one lung (black) and (E) decreased resistance in the other lung (blue). Orange dotted line indicates increase or decrease in resistance. Green dotted line indicates return of baseline ventilation parameters upon titration of the valves. Waveforms from each lung are slightly offset to enable visualization.
artificial lung. Throughout these tests, end-tidal carbon dioxide (EtCO₂) and oxygen saturation (SpO₂) were maintained between 38 and 41 mmHg and 91 and 98%, respectively, at baseline and after titration.

After euthanasia, the animal’s lungs were filled with 750 ml of saline, effectively decreasing their compliance and resulting in a lower Vₜ. With repeated breaths, flow was further diminished despite efforts by the automatic adjustments of the ventilator in increasing the pressure (fig. S8E). Titration of the flow valve enabled restoration of the desired volume. We also simulated scenarios such as tube clogging and aspiration, in which the endotracheal tube of the artificial lung was mechanically obstructed. This immediately increased flow to the animal by 30% (fig. S8F) and was quickly resolved by closing the valve to the artificial lung circuit and adjusting flow to the animal.

**Validating iSAVE ventilation against standard ventilation**

We then investigated whether the iSAVE could ventilate two large animals (Yorkshire swine, 74 and 88 kg) as adequately as ventilating each animal on its own closed-circuit ventilator (Fig. 3, A and B) in a three-stage experiment. In stage 1, pigs were ventilated individually. Pig A required Vₜ = 690 ml, whereas pig B required Vₜ = 880 ml. We ventilated each animal for about 45 min under stable EtCO₂ (33 to 38 mmHg), SpO₂ (93 to 99%), desired tidal volumes, and arterial blood gases (Fig. 3, C and D). In stage 2, we ventilated both animals on the same closed-circuit ventilator using iSAVE. Representative individual pressure, flow, and volume traces are provided in Fig. 3E, and the means and SD of 300 breathing cycles are provided in Fig. 3F. No significant differences were observed between stages 1 and 2, and all values were within the physiologically healthy range for swine. The difference in mean tidal volume between stages 1 and 2 was 5 ml for pig A and 2 ml for pig B. The difference in mean driving pressure between stages was 1.8 cm H₂O for pig A and 0.4 cm H₂O for pig B. Blood electrolytes, anion gap, glucose, blood urea nitrogen, hematocrit, and hemoglobin remained stable throughout (table S4). We measured the respiratory mechanics of both animals during both stages of ventilation. During shared ventilation, Pplat was simultaneously determined using the end-inspiratory hold feature for both animals. The computed compliance and resistances (based on the Pplat from the main ventilator) matched those displayed by the individual respiratory monitors.

In the third stage, we ventilated the two animals with differential PEEP’s (pig A, PEEP = 5 cm H₂O; pig B, PEEP = 10 cm H₂O; Fig. 4, A and B) using iSAVE on a closed-circuit ventilator. During ventilation, we simulated several practically challenging scenarios including the following: (i) adding an animal to the iSAVE circuit; (ii) quickly removing an animal from the circuit without leaking the airflow (containing isoflurane) into the atmosphere; (iii) adjusting ventilation parameters as the animal’s respiratory mechanics changed. In all cases, iSAVE enabled stable management of ventilation.

**DISCUSSION**

In this study, we demonstrate how the iSAVE enables individualized management of ventilation using valves, sensors, and alarms. Through benchtop and in vivo testing, we demonstrate that we can not only individualize tidal volume and PEEP but also rebalance ventilation to accommodate changes in respiratory mechanics in one channel that could otherwise jeopardize flow to a second connected channel. The data show that the system can support the flows and pressures required to manage the ventilation of lungs with properties of ARDS for two individuals over several hours. We tested several clinical scenarios associated with multiplexing ventilation to address alarm mechanisms, connection and disconnection processes, mitigation of contamination risk, and measurement of pulmonary mechanics. Moreover, the stability of tidal volume, PEEP, respiratory mechanics (resistance and compliance), and gas exchange (SpO₂ and EtCO₂), as indicated by low SD values during the ventilation of two pigs, suggests that it may be possible to integrate iSAVE in a protective ventilation strategy.

The iSAVE’s pressure release valve, flow control valve, one-way valve, pressure sensor, and CO₂ sensor are commercially available medical-grade parts commonly found in hospitals, lowering the barrier to implementation. In the event of shortages, these parts could be procured from plumbing and ventilation departments in hardware stores and autoclaved for sterile usage. If standard adapters do not interface these parts, then adapters can be made from standard piping or three-dimensional printing. As specified, the iSAVE permits individualized ventilation through alteration in tidal volume for two channels. Future iterations could incorporate closed-loop control, directly modulating flow according to flow meter readings and valves to allow for independent control of RR and FiO₂ in each patient circuit. For ventilators requiring closed-loop flow control, standard flow sensors can be placed in line and rerouted through stopcocks to enable normal functioning of the ventilator’s self-check/calibration mechanisms (fig. S9). In the envisioned setup, individual patient monitors (such as the Philips NM3) would be used to set the initial conditions of the valves, perform periodic checks, and determine changes in the circuit. Standard humidification devices for airstreams would be connected in series with the iSAVE on the inspiratory channel. In addition to the components described here, a whistle ring could be added to the pressure release valve to provide an auditory alarm when high pressure develops in one segment (fig. S10). Independent monitoring mechanisms should also be developed.

Although we tested scenarios involving only two individuals and splitting among two channels under preclinical conditions, in theory, the iSAVE should be able to accommodate more than two channels. Our testing thus far has provided a proof of concept for the iSAVE using a single ventilator with two individuals (1:2 ratio). Ventilator volume can be linearly divided to additional patients. Thus, the maximal capacity of this system is defined by

\[
\text{Capacity} = \frac{V_{T,\text{Ventilator}}}{V_{T,\text{Patient }1} + V_{T,\text{Patient }2} + V_{T,\text{Patient }3} + \ldots}
\]

Eq. 1

Because most ICU ventilators provide up to 2500 ml, it is estimated that at least six individuals could be simultaneously ventilated. However, physical and practical challenges will limit implementation, particularly in the ICU setting. Dead space, the volume of air that does not participate in gas exchange, accumulated in tubing cannot be greater than the lowest individual tidal volume required. Further, with the addition of more than two patients to one ventilator circuit, determining flow changes and titration will entail a more involved process, requiring monitors for each patient. In the case of n patients, data from n – 1 circuits would be required to make adjustments. If the supply of respiratory monitors is constrained, then this limitation could potentially be overcome by sharing monitors among patients and using monitors to perform frequent checks.

On the basis of the iSAVE’s individualization capacity, matching criteria for patients may not need to be as stringent as other protocols designed to multiplex ventilation without individualized control (10). Nevertheless, certain ventilatory parameters (RR, FiO2, and I:E) will remain shared among patients. In addition, patients need to remain sedated (and/or paralyzed) to prevent spontaneous breathing, which would lead to asynchrony in the system. Thus, patients should be matched as closely as possible in terms of degree of illness and ventilatory needs to optimize functioning. A list of recommendations for patient stratification is provided in table S5. These guidelines were derived from the highest bounds of variance tolerable by the iSAVE from our in vitro and preclinical testing, with a safety factor of 2 to 3, in combination with feedback from standardized treatment protocols (12, 14). Other considerations include hemodynamic stability, anticipated invasive ventilation time, proning, coinfection, and the logistics of space allocation for patients. Ideally, patients would be ventilated in negative pressure rooms, with their heads placed as close as possible to the ventilator. Recovering patients would need to be transitioned to an individual ventilator when spontaneous breathing becomes viable (11).

Fig. 3. Ventilation of two pigs on the iSAVE. (A) Experimental setup for stages 2 and 3 of shared ventilation of pig A (74 kg) and pig B (88 kg) with iSAVE using closed-circuit ventilation. (B) Photograph of the experimental setup. Pressure, flow, and volume waveforms for (C) pig A ventilated individually (stage 1), (D) pig B ventilated individually (stage 1), and (E) pigs A and B ventilated together on the iSAVE (stage 3). (F) Table summarizing ventilatory and respiratory parameters and arterial blood gasses for (C) to (E). Means ± SD were calculated from 300 breathing cycles. No significant differences were found between the individual and shared ventilation approaches (homoscedastic two-tailed t test, P > 0.05).
This study only tested the iSAVE on test lungs and in a small number of large animals without active disease pathology. Further, ventilation was performed for a short period of time in vivo to serve as a proof of concept. Our overall goal is to ensure that the iSAVE can be reliably implemented and used across intensive care clinical settings to address ventilator shortages. To this end, further steps must be taken to address current limitations before clinical implementation. Because of differences in performance characteristics (mechanism of pressure/flow monitoring) of ICU ventilators, this approach must be tested across a range of ventilators. The iSAVE must be evaluated under conditions reflecting the real-life variability of intensive care practice. The procurement and sterilization processes for nonstandard components must be addressed. Toward facilitating this evaluation by our team and others, we have assembled a list of components, instructions for assembly, links to three-dimensional printable adaptors (barb, push-to-connect, or luer lock fittings), and clinical consideration guidelines (https://i-save.mit.edu/). Modules to train personnel in using this system should also be developed. Although this system mitigates several challenges associated with splitting ventilation, it has several drawbacks and unknown limitations and does not fully address safety concerns associated with this and other approaches for ventilator splitting. Ventilator sharing is strongly discouraged by critical care practitioners due to safety concerns. In a setting of severe shortage of ventilators, sharing a ventilator among two patients could potentially mitigate the need to triage which patients receive ventilatory support and which do not. This may present societal value in comparison to potentially losing or prioritizing one life. Further ethical and policy-based discussions are necessary before implementation. The authors caution that the iSAVE approach is not a standard of clinical care and that this is a preclinical study. Rigorous further testing and validation of this approach are necessary before implementation could be considered under the Emergency Use Authorization issued by the U.S. Food and Drug Administration (13). Here, we have demonstrated that the iSAVE provides a solution for expansion of respiratory support using readily available medical-grade materials and existing ventilators. The iSAVE can at least double the existing ventilator capacity while retaining personalized ventilation settings for two individuals. Health care systems worldwide could potentially benefit from this system as they strive to care for the increasing volume of patients with ARDS associated with COVID-19 infection.

### MATERIALS AND METHODS

#### Study design

The goal of this study was to design and validate a system using medical-grade filters, valves, and sensors to provide individualized ventilation using one ventilator shared among at least two individuals. We performed benchtop testing of the iSAVE using linear test lungs to ensure its ability to provide differential ventilation, accommodate for changes in respiratory mechanics, and manage acute changes to patient status. We also evaluated issues of cross contamination, adding and removing individuals from the circuits, and circuit design as they related to specific ventilator models. We performed ventilation of one pig and one test lung, wherein we modulated the respiratory mechanics of each lung to test the management of rebalancing differential ventilation to both lungs. We then performed the simultaneous ventilation of two pigs to evaluate the iSAVE’s performance in an in vivo setting, simulating several clinical scenarios. Animal experiments were approved by the Committee on Animal Care at the Massachusetts Institute of Technology (MIT). No blinding or randomization was performed as this was a proof-of-concept study.

#### Open-circuit ventilator assembly

A Philips Trilogy portable ventilator was used as a representative open-circuit ventilator for testing. Standard flex corrugated tubing (22-mm outer diameter) was used to connect two respiratory circuits, each consisting of a (i) ball valve (EKWB G ¼” Nickel,
Microcenter), (ii) bacterial/viral filter (Main Flow Bacterial/Viral Filter, Teleflex Medical), (iii) pressure sensor (TD160D, Biopac Systems), (iv) airflow sensor (BSL Medium Airflow Transducer SS11LA and AFT 20, Biopac Systems), and (v) Whisper Swivel valve (passive exhalation port, Philips). This was connected to the expiratory side of a Y piece to allow for the inclusion of a bacterial/viral filter before venting exhaled gas to the ambient. See the circuit diagram provided in Fig. 1A. The airflow and pressure sensors were connected to DA100C differential amplifiers with 1000× gain and processed through the MP160WSW signal processing unit (Biopac Systems), sampling at 2 kHz. Volume control mode was used, and settings were adjusted to deliver the desired $V_T$, RR, and PEEP. Artificial linear test lungs (IngMar Medical) were used for simulations with the open-circuit ventilator.

Closed-circuit ventilator assembly
The iSAVE is designed to work with closed-circuit ventilators currently found in ICUs. To this end, the system was tested on a Philips VX850 ventilator, a Hamilton G5 ventilator, and a Puritan Bennett 840 ventilator (fig. S1). These ventilators were chosen because they represent the most common brands used in the United States. Y connectors were connected to the inspiratory and expiratory limbs of the ventilator to enable individual channels for each lung. In the inspiratory limbs, the following components were connected in series: (i) a bacterial/viral filter (Main Flow Bacterial/Viral Filter, Teleflex Medical), (ii) ball valve (EKWB G ¼” Nickel, Microcenter), (iii) one-way valve (a Threshold PEP device, Philips, was used as a surrogate for one-way valves), (iv) pressure sensor, (v) flow sensor, and (vi) capnastat adapter. In the expiratory limb, the following components were connected in series: (i) a bacterial/viral filter, (ii) a pressure release valve (PEEP valve), and (iii) an optional one-way valve before connection with the ventilator (Threshold PEP). Polytetrafluoroethylene tubing or rubber adapters were used to adapt the ball valves to the standard 2-mm–outer diameter corrugated flex tubing. A dual adult lung simulator (Model 5600i, Michigan Instruments) was used to perform simulations with the closed-circuit ventilator. Pressure and flow were displayed and recorded by the Philips NM3 monitors and associated sensors, one for each channel. For ventilators requiring continuous flow measurements for calibration and closed-loop control, such as the Hamilton G5, the standard flow sensors can be reconfigured using stopcocks as elaborated in fig. S9.

Setup and testing protocol
The following main ventilator settings were selected: (i) Volume control mode is selected with triggering turned off; (ii) RR is determined on the basis of the minute ventilation needs for both sets of lungs; (iii) FiO₂ is set to the desired value to be shared by both patients; (iv) I:E ratio is set with consideration of the tau (τ = RC) parameter of each patient, which must initially be estimated for each patient. The longer τ will dictate the expiratory window and prevent autoPEEP; (v) the alarm for minute volume was set to the following lower and upper bounds [10% of RR($V_T$ + $V_T$), 10% of RR($V_T$ + $V_T$)]. Next, we checked for leaks and alarms by first testing the ability of the system to perform ventilation without leaks. We then disconnected a lung to ensure the activation of standard alarms on the ventilator. To produce variable tidal volumes, starting with both flow valves fully open, we measured the flow and pressure delivered to each lung. Then, we gradually closed one of the valves, measuring the distribution of flow, to map the range of volume distribution capable of the system. To maintain $V_T$ after static changes to compliance and resistance, we modulated the compliance and/or resistance of one test lung, measured the effect of the intervention, and titrated the valve until the baseline parameters were reached. Resistors (Rp5, Rp20, Rp50, and Rp500, Michigan Instruments) were used for all benchtop testing. We performed these tests with baseline parameters set to those of (i) a healthy lung ($C = 50$ cm H₂O, $R = Rp5$) and (ii) a lung with ARDS ($C = 20$ cm H₂O, $R = Rp5$ to Rp50). The flow valve was the main variable we modulated, but, in some cases, the ventilator’s tidal volume or I:E ratio was also adjusted. These cases are specifically mentioned in Results. Last, to maintain desired ventilation after dynamic changes to compliance and resistance, we simulated acute changes by disconnecting or clamping tubes and monitoring the ventilator’s response. The flow valves were also titrated to return the circuit to its baseline.

Cross-contamination testing
Five milliliters of trypan blue (0.4%; 15250061, Thermo Fisher Scientific) was placed in a nebulizer attached to the patient inflow/outflow segment of one of two circuits containing a test lung. Air from a separate line was used to nebulize droplet particles into the main tubing. We then performed tests ranging the following parameters, including the maximal combination designed to create the most turbulent flow conditions: valves, fully open to fully closed; airflow pressure used to nebulize, 1 to 2800 cm H₂O; duration of nebulization, 1 to 10 min; $V_T$ setting, 100 to 900 ml per lung; PEEP, 2 to 10 cm H₂O; RR, 2 to 30 bpm. After each test, filters were manually inspected for contamination, and a wipe test was performed in each of the segments of tubing. Each wipe was then incubated in deionized H₂O for 5 min. One milliliter from each sample was used to perform ultraviolet-visible absorption spectroscopy at a wavelength of 580 nm. Each sample was measured in triplicate.

In vivo testing
The first experiment was performed as part of a terminal procedure on a 70-kg female swine ($n = 1$). The iSAVE was connected to a veterinary anesthesia ventilator (Model 2001E, Hallowell EMC) delivering 2% isoflurane in oxygen. One inspiratory circuit was connected to the anesthesia machine delivering gas to the animal, while the other inspiratory circuit was connected to an artificial linear test lung (IngMar Medical). Pressure and flow measurements were recorded on the inspiratory limb of the animal. A VetTrends Vital Signs Monitor was used to measure $SpO_2$, ETCO₂, RR, and other physiological parameters. $V_T$ (600 ml) was equally distributed between the animal and test lung. An RR between 18 and 20 per minute was set on the ventilator. We carried out the same tests outlined in the benchtop testing protocol, modulating the parameters of the test lung to validate the capabilities of the iSAVE to restore the system to baseline. After the animal was euthanized, to acutely change the compliance of the pig’s lung, we used an endoscope (Pentax) to deliver 750 ml of phosphate-buffered saline (Sigma–Aldrich) into the left and right bronchi. Ventilation was performed, and the valves were titrated to achieve desired flow parameters.

In the second experiment, one 74-kg and one 88-kg female swine were used in a survival approach. These pigs were sedated with midazolam (0.25 mg/kg, 5 mg/ml) and dexmedetomidine (0.03 mg/kg, 0.5 mg/ml). The experiment was divided into three stages: individual ventilation of each animal (PEEP = 0) (stage 1); iSAVE ventilation (differential $V_T$, PEEP = 0) (stage 2); and iSAVE ventilation (differential $V_T$, differential PEEP) (stage 3). Ventilation was performed...
with the Philips Respironics Esprit ventilator in series with the anesthesia machine. The target tidal volume for each animal was calculated at 10 ml/kg, although ventilation was optimized to maintain an SpO2 of >94% and an EtCO2 of ~30 to 35 mmHg in all stages. The PEEP setting on the main ventilator was set to 0 cm H2O. After 30 min of stable ventilation, arterial blood gases were measured using an i-STAT handheld blood analyzer (Abbott). Respiratory flow, pressure, and volume were continuously logged using the Philips NM3 monitors and the Biopac Systems described above. Reported values represent the mean and SDs of 300 breathing cycles. In stage 3, the PEEP settings were changed by adjusting the PEEP valve to enable the animals to be ventilated with differential PEEP values (pig A, PEEP = 5 cm H2O; pig B, PEEP = 10 cm H2O). Compliance and resistance were measured for each animal by performing an end-inspiratory pause of 1 s to ensure that the iSAVE enabled these measurements. The computed compliance and resistance values matched those that were measured on the monitor. The duration of each stage lasted about 45 min.

**Data and statistical analysis**

Pressure and flow data from the Biopac System was analyzed using MATLAB (2018a). Flow data were integrated every respiratory cycle to derive the flow. In cases where pressure and flow data were not simultaneously obtained from both limbs of the circuit, data were derived on the basis of the settings on the main ventilator and the measurements on one limb. Homoscedastic, two-tailed t tests were performed to assess significance from the data from the contamination testing with a threshold of P < 0.05. A homoscedastic, two-tailed t test was used to compare the values from 300 breathing cycles between the two conditions for the in vivo study with a threshold of P < 0.05.

**SUPPLEMENTARY MATERIALS**

stm.sciencemag.org/cgi/content/full/12/549/eabb9401/DC1

Fig. S1. Photographs of the iSAVE setup.

Fig. S2. Differential tidal volume and PEEP delivery on the iSAVE using an open-circuit ventilator and test lungs.

Fig. S3. Accommodation to changes in compliance of one test lung using the iSAVE and an open-circuit ventilator.

Fig. S4. Ventilation at high lung resistances.

Fig. S5. Ventilator alarm response to occlusion.

Fig. S6. Adding a test lung to the circuit.

Fig. S7. Cross-contamination validation using artificial lungs on a closed-circuit ventilator.

Fig. S8. Individualized management of ventilation using the iSAVE on a pig lung and a test lung.

Fig. S9. Modification of sensing circuit for a Hamilton GS ventilator.

Fig. S10. Whistle ring designs for two types of common pressure release (PEEP) valves.

Table S1. List of components required for the assembly of the iSAVE.

Table S2. Mechanical components used in the iSAVE and their readily available medical industry equivalents.

Table S3. Measurement of respiratory mechanics for iSAVE system using test lungs.

Table S4. Blood electrolytes and chemistry during ventilation in pigs.

Table S5. Stratification for patient matching.

View/request a protocol for this paper from Bio-protocol.

**REFERENCES AND NOTES**


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**Author contributions:** S.S.S. performed the conceptualization, investigation, data analysis, and writing. A.H. and D.L. assisted with in vivo test setup and flow, pressure, and volume data analysis. A.H. performed the conceptualization and editing of the manuscript. K.B.R. contributed to the conceptualization and writing of the manuscript. F.V. performed the investigation and data curation and edited the manuscript. D.G. assisted with prototyping. R.L. and J.J.F., D.G., R.M.B., and G.T. contributed to the conceptualization and editing of the manuscript. All authors approved the manuscript. Competing interests: F.V. and J.J.F. serve as employees for Philips North America, a maker of health care devices and monitoring solutions including patient ventilators. R.M.B. is part of an advisory board for Merck. Complete details of all relationships for profit and not for profit for G.T. can be found at the following link: https://dropbox.com/sh/szii7vnr4a2aj56s/AAo5sN5sDQ9AT11qIAE-T5a7d1l?fbclid=IwAR3EMFwFYtEzAI0e0B40QD102l8kT85Wz90ID12fjWz90f8MzDy12fG4fKc. All other authors declared that they have no competing interests related to the work reported here.

Data and materials availability: All data associated with this study are in the paper or the Supplementary Materials. Further information including videos and details on components, assembly, clinical considerations, partnerships, and the implementation can be found at https://i-save.mit.edu. This work is licensed under a Creative Commons Attribution 4.0 International (CC BY 4.0) license, which permits unrestricted use, distribution, and reproduction in any medium, provided that the original work is properly cited. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/. This license does not apply to figures/photos/artwork or other content included in the article that is credited to a third party and obtain authorization from the rights holder before using such material.

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A rapidly deployable individualized system for augmenting ventilator capacity
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Divide and conquer
Ventilators, machines that move air into and out of lungs, can be shared among patients in emergency situations, although this practice poses safety concerns. Srinivasan et al. developed a system of tubing, sensors, filters, and valves to provide individualized volume and pressure control to two or more individuals from a shared ventilator. The system was applied to open- and closed-circuit ventilators, provided temporary respiratory support to two artificial test lungs or pigs with differing pulmonary compliance and resistance, measured pulmonary mechanics, and accommodated sudden flow changes or the addition or removal of an individual from shared ventilation. Use of readily available materials enables the rapid deployment of this system.