

## MARFAN SYNDROME

## Oxytocin antagonism prevents pregnancy-associated aortic dissection in a mouse model of Marfan syndrome

Jennifer Pardo Habashi<sup>1\*</sup>, Elena Gallo MacFarlane<sup>2\*</sup>, Rostam Bagirzadeh<sup>2</sup>, Caitlin Bowen<sup>2</sup>, Nicholas Huso<sup>2</sup>, Yichun Chen<sup>2</sup>, Djahida Bedja<sup>3</sup>, Tyler J. Creamer<sup>4</sup>, Graham Rykiel<sup>2</sup>, Maurice Manning<sup>5</sup>, David Huso<sup>3†</sup>, Harry C. Dietz<sup>2,6‡</sup>

Copyright © 2019  
The Authors, some  
rights reserved;  
exclusive licensee  
American Association  
for the Advancement  
of Science. No claim  
to original U.S.  
Government Works

Women with Marfan syndrome (MFS) are at high risk for pregnancy-associated aortic dissection. Pathogenic models that singularly invoke hemodynamic stress are difficult to reconcile with predominant postnatal occurrence of aortic tear, often occurring weeks to months after delivery. In consideration of events that peak at term, are sustained after delivery, and might synergize with previously defined signaling pathways implicated in aneurysm progression, we examined the hormone oxytocin, which initiates uterine contraction and milk letdown for the duration of lactation through phosphorylation of extracellular signal-regulated kinase (ERK). In a mouse model of MFS that shows highly penetrant postnatal aortic dissection, risk was strongly attenuated by preventing lactation or use of an oxytocin receptor antagonist. Survival correlated inversely with the extent of ERK activation in the aortic wall, and strong protection was observed upon attenuation of ERK phosphorylation using an inhibitor of ERK kinase (MEK) or the U.S. Food and Drug Administration–approved medication hydralazine, offering potential therapeutic strategies for pregnancy-associated vascular catastrophe in the setting of MFS.

## INTRODUCTION

Marfan syndrome (MFS) is an autosomal dominant connective tissue disorder that affects about 1 in 5000 people worldwide and is caused by mutations in the fibrillin-1 gene (*FBN1*) encoding the extracellular matrix protein fibrillin-1. Cardinal manifestations occur in the ocular, musculoskeletal, and cardiovascular systems, including dislocation of the lens of the eye, long bone overgrowth, scoliosis, and progressive aortic root enlargement resulting in vessel tear (dissection) and death. The current recommendation is to perform aortic root replacement once the aorta reaches an absolute diameter of 5.0 cm (1). There is an increased risk of accelerated aneurysm progression and aortic dissection associated with pregnancy in MFS once the maximal aortic diameter exceeds 4.0 cm; however, precise risk counseling has been difficult because of the small number and size of studies that address this issue (2–7). Contrary to intuition and the commonly invoked mechanistic hypothesis highlighting the importance of hemodynamic stress, most of the pregnancy-associated catastrophic events do not occur during labor (8). Although the incidence of dissection rises in the third trimester, it peaks during the first few weeks postpartum. One study by Goland and Elkayam (2) estimated the overall risk of aortic dissection during pregnancy in MFS to be 3%, but only about 1% when the diameter of the aortic root was <4.0 cm around the time of conception. The risk was estimated at 10% if the aortic diameter measured  $\geq 4.0$  cm. Pacini and colleagues (6) found that the risk of aortic dissection with each pregnancy in MFS was 4.4%, and that the time of highest risk extends from the late third trimester to several

weeks to months postpartum. A more recent study examined the incidence of aortic complications during pregnancy in 184 women with an established diagnosis of MFS: 10.6% experienced an aortic-related complication during pregnancy, and 75% of dissections occurred in the postpartum period (8). With the advent of valve-sparing aortic root replacement (precluding the need for chronic anticoagulation), some women choose surgery before conception with aortic dimensions below conventional surgical thresholds to mitigate pregnancy-associated risks. However, Sayama and colleagues (9) followed 14 women (17 pregnancies) with confirmed MFS and reported an increased rate of descending thoracic (type B) aortic dissections in association with pregnancy in those women who had undergone elective aortic root replacement (60%) versus those who had not (8.3%). This demonstrates the ongoing need for improved preventative treatment options for women with MFS during pregnancy.

Aspects of the pathogenesis of aneurysm growth and tear have been elucidated in MFS. A primary deficiency of fibrillin-1 in the extracellular matrix associates with increased transforming growth factor  $\beta$  (TGF $\beta$ ) signaling in the aortic media in both patients and mouse models, as evidenced by increased activation of receptor-activated SMAD proteins and increased expression of TGF $\beta$ -responsive genes (10). Furthermore, treatment of MFS mice with a TGF $\beta$ -neutralizing antibody (TGF $\beta$ NAb) after the immediate perinatal period can slow aortic root growth and diminish elastic fiber fragmentation and excessive collagen deposition in the aortic wall (11–14). Losartan, an angiotensin II type 1 (AT1) receptor blocker (ARB) that both lowers blood pressure and attenuates TGF $\beta$  expression, activation, and signaling, achieved cessation of aneurysm progression, preservation of aortic wall architecture, and delay or even prevention of aortic dissection in Marfan mice (11). More recently, activation (phosphorylation) of extracellular signal-regulated kinase 1/2 (pERK1/2) has been shown to be a prominent distal determinant of aneurysm progression in the MFS mouse model; a selective antagonist of the kinase that phosphorylates ERK1/2 (MEK) was as effective as losartan in preventing aneurysm growth. Losartan both reduces ERK activation by AT1 and augments inhibition of ERK by the type 2 angiotensin receptor

<sup>1</sup>Division of Pediatric Cardiology, Johns Hopkins University, Baltimore, MD 21287, USA. <sup>2</sup>Center for Medical Genetics, Johns Hopkins University, Baltimore, MD 21287, USA. <sup>3</sup>Division of Comparative Medicine, Johns Hopkins University, Baltimore, MD 21287, USA. <sup>4</sup>Department of Surgery, Johns Hopkins University, Baltimore, MD 21287, USA. <sup>5</sup>Department of Cancer Biology, University of Toledo College of Medicine, Toledo, OH 43614, USA. <sup>6</sup>Howard Hughes Medical Institute, Chevy Chase, MD 20815, USA.

\*These authors contributed equally to this work.

†Deceased.

‡Corresponding author. Email: hdietz@jhmi.edu

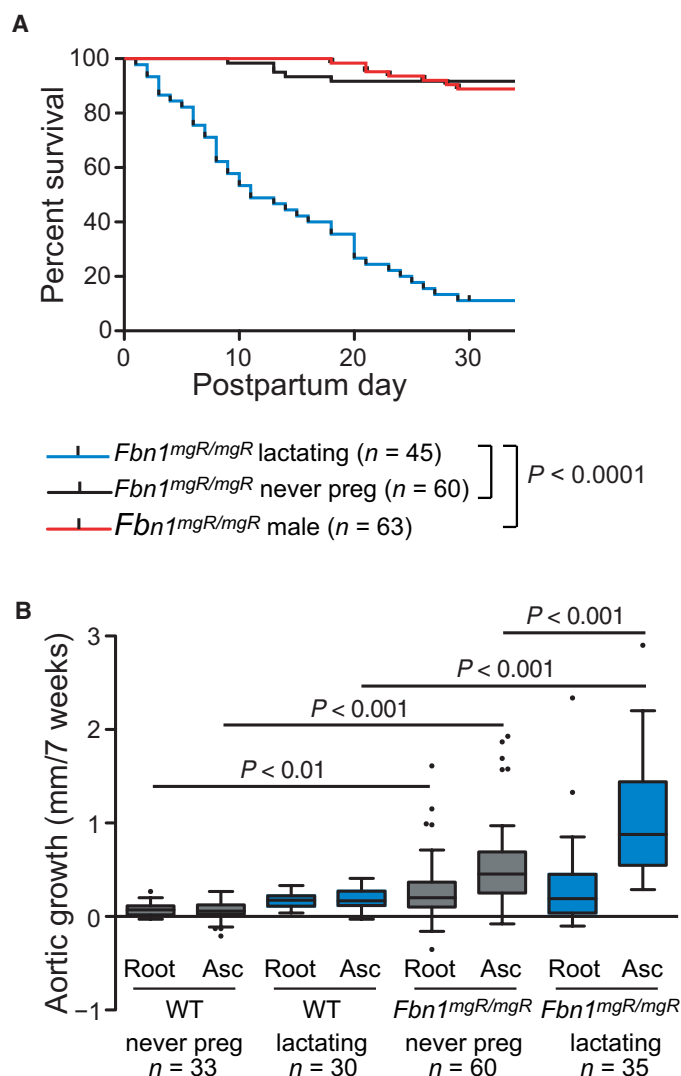
(AT2) (12). Pathologic ERK1/2 activation was also shown, at least in part, to be dependent upon TGF $\beta$  signaling in MFS mice as it was diminished in the aortic wall upon treatment with TGF $\beta$ Ab (12–14). ERK1/2 activation was subsequently linked to a wide variety of genetically induced presentations of aortic aneurysm (15) and to environmental provocations that accelerated aneurysm progression and dissection in MFS mice. Together, these data suggest that the predisposition for aortic aneurysm growth and tear can integrate the influence of diverse etiologies for ERK1/2 activation.

In consideration of putative pathogenic events that would initiate toward the end of pregnancy, be maintained after delivery, and might synergize with signaling events previously implicated in aortic disease, our focus turned to the hormone oxytocin. Oxytocin release by the hypothalamus increases throughout pregnancy, particularly in the late third trimester, and contributes to the initiation of uterine contraction (16). High circulating oxytocin is sustained postnatally in association with lactation to promote milk letdown (17). The expression of oxytocin receptor (OR) in the aorta is enhanced by estrogen and pregnancy (18), and oxytocin exerts its effects on peripheral tissues including the uterus and breast through ERK1/2 activation (19–21). On this basis, we hypothesized that oxytocin mediates the enhanced pregnancy-associated risk of aortic dissection in MFS and related disorders.

## RESULTS

### Decreased survival and accelerated aortic growth in pregnant MFS mice

The *Fbn1*<sup>mgR/mgR</sup> mouse is an exaggerated homozygous model of MFS, in which there is a severe deficiency of fibrillin-1 expression (~15% of normal) early in development and throughout postnatal life due to transcriptional interference imposed by targeted insertion of a neomycin resistance cassette in the *Fbn1* gene. Unlike people and mice with MFS that harbor heterozygous *Fbn1* mutations, where the strongest predisposition for aneurysm is at the sinuses of Valsalva (the aortic root), *Fbn1*<sup>mgR/mgR</sup> mice show predominant dilatation in the more distal ascending aorta, culminating in ascending aortic dissection at ~2 to 8 months of age (22). To investigate the impact of pregnancy on survival, *Fbn1*<sup>mgR/mgR</sup> females were bred at 7 to 9 weeks of age and monitored for survival. All mice found dead demonstrated hemothorax, with the majority also demonstrating hemopericardium, suggesting a predominance of type A (ascending) aortic dissections. We found an extreme risk of pregnancy-associated death from dissection, with 91.1% of *Fbn1*<sup>mgR/mgR</sup> mice ( $n = 45$ ) dying from aortic dissection within the first 4 weeks postpartum (median age of death was 11 days postpartum) as compared to 6.7% of age-matched never-pregnant *Fbn1*<sup>mgR/mgR</sup> females ( $n = 60$ ;  $P \leq 0.0001$ ) and 9.5% of males ( $n = 63$ ;  $P \leq 0.0001$ ) followed over the same age-matched time period. There was no difference in survival between never-pregnant *Fbn1*<sup>mgR/mgR</sup> females and corresponding males (Fig. 1A). In comparison, no wild-type (WT) females ( $n = 60$ ) or lactating WT mice ( $n = 45$ ) died during the same age-matched time course. We followed mice by echocardiography over the 7-week time course spanning pregnancy (3 weeks) and lactation (4 weeks) and found a significant increase in aortic root growth and ascending aortic growth in never-pregnant *Fbn1*<sup>mgR/mgR</sup> female mice as compared to never-pregnant WT mice ( $0.07 \pm 0.07$  versus  $0.29 \pm 0.32$  mm/7 weeks,  $P \leq 0.01$ , and  $0.48 \pm 0.48$  versus  $0.07 \pm 0.12$  mm/7 weeks,  $P \leq 0.001$ , respectively). Whereas there was no pregnancy-associated effect on



**Fig. 1. The effects of pregnancy in WT and *Fbn1*<sup>mgR/mgR</sup> mice.** (A) Kaplan-Meier survival curve for lactating *Fbn1*<sup>mgR/mgR</sup> mice ( $n = 45$ ) compared to male ( $n = 63$ ) or never-pregnant female ( $n = 60$ ) *Fbn1*<sup>mgR/mgR</sup> mice. (B) Average aortic root and ascending aortic growth over the 7-week period spanning pregnancy (3 weeks) and lactation (4 weeks) in WT never-pregnant ( $n = 33$ ), WT lactating ( $n = 30$ ), *Fbn1*<sup>mgR/mgR</sup> never-pregnant ( $n = 60$ ), and *Fbn1*<sup>mgR/mgR</sup> lactating ( $n = 35$ ) mice, as measured by echocardiogram. Survival was statistically evaluated using a log-rank (Mantel-Cox) test. When data are presented as boxplots, the box extends from the 25th to 75th percentiles, median is denoted by the internal line, and whiskers indicate the range calculated using the Tukey method, with data points outside the whiskers shown as individual points. All significant  $P$  values ( $P < 0.05$ ) are noted in the figure.

aortic root growth, there was a significant pregnancy-associated increase in ascending aortic growth in *Fbn1*<sup>mgR/mgR</sup> females ( $1.02 \pm 0.61$  mm/7 weeks) compared to never-pregnant *Fbn1*<sup>mgR/mgR</sup> females ( $P \leq 0.001$ ) and postpartum WT mice ( $0.19 \pm 0.11$  mm/7 weeks,  $P \leq 0.001$ ) (Fig. 1B and table S1).

### Survival improved by prevention of lactation

To determine whether lactation played a role in pregnancy-associated aortic dissection, we removed *Fbn1*<sup>mgR/mgR</sup> mothers from their pups on the day of delivery, effectively eliminating lactation-induced oxytocin

release in the postpartum period. This isolated manipulation reduced the incidence of pregnancy-associated death due to aortic dissection from 91.1 to 25.9% ( $P \leq 0.0001$ ) in *Fbn1*<sup>mgR/mgR</sup> mice (Fig. 2A). In addition, although the growth of the aortic root ( $0.15 \pm 0.21$  mm/7 weeks) was not significantly decreased, growth of the ascending aorta ( $0.54 \pm 0.43$  mm/7 weeks) was significantly less during the 7-week time period spanning pregnancy and the first 4 weeks postpartum in *Fbn1*<sup>mgR/mgR</sup> mice not allowed to lactate ( $P \leq 0.01$ ) (Fig. 2B and tables S1 and S2). Evaluation of the proximal ascending aorta with Verhoeff–van Gieson (VVG) staining demonstrated increased elastic fiber fragmentation in *Fbn1*<sup>mgR/mgR</sup> mice compared to WT controls. Marked thickening of the adventitia with an evident fibroproliferative response was observed in lactating *Fbn1*<sup>mgR/mgR</sup> mice and was greatly reduced by preventing lactation (Fig. 2C).

### Survival improved by OR antagonist

To determine whether reduction of oxytocin was responsible for the improved survival resulting from prevention of lactation, we treated *Fbn1*<sup>mgR/mgR</sup> pregnant mice with a continuous infusion of a highly specific OR antagonist [OTA; desGly-NH<sub>2</sub>-d(CH<sub>2</sub>)<sub>5</sub>[D-Tyr<sup>2</sup>,Thr<sup>4</sup>]OVT (ornithine vasotocin)], which has 95 times greater specificity for the OR compared to the vasopressin receptor (23). Mini-osmotic pumps were implanted subcutaneously in the mice at the beginning of the third trimester and delivered OTA for the remaining 1 week of pregnancy and first 4 weeks postpartum. We observed a reduction in death from aortic dissection from 91.1 to 6.7% in treated mice ( $P \leq 0.0001$ ) (Fig. 2D). The administration of OTA resulted in an incomplete antagonism of oxytocin, as evidenced by the retained ability of the mice to deliver and to support their pups through lactation. In comparison, genetically targeted mice that are completely deficient in oxytocin are still able to deliver normally, but are not able to successfully nurse their pups due to lack of milk letdown (24). There was no significant difference in aortic root growth ( $0.32 \pm 0.20$  mm/7 weeks) in OTA-treated pregnant *Fbn1*<sup>mgR/mgR</sup> mice when compared to either never-pregnant or lactating *Fbn1*<sup>mgR/mgR</sup> animals; however, ascending aortic growth ( $0.44 \pm 0.49$  mm/7 weeks) was significantly decreased compared to lactating *Fbn1*<sup>mgR/mgR</sup> mice ( $P \leq 0.01$ ) and was no different than that observed in never-pregnant *Fbn1*<sup>mgR/mgR</sup> females (Fig. 2E and tables S1 and S2). No protection was observed in mice receiving osmotic pumps delivering vehicle alone (fig. S1), and OTA delivery was not associated with a reduction in blood pressure (fig. S2 and table S3).

### Administration of oxytocin agonist not sufficient to induce dissection

To determine whether oxytocin is sufficient to induce aortic dissection, we first administered a continuous infusion of a highly specific oxytocin agonist [OA; (Thr<sub>4</sub>,Gly<sub>7</sub>)OT] (23) via mini-osmotic pumps to age-matched never-pregnant *Fbn1*<sup>mgR/mgR</sup> females. This did not significantly accelerate dissection (12.2 versus 8.9% died from aortic dissection,  $P = 0.20$ ) (Fig. 2F). We then administered OA to pregnant *Fbn1*<sup>mgR/mgR</sup> mice at the beginning of the third trimester and continued for 5 weeks. There was no significant difference in the incidence of death from aortic dissection or the timing of dissection over the 5-week time period compared to vehicle-treated lactating animals (90.0 versus 91.1%,  $P = 0.37$ ) (Fig. 2G). We went on to evaluate OR gene (*Oxtr*) expression in the aorta and observed increased *Oxtr* mRNA during pregnancy in both WT and *Fbn1*<sup>mgR/mgR</sup> mice compared to their never-pregnant counterparts ( $P < 0.05$ ), with no sig-

nificant difference noted between never-pregnant WT and *Fbn1*<sup>mgR/mgR</sup> mice and between lactating WT and *Fbn1*<sup>mgR/mgR</sup> mice ( $P = 0.69$ ) (fig. S3 and table S4), suggesting a pregnancy-specific effect independent of genotype.

### Treatment with $\beta$ -blockers, the current standard of care for pregnancy in MFS

$\beta$ -Adrenergic receptor blockers such as atenolol or propranolol have been shown to achieve a suppression in aortic root growth rate in a different mouse model of MFS harboring a missense mutation, *Fbn1*<sup>C1039G/+</sup> (11), and are currently considered standard of care for pregnant women with MFS (2,3). Pregnant *Fbn1*<sup>mgR/mgR</sup> mice were treated with propranolol in their drinking water starting at the beginning of the third trimester and continuing for 5 weeks. Propranolol treatment improved survival from 9 to 57% ( $P \leq 0.05$ ) in lactating *Fbn1*<sup>mgR/mgR</sup> mice (Fig. 3A), but did not result in a significant reduction in either aortic root ( $0.24 \pm 0.23$  mm/7 weeks) or ascending aortic growth ( $0.99 \pm 0.80$  mm/7 weeks), as compared to placebo-treated lactating *Fbn1*<sup>mgR/mgR</sup> animals (Fig. 3B and tables S1 and S2).

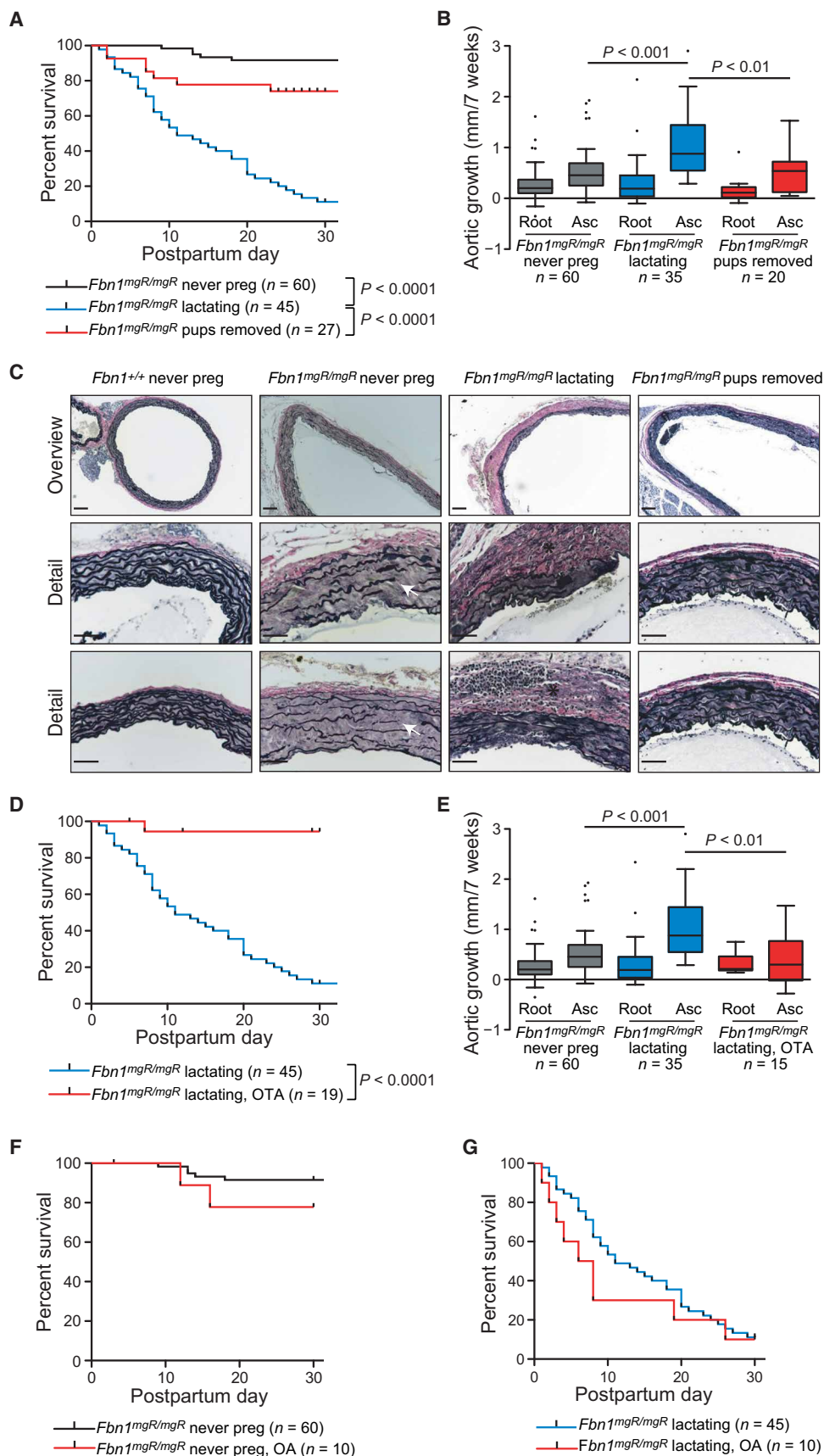
### Differential effect on survival of propranolol versus hydralazine in pregnancy

We went on to treat with hydralazine, another antihypertensive agent approved for use in pregnancy that was previously demonstrated to markedly reduce aortic root growth in the *Fbn1*<sup>C1039G/+</sup> mouse model (25). Treatment of *Fbn1*<sup>mgR/mgR</sup> mice, again initiated at the beginning of the third trimester and continued for 5 weeks, resulted in 95% freedom from aortic dissection, which was significantly greater than in untreated pregnant *Fbn1*<sup>mgR/mgR</sup> mice ( $P \leq 0.0001$ ). There was no significant difference in the performance of hydralazine when compared to propranolol ( $P = 0.10$ ) (Fig. 3A). In contrast to propranolol, hydralazine significantly reduced ascending aortic growth rate ( $0.59 \pm 0.28$  mm/7 weeks) as compared to untreated lactating females ( $P \leq 0.01$ ; Fig. 3B and tables S1 and S2). Both propranolol and hydralazine resulted in a decline in blood pressure, and there was no significant difference in reduction between the two (Fig. 3C and table S5).

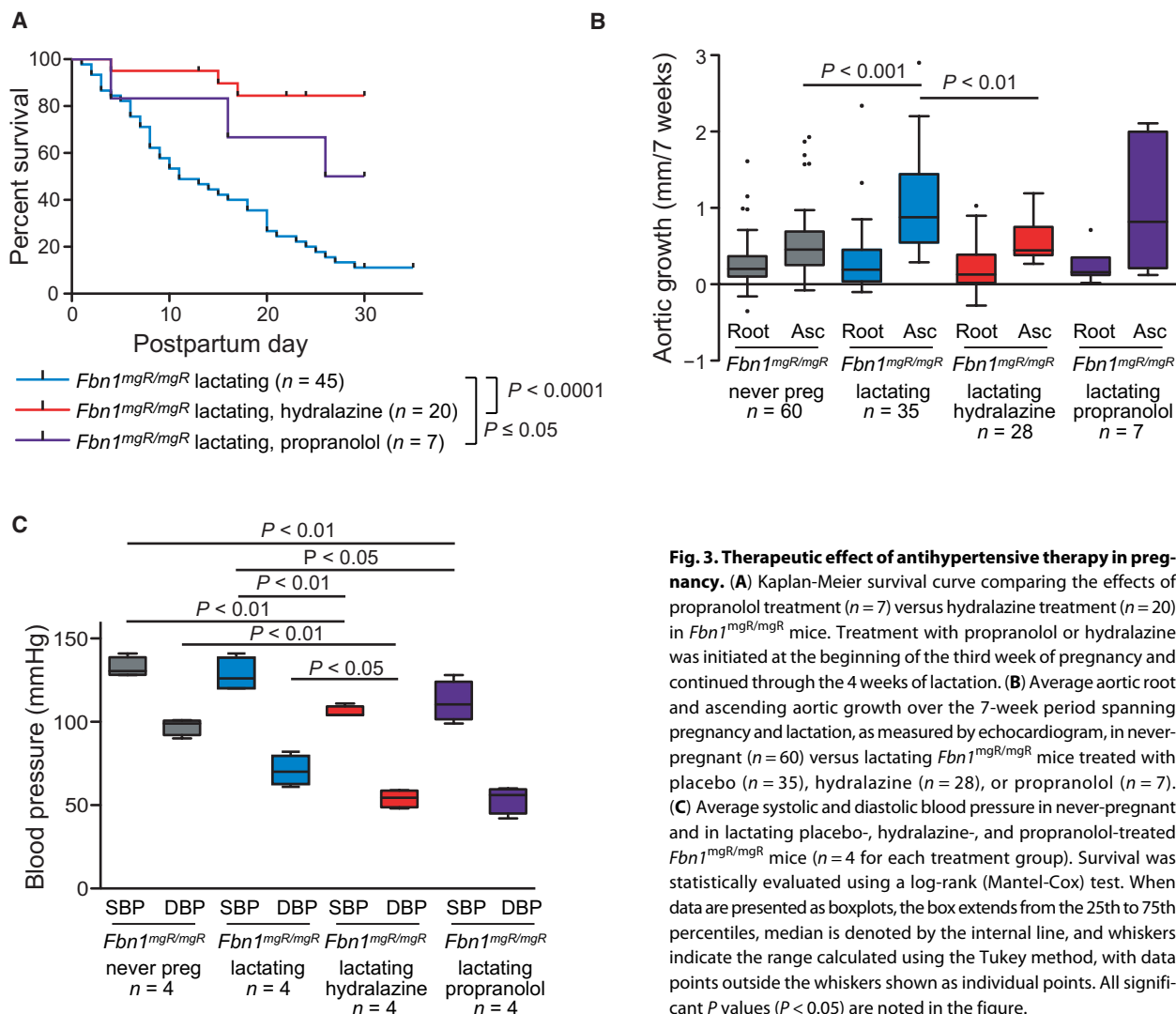
### Correlation of pregnancy outcomes with ERK1/2 activation

In keeping with our hypothesis that ERK1/2 signaling drives abnormal aortic growth, there was increased pERK1/2 in the aortic media of never-pregnant *Fbn1*<sup>mgR/mgR</sup> females as compared to never-pregnant WT females ( $P \leq 0.05$ ), with a significant further up-regulation induced by pregnancy and lactation ( $P \leq 0.01$ ). Furthermore, lactating *Fbn1*<sup>mgR/mgR</sup> mice had increased pERK1/2 as compared to lactating WT females ( $P < 0.001$ ) (Fig. 4A and table S6). Preventing lactation by removing pups or treatment with hydralazine resulted in a marked reduction of pERK1/2 in the ascending aorta, as compared to lactating *Fbn1*<sup>mgR/mgR</sup> mice ( $P \leq 0.05$  for both); a significant reduction was not observed upon treatment with OTA ( $P = 0.14$ ). Removal of pups or treatment with OTA or hydralazine resulted in ERK1/2 phosphorylation in the aortic wall of *Fbn1*<sup>mgR/mgR</sup> mice that was indistinguishable from that observed in lactating WT mice (Fig. 4B and table S7). Immunofluorescence (IF) staining for pERK1/2 revealed an increase in the aortic media in the aortic root and ascending aorta (Fig. 4C, most notably in the ascending aorta, in *Fbn1*<sup>mgR/mgR</sup> mice compared to WT animals. A further increase was noted in lactating *Fbn1*<sup>mgR/mgR</sup> mice, and this was reduced with OTA administration (Fig. 4C).





**Fig. 2. Therapeutic manipulations that alter the survival and aortic growth in pregnancy.** (A) Kaplan-Meier survival curve comparing *Fbn1<sup>mgR/mgR</sup>* never-pregnant (n = 60) and *Fbn1<sup>mgR/mgR</sup>* lactating (n = 45) mice to *Fbn1<sup>mgR/mgR</sup>* females with pups removed on the day of delivery, thereby preventing lactation and eliminating the lactation-induced prolonged elevation of oxytocin (n = 27). (B) Average aortic root and ascending aortic growth over the 7-week period spanning pregnancy (3 weeks) and lactation (4 weeks) in *Fbn1<sup>mgR/mgR</sup>* never-pregnant (n = 60), *Fbn1<sup>mgR/mgR</sup>* lactating (n = 35), and *Fbn1<sup>mgR/mgR</sup>* females with pups removed (n = 20). (C) Representative proximal ascending aortic wall sections stained with VVG for elastin, demonstrating elastic fiber breaks (white arrows), cellularity, and thickness of the adventitia (black asterisks) in the *Fbn1<sup>mgR/mgR</sup>* and WT mice. (D) Kaplan-Meier survival curve comparing *Fbn1<sup>mgR/mgR</sup>* lactating mice (n = 45) to *Fbn1<sup>mgR/mgR</sup>* females with OTA administered via a continuous subcutaneous infusion pump implanted at the beginning of the third week of gestation and continued through the 4 weeks of lactation for a total of 5 weeks of treatment (n = 19). (E) Average aortic root and ascending aortic growth over the 7-week period spanning pregnancy and lactation in never-pregnant (n = 60), lactating (n = 35), and OTA-treated (n = 15) *Fbn1<sup>mgR/mgR</sup>* mice. (F) Kaplan-Meier survival curve assessing the effects of OA administered to *Fbn1<sup>mgR/mgR</sup>* mice (n = 10) via a continuous subcutaneous infusion pump implanted at the beginning of the third week of gestation and continued through the 4 weeks of lactation. (G) Kaplan-Meier survival curve assessing the effects of OA administered to *Fbn1<sup>mgR/mgR</sup>* mice (n = 10) via a continuous subcutaneous infusion pump implanted at the beginning of the third week of gestation and continued through the 4 weeks of lactation for a total of 5 weeks of treatment. Survival was statistically evaluated using a log-rank (Mantel-Cox) test. When data are presented as boxplots, the box extends from the 25th to 75th percentiles, median is denoted by the internal line, and whiskers indicate the range calculated using the Tukey method, with data points outside the whiskers shown as individual points. All significant P values (P < 0.05) are noted in the figure.



**Fig. 3. Therapeutic effect of antihypertensive therapy in pregnancy.** (A) Kaplan-Meier survival curve comparing the effects of propranolol treatment (n = 7) versus hydralazine treatment (n = 20) in  $Fbn1^{mgR/mgR}$  mice. Treatment with propranolol or hydralazine was initiated at the beginning of the third week of pregnancy and continued through the 4 weeks of lactation. (B) Average aortic root and ascending aortic growth over the 7-week period spanning pregnancy and lactation, as measured by echocardiogram, in never-pregnant (n = 60) versus lactating  $Fbn1^{mgR/mgR}$  mice treated with placebo (n = 35), hydralazine (n = 28), or propranolol (n = 7). (C) Average systolic and diastolic blood pressure in never-pregnant and in lactating placebo-, hydralazine-, and propranolol-treated  $Fbn1^{mgR/mgR}$  mice (n = 4 for each treatment group). Survival was statistically evaluated using a log-rank (Mantel-Cox) test. When data are presented as boxplots, the box extends from the 25th to 75th percentiles, median is denoted by the internal line, and whiskers indicate the range calculated using the Tukey method, with data points outside the whiskers shown as individual points. All significant P values ( $P < 0.05$ ) are noted in the figure.

### Treatment with MEK inhibitor

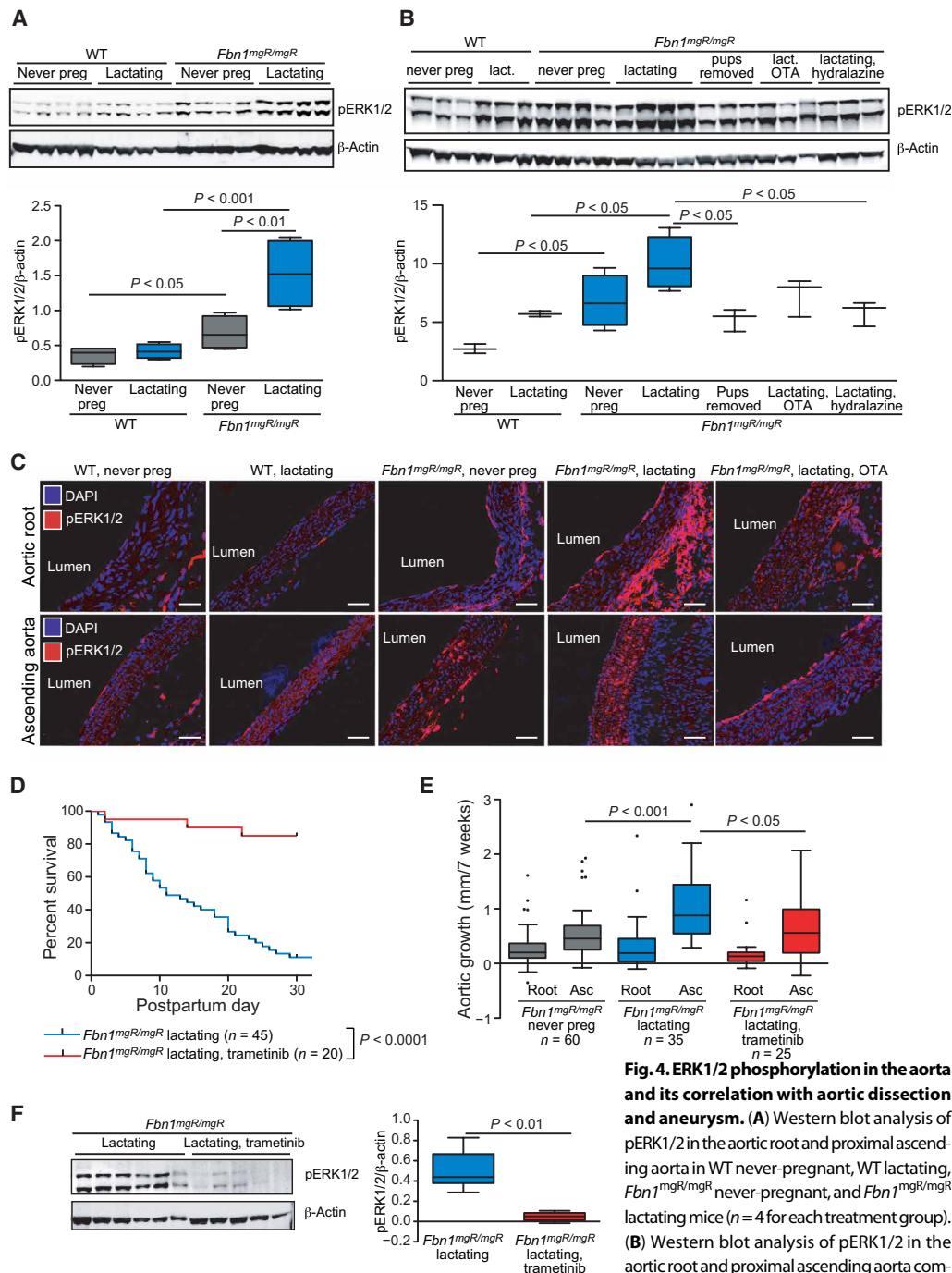
To further clarify the role of ERK activation in pregnancy-associated aortic growth and dissection, we treated pregnant  $Fbn1^{mgR/mgR}$  mice with trametinib, a U.S. Food and Drug Administration–approved oral MEK inhibitor (MEKi), beginning at the third trimester and continuing for 5 weeks. Trametinib significantly improved freedom from aortic dissection, with 85% of mice surviving ( $P \leq 0.0001$ ) (Fig. 4D). Trametinib significantly reduced ascending aortic growth ( $0.64 \pm 0.55$  mm/7 weeks,  $P \leq 0.05$ ) as compared to lactating  $Fbn1^{mgR/mgR}$  mice, with the result indistinguishable from never-pregnant  $Fbn1^{mgR/mgR}$  females (Fig. 4E and tables S1 and S2). There was no significant reduction in blood pressure upon treatment with trametinib (fig. S4 and table S8). The phosphorylation of ERK1/2 in the ascending aorta was significantly decreased in lactating  $Fbn1^{mgR/mgR}$  mice treated with MEKi ( $P \leq 0.001$ ) (Fig. 4F and table S9), demonstrating that oral trametinib was having the expected effect in this experimental system.

### DISCUSSION

Although a variety of hypotheses regarding the mechanism of pregnancy-associated aortic dissection have been proposed, alter-

ation of hemodynamic parameters is typically considered the dominant determinant of predisposition. Contributing factors could include the increased circulating volume and heart rate during pregnancy, the marked isometric strain that attends vaginal delivery, or the rapid fluid shifts and obligate diuresis in the immediate peripartum period. It is difficult to reconcile a strong predisposition for vascular catastrophe in the weeks to months after delivery with pathogenic models that singularly invoke the acute hemodynamic consequences of pregnancy. The observation of delayed risk of postpartum vascular dissection not only is restricted to MFS but also pertains to other heritable connective tissue disorders such as vascular Ehlers-Danlos syndrome or Loeys-Dietz syndrome and to more common nonsyndromic conditions without a defined genetic contribution such as pregnancy-associated spontaneous cervical or coronary artery dissection (26,27).

This study using a genetically defined and validated mouse model of MFS recapitulates a strong predisposition for postnatal aortic dissection and provides complementary and compelling evidence against a dominant hemodynamic mechanism. First, antihypertensive agents that achieved a comparable lowering of blood pressure during and after pregnancy were associated with variable outcomes,



**Fig. 4. ERK1/2 phosphorylation in the aorta and its correlation with aortic dissection and aneurysm.** (A) Western blot analysis of pERK1/2 in the aortic root and proximal ascending aorta in WT never-pregnant, WT lactating, *Fbn1<sup>mgR/mgR</sup>* never-pregnant, and *Fbn1<sup>mgR/mgR</sup>* lactating mice ( $n = 4$  for each treatment group). (B) Western blot analysis of pERK1/2 in the aortic root and proximal ascending aorta comparing

WT never-pregnant ( $n = 3$ ) and lactating mice ( $n = 3$ ), *Fbn1<sup>mgR/mgR</sup>* never-pregnant ( $n = 4$ ) and lactating ( $n = 4$ ) mice, *Fbn1<sup>mgR/mgR</sup>* mice with pups removed thereby preventing lactation ( $n = 3$ ), and *Fbn1<sup>mgR/mgR</sup>* mice treated with OTA ( $n = 3$ ) or hyalalazine ( $n = 3$ ). (C) pERK1/2 IF of the aortic root and ascending aorta in representative WT and *Fbn1<sup>mgR/mgR</sup>* never-pregnant, lactating, and lactating *Fbn1<sup>mgR/mgR</sup>* treated with OTA. Scale bars, 50  $\mu\text{m}$ . DAPI, 4',6'-diamidino-2-phenylindole. (D) Kaplan-Meier curve demonstrating the survival of *Fbn1<sup>mgR/mgR</sup>* lactating mice treated with trametinib ( $n = 20$ ), initiated at the start of the third week of pregnancy and continued through 4 weeks of lactation, in comparison to untreated *Fbn1<sup>mgR/mgR</sup>* mice ( $n = 45$ ). (E) Average aortic root and proximal ascending aortic growth over the 7 weeks spanning pregnancy and lactation in *Fbn1<sup>mgR/mgR</sup>* never-pregnant ( $n = 60$ ) or lactating *Fbn1<sup>mgR/mgR</sup>* mice treated with placebo ( $n = 35$ ) or trametinib ( $n = 25$ ). (F) Western blot analysis of pERK1/2 in the aortic root and proximal ascending aorta comparing lactating *Fbn1<sup>mgR/mgR</sup>* mice treated with placebo ( $n = 6$ ) or trametinib ( $n = 5$ ). Survival was statistically evaluated using a log-rank (Mantel-Cox) test. When data are presented as boxplots, the box extends from the 25th to 75th percentiles, median is denoted by the internal line, and whiskers indicate the range calculated using the Tukey method, with data points outside the whiskers shown as individual points. All significant  $P$  values ( $P < 0.05$ ) are noted in the figure.

ranging from modest (propranolol) to substantial (hydralazine) protection. Second, the incidence of dissection was potentially reduced by either removal of the pups or administration of a selective OTA, strongly implicating lactation as a major contributory risk factor. The suggestion of differential effect of oxytocin agonism in nonpregnant versus pregnant *Fbn1<sup>mgR/mgR</sup>* mice, along with the up-regulation of OR expression in the aortic wall in both WT and *Fbn1<sup>mgR/mgR</sup>* mice, supports the hypothesis that pregnancy induces a more vulnerable aortic environment and suggests that oxytocin is a major contributor but is not sufficient for acceleration of aneurysm or dissection and that other factors associated with pregnancy, including up-regulation of the OR, are necessary. Although in vitro studies have documented that oxytocin has vasoactive properties, with dilatation or constriction at low and high concentrations, respectively (28), this study failed to demonstrate any blood pressure differences imposed by lactation or oxytocin antagonism. Third, the marked protection afforded by inhibition of MEK-mediated ERK activation was not associated with any apparent hemodynamic consequence.

Oxytocin has been reported to be expressed in both endothelial cells and cultured vascular smooth muscle cells (VSMCs), where it can support calcium-mediated responses including ERK activation. A parsimonious explanation for the observed increase in pERK in the aortic media in lactating *Fbn1<sup>mgR/mgR</sup>* mice would invoke oxytocin signaling in VSMCs, but more complex paracrine interactions that involve the endothelium cannot be excluded (29–31).

The deleterious gene-by-environment interaction imposed by pregnancy and lactation in MFS mice shows apparent correlates with the acceleration of aortic aneurysm growth and tear observed in MFS mice treated with calcium channel blockers (CCBs) (25). CCBs induced maximal dilatation in the



more distal ascending aorta in association with excessive ERK phosphorylation that was ultimately linked to a phospholipase C (PLC)–inositol triphosphate (IP<sub>3</sub>)/diacylglycerol (DAG)–protein kinase C (PKC) axis of activation. Normalization of both clinical and biochemical parameters was observed upon treatment with the IP<sub>3</sub> or PKC antagonist hydralazine or enzastaurin, respectively (25). In this light, it is notable that both the OR and AT1 (the target of losartan) are G protein–coupled receptors that signal through the Gαq-PLC-IP<sub>3</sub>/DAG-PKC-ERK axis. Desensitization of the OR, and hence inhibition of uterine contraction, is dependent on β-arrestins that paradoxically enhance ERK phosphorylation (in response to either OR or AT1 stimulation) but suppress nuclear localization and transcriptional responses (32,33). Previous work has suggested that enhanced TGFβ activity can be either an upstream effector or a downstream consequence of ERK activation in the pathogenesis of MFS in a context-dependent manner (13, 34). Together, this study and previous work suggest that the timing and severity of aneurysm in MFS correlates with the extent of ERK signaling in the aortic wall. It appears that the total phosphorylation of ERK in pregnancy integrates parallel inputs, because prevention of lactation or oxytocin antagonism lowers pERK to the level seen in never-pregnant MFS mice, but not the lower level observed in WT animals.

We note that our study has several limitations. The period of observation (3 weeks of gestation and 4 weeks postpartum) provided limited time for aortic growth, which is largely restricted to the more distal ascending aorta in the *Fbn1*<sup>mgR/mgR</sup> mouse model. This precluded our ability to assess for differences in growth of the aortic root, an aortic segment more typically involved in people with MFS. Western blot analyses were performed on protein lysates derived from tissue extending from just above the aortic valve to the origin of the brachiocephalic artery. This practice did not allow us to distinguish between the protein expression contribution of the aortic root and the more distal ascending aorta. In addition, although careful attention was given to controlling for age and mouse background, there was variability in the absolute aortic dimensions within a given state (genotype and treatment arm) that may have resulted in underestimation of treatment effects. This was offset by using death due to aortic dissection as a discrete outcome parameter.

This study has multiple potential therapeutic implications that will require further study and validation in people. The most obvious and immediately applicable intervention would be the avoidance of lactation and/or the use of OTAs late in gestation and in the postpartum period. OTAs are currently in clinical use in Europe for the suppression of preterm labor. It is notable that OTA-treated MFS mice retained the ability to deliver spontaneously and to support their pups through lactation, suggesting that protection from pregnancy-associated aortic dissection can be achieved with incomplete OR antagonism. In contrast, *Otxr*-null mice support spontaneous labor and delivery, but show a critical defect in milk letdown (24). The use of Pitocin (a synthetic form of oxytocin) to induce labor or as a routine practice after delivery to suppress uterine bleeding will need to be critically evaluated in at-risk populations. Unlike ARBs, hydralazine can be used during pregnancy and could be considered as an alternative or adjunct to β-blocker therapy. Studies have documented that oxytocin concentrations in women are measurably increased at 12 weeks of gestation, rise throughout pregnancy, and peak at delivery. Concentrations return to baseline by 8 weeks postpartum in the absence of lactation, but are sustained at delivery-associated values throughout lactation (16). If epidemiologic studies confirm a role

for oxytocin in pregnancy-associated vascular events in women with MFS and related disorders, there is the potential for rapid translation of therapeutic strategies that include modification of delivery and breastfeeding practices and pharmacologic interventions that include hydralazine and OTAs.

## MATERIALS AND METHODS

### Study design

The objective of the study was to investigate factors contributing to the increased risk of pregnancy-associated aortic aneurysm progression and dissection in MFS. We hypothesized that oxytocin played a role by increasing activation of the ERK signaling pathway and that inhibiting oxytocin or ERK directly could decrease the risk of dissection. We used an established mouse model for MFS (*Fbn1*<sup>mgR/mgR</sup>) and observed an increased risk of dissection in the 4 weeks postpartum, the period of lactation. All drug interventions were initiated at the beginning of the third trimester to allow for completion of organ development, and continued postnatally for the 4-week period of lactation. A minimum of seven mice were included in each treatment arm. All experiments used mice of the same age and background that were randomly assigned to treatment arms. Although each treatment trial included contemporaneous control animals, the performance of untreated mice remained constant for the full duration of this study, allowing pooling of controls for robust comparisons.

### Mice

All mice were cared for under strict adherence to the guidelines of the Animal Care and Use Committee of the Johns Hopkins University School of Medicine. Heterozygous mutant mice (*Fbn1*<sup>mgR/+</sup>) were purchased from The Jackson Laboratory in a pure C57BL/6 background, and homozygous mutant mice (*Fbn1*<sup>mgR/mgR</sup>) were generated. Heterozygous mutant mice (*Fbn1*<sup>mgR/+</sup>) were used to surrogate nurse the pups of the homozygous females who were sacrificed or died during lactation. Control mice were pure C57BL/6 WT mice. Genotyping was performed at 3 weeks of life by polymerase chain reaction (PCR). DNA was isolated from tags using standard methods. Quantitative PCR (qPCR) was performed with four amplicons (Alb1-FOR:CTGCAATCCTGAACCGTGT, Alb1-REV:TTCCACCAGG-GATCCACTAC; qNEO1-FOR:TGAATGAACTGCAGGACGAG, qNEO1-REV:AGTGACAACGTCGAGCACAG; qNEO2-FOR:TCTCCTGTATCTCACCTTGC, qNEO2-REV:GTAGCCGGAT-CAAGCGTATG; and qNEO3-FOR:TCTGGATTTCATCGACTGT-GG, qNEO3-REV:TTCAGCAATATCACGGGTAGC) detecting the introduced neomycin cassette.

Seven- to 9-week-old female *Fbn1*<sup>mgR/+</sup> and *Fbn1*<sup>mgR/mgR</sup> mice underwent timed mating to male *Fbn1*<sup>mgR/mgR</sup> mice. Survival was assessed for the 4 weeks after delivery, during which females remained with their pups for nursing. Mice were sacrificed with an inhalation overdose of 2-bromo-2-chloro-1,1,1-trifluoroethane (Sigma). Mice underwent immediate laparotomy and descending abdominal aortic transection, and phosphate-buffered saline (PBS) (pH 7.4) was infused through the right and left ventricles to flush out the blood. Mice that were used for Western blot analysis had their aortic root and ascending aortas (aortic root to right brachiocephalic trunk) immediately dissected out, flash-frozen in liquid nitrogen, and stored at –80°C until further processing. Mice that were analyzed for histology had their abdominal aorta transected before the bifurcation, and PBS, followed by latex (Ward's Science), was infused into the

left ventricle. Latex-infused aortas were then transected at the level of the aortic valve, and 3-mm transverse sections were mounted in 4% bacto-agar (BD Biosciences) and submitted for paraffin fixation. Concentric aortic rings (5  $\mu$ m thick) were mounted on glass slides, stained with VVG, and imaged at  $\times 10$  and  $\times 40$  magnification using a Nikon Eclipse E400 microscope. All mice found dead were assessed for cause of death by necropsy, noting in particular hemothorax and hemopericardium.

### Delivery of medication

Mice were initiated on medication at the beginning of the third week of gestation and continued for 5 weeks: 1 week of pregnancy and 4 weeks of lactation. Both the selective OTA [desGly-NH<sub>2</sub>-d(CH<sub>2</sub>)<sub>5</sub>[D-Tyr<sup>2</sup>,Thr<sup>4</sup>]OVT] and the selective OA [(Thr<sup>4</sup>,Gly<sup>7</sup>)OT] were synthesized in the Manning laboratory (23), dissolved in PBS to reach a final concentration of 0.2 mg/ml, and delivered via continuous infusion for 5 weeks with a mini Alzet pump implanted subcutaneously between the scapulae at 3 weeks of gestation for a dose of 1  $\mu$ g/kg per hour. Placebo-treated animals underwent pump implantation and received continuous infusion of PBS. Propranolol (QualiTest) was dissolved in the drinking water and filtered to reach a final concentration of 0.5 g/liter, giving an estimated daily dose of 40 mg/kg per day. Hydralazine (AmerisourceBergen) was dissolved in the drinking water and filtered to reach a final concentration of 0.16 g/liter, giving an estimated daily dose of 16 mg/kg per day. Trametinib (GlaxoSmithKline) was dissolved in PBS with 5% dimethyl sulfoxide and administered by oral gavage once a day, giving an estimated daily dose of 1 mg/kg per day. Placebo-treated animals received regular drinking water.

### Echocardiography

Nair hair removal cream was used on all mice the day before echocardiograms. All echocardiograms were performed on awake, unsedated mice using a VisualSonics Vevo 660 V1.3.6 imaging system and a 30-MHz transducer. Mice were imaged at baseline and weekly thereafter until death or the time of sacrifice. The aorta was imaged using a parasternal long-axis view. Three separate measurements of the maximal internal dimension at the aortic root and proximal ascending aorta were made from distinct captured images and averaged. All imaging and measurements were performed by a cardiologist (J.P.H.) who was blinded to genotype and treatment arm.

### Blood pressure analysis

Blood pressures were analyzed by taking 20 tail-cuff blood pressures per day over 2 to 3 days in each mouse to habituate the mice to the tail-cuff pressure system, and then 10 to 20 blood pressure measurements were obtained on the last day and averaged. At least four mice for each treatment group were analyzed.

### Antibodies and Western blot analysis

Mouse aortic root and ascending aortas (aortic root excluding the aortic valve to origin of right brachiocephalic trunk) were harvested, snap-frozen in liquid nitrogen, and stored at  $-80^{\circ}\text{C}$  until processed. Protein was extracted using an automatic bead homogenizer in conjunction with reagents from the Protein Extraction Kit (Full Moon BioSystems). All protein lysis buffers contained protease and phosphatase inhibitors (Millipore). Western blotting was performed using LI-COR buffer and species-appropriate secondary antibodies conjugated to IRDye 700 (IRDye 680LT donkey anti-mouse, catalog

no. 926-68022, LI-COR Biosciences) or IRDye 800 (IRDye 800CW donkey anti-rabbit, catalog no. 926-32213, LI-COR Biosciences), according to the manufacturer's guidelines, and analyzed using LI-COR Odyssey. The following primary antibodies were used: anti-actin (Sigma-Aldrich, clone AC-74) and anti-pERK1/2 (Cell Signaling Technology, 4370).

pERK amounts were normalized to  $\beta$ -actin as opposed to total ERK for a variety of practical reasons. First, the functional consequences for the cell depend on the amount of pERK but not the pERK/total ERK ratio. Second, the amount of total ERK has never been shown to be limiting in *in vitro* contexts. Earlier work has shown the abundant recruitment of cells into the aortic wall of aneurysm models (32,33) that have high expression of total ERK but not pERK, markedly diluting the pERK signal when averaging techniques such as immunoblots are performed using protein lysates from the entire aortic wall and total ERK for normalization. This practice results in extreme intrastate variability that is minimized when a housekeeping gene is used as loading control.

### IF on frozen aortic tissue sections

Mice were euthanized by halothane inhalation, and the left common iliac artery was transected to allow for drainage. A total of 20 ml of PBS (pH 7.4) and then about 20 ml of PBS containing 4% paraformaldehyde were flushed through the left ventricle. The heart and thoracic aorta were then removed en bloc and fixed in fresh 4% paraformaldehyde in PBS at  $4^{\circ}\text{C}$  overnight. Tissue blocks were immersed in antigen retrieval solution (10 mM sodium citrate buffer, pH 6.0) at  $4^{\circ}\text{C}$  overnight and then in boiling antigen retrieval solution for 3 min. Immediately after that, tissue was placed in cold 30% sucrose in PBS and incubated at  $4^{\circ}\text{C}$  overnight, embedded in Tissue-Tek O.C.T. Compound, and frozen using ethanol mixed with dry ice and stored at  $-80^{\circ}\text{C}$ . Frozen 10- $\mu$ m long-axis view sections were obtained with a cryostat, mounted on glass slides, and dried at room temperature for at least 1 day before staining. Sections were permeabilized in staining buffer (PBS containing 0.1% Triton X-100) for 15 min and then incubated with Fc Receptor Block (Innovex Biosciences, catalog no. NB309) for 30 min at room temperature, washed in staining buffer, and then incubated again in Background Buster blocking solution (Innovex Biosciences, catalog no. NB306) for 30 min. Anti-pERK1/2 (Cell Signaling Technology, 4370) was diluted at 1:100 in staining buffer and incubated overnight at  $4^{\circ}\text{C}$ . Three consecutive washes with staining buffer were performed before incubation with goat anti-rabbit secondary antibody conjugated to Alexa Fluor 555 [goat anti-rabbit IgG (H+L) secondary antibody, Alexa Fluor 555 conjugate, #A-21428 from Thermo Fisher Scientific] at 1:200 for 1 hour. Images were acquired on Zeiss AxioExaminer with 780NLO-Meta multiphoton confocal microscope at a  $\times 25$  magnification.

### OR expression

Aortas were dissected as described above, flushed in PBS, and directly stored in TRIzol (Invitrogen). RNA was extracted according to the manufacturer's instruction and purified with RNeasy mini columns (Qiagen). An on-column deoxyribonuclease digest (Qiagen) was performed before the clean-up step to eliminate residual genomic DNA. Complementary DNA (cDNA) was generated using TaqMan High Capacity cDNA Reverse Transcription reagents, and qPCR was performed in triplicate with TaqMan Universal PCR Master Mix (Applied Biosystems). The following prevalidated TaqMan probes



were used: Mm01182684\_m1 (*Oxtr*) and Mm99999915\_g1 (*Gapdh*) (34). Relative quantification for each transcript was obtained by normalizing against *Gapdh* transcript abundance according to the formula  $2^{-(Ct)}/2^{-(Ct \text{ } Gapdh)}$ .

### Statistical analysis

Quantitative data are presented as boxplots with the box extending from the 25th to 75th percentile, the internal line represents the median, and the whiskers (indicating range) are calculated using the Tukey method (the upper whisker represents 75th percentile plus 1.5 times the interquartile distance, and the lower whisker represents 25th percentile minus 1.5 times the interquartile difference); data points that fall above or below the whiskers (outliers) are shown as individual points but included in statistical analyses. One-way analysis of variance (ANOVA) and two-way ANOVA were performed to evaluate significance of comparisons between groups using Tukey's multiple comparison test, with a *P* value of <0.05 considered statistically significant. Kaplan-Meier survival curves were compared using a log-rank (Mantel-Cox) test.

### SUPPLEMENTARY MATERIALS

stm.sciencemag.org/cgi/content/full/11/490/eaat4822/DC1

Fig. S1. Subcutaneous pump placement effect on survival.

Fig. S2. Blood pressure effect in *Fbn1*<sup>mgR/mgR</sup> mice.

Fig. S3. OR mRNA expression in the proximal ascending aorta.

Fig. S4. Blood pressure effect in *Fbn1*<sup>mgR/mgR</sup> mice.

Table S1. Average aortic root and ascending aortic growth over the 7-week period spanning pregnancy and lactation in untreated mice.

Table S2. Average aortic root and ascending aortic growth over the 7-week period spanning pregnancy and lactation in treated mice.

Table S3. Effect of OTA treatment on the average systolic and diastolic blood pressure.

Table S4. Average OR expression in the aorta.

Table S5. Effect of hydralazine or propranolol on the average systolic and diastolic blood pressure.

Table S6. Average ERK1/2 phosphorylation in untreated mice.

Table S7. Average ERK1/2 phosphorylation in treated mice.

Table S8. Effect of trametinib on average systolic and diastolic blood pressure.

Table S9. Average ERK1/2 phosphorylation in trametinib-treated mice.

### REFERENCES AND NOTES

- D. E. Cameron, D. E. Alejo, N. D. Patel, L. U. Nwakanma, E. S. Weiss, L. A. Vricella, H. C. Dietz, P. J. Spevak, J. A. Williams, B. T. Bethea, T. P. Fitton, V. L. Gott, Aortic root replacement in 372 Marfan patients: Evolution of operative repair over 30 years. *Ann. Thorac. Surg.* **87**, 1344–1349 (2009).
- S. Goland, U. Elkayam, Cardiovascular problems in pregnant women with Marfan syndrome. *Circulation* **119**, 619–623 (2009).
- U. Elkayam, E. Ostrzega, A. Shotan, A. Mehra, Cardiovascular problems in pregnant women with the Marfan syndrome. *Ann. Intern. Med.* **123**, 117–122 (1995).
- L. J. Meijboom, F. E. Vos, J. Timmermans, G. H. Boers, A. H. Zwinderman, B. J. M. Mulder, Pregnancy and aortic root growth in the Marfan syndrome: A prospective study. *Eur. Heart J.* **26**, 914–920 (2005).
- S. Lalchandani, M. Wingfield, Pregnancy in women with Marfan's syndrome. *Eur. J. Obstet. Gynecol. Reprod. Biol.* **110**, 125–130 (2003).
- L. Pacini, F. Digne, A. Boumendil, C. Muti, D. Detaint, C. Boileau, G. Jondeau, Maternal complication of pregnancy in Marfan syndrome. *Int. J. Cardiol.* **136**, 156–161 (2009).
- L. Houston, M. Tuuli, G. Macones, Marfan syndrome and aortic dissection in pregnancy. *Obstet. Gynecol.* **117**, 956–960 (2011).
- M. J. Roman, N. L. Pugh, T. P. Hendershot, R. B. Devereux, H. Dietz, K. Holmes, K. A. Eagle, S. A. LeMaire, D. M. Milewicz, S. A. Morris, R. E. Pyeritz, W. J. Ravekes, R. V. Shohet, M. Silberbach; GenTAC Investigators, Aortic complications associated with pregnancy in Marfan syndrome: The NHLBI national registry of genetically triggered thoracic aortic aneurysms and cardiovascular conditions (GenTAC). *J. Am. Heart Assoc.* **5**, e004052 (2016).
- S. Sayama, N. Takeda, T. Iriyama, R. Inuzuka, S. Maemura, D. Fujita, H. Yamauchi, K. Nawata, M. Bougaki, H. Hyodo, R. Shitara, T. Nakayama, A. Komatsu, T. Nagamatsu, Y. Osuga, T. Fujii, Peripartum type B dissection in patients with Marfan syndrome who underwent aortic root replacement: A case series study. *BJOG* **125**, 487–493 (2018).
- E. R. Neptune, P. A. Frischmeyer, D. E. Arking, L. Myers, T. E. Bunton, B. Gayraud, F. Ramirez, L. Y. Sakai, H. C. Dietz, Dysregulation of TGF- $\beta$  activation contributes to pathogenesis in Marfan syndrome. *Nat. Genet.* **33**, 407–411 (2003).
- J. P. Habashi, D. P. Judge, T. M. Holm, R. D. Cohn, B. L. Loeys, T. K. Cooper, L. Myers, E. C. Klein, G. Liu, C. Calvi, M. Podowski, E. R. Neptune, M. K. Halushka, D. Bedja, K. Gabrielson, D. B. Rifkin, L. Carta, F. Ramirez, D. L. Huso, H. C. Dietz, Losartan, an AT1 antagonist, prevents aortic aneurysm in a mouse model of Marfan syndrome. *Science* **312**, 117–121 (2006).
- J. P. Habashi, J. J. Doyle, T. M. Holm, H. Aziz, F. Schoenhoff, D. Bedja, Y. Chen, A. N. Modiri, D. P. Judge, H. C. Dietz, Angiotensin II type 2 receptor signaling attenuates aortic aneurysm in mice through ERK antagonism. *Science* **332**, 361–365 (2011).
- J. R. Cook, N. P. Clayton, L. Carta, J. Galatioto, E. Chiu, S. Smaldone, C. A. Nelson, S. H. Cheng, B. M. Wentworth, F. Ramirez, Dimorphic effects of transforming growth factor- $\beta$  signaling during aortic aneurysm progression in mice suggest a combinatorial therapy for Marfan syndrome. *Arterioscler. Thromb. Vasc. Biol.* **35**, 911–917 (2015).
- T. M. Holm, J. P. Habashi, J. J. Doyle, D. Bedja, Y. Chen, C. van Erp, M. E. Lindsay, D. Kim, F. Schoenhoff, R. D. Cohn, B. L. Loeys, C. J. Thomas, S. Patnaik, J. J. Marugan, D. P. Judge, H. C. Dietz, Noncanonical TGF $\beta$  signaling contributes to aortic aneurysm progression in Marfan syndrome mice. *Science* **332**, 358–361 (2011).
- J. R. Cook, L. Carta, J. Galatioto, F. Ramirez, Cardiovascular manifestations in Marfan syndrome and related diseases; multiple genes causing similar phenotypes. *Clin. Genet.* **87**, 11–20 (2015).
- S. Stock, K. Bremme, K. Uvnäs-Moberg, Plasma levels of oxytocin during the menstrual cycle, pregnancy and following treatment with HMG. *Hum. Reprod.* **6**, 1056–1062 (1991).
- K. Uvnäs-Moberg, A.-M. Widström, S. Werner, A.-S. Matthiesen, J. Winberg, Oxytocin and prolactin levels in breast-feeding women. *Acta Obstet. Gynecol. Scand.* **69**, 301–306 (1990).
- M. Jankowski, D. Wang, F. Hajjar, S. Mukaddam-Daher, S. M. McCann, J. Gutkowska, Oxytocin and its receptors are synthesized in the rat vasculature. *Proc. Natl. Acad. Sci. U.S.A.* **97**, 6207–6211 (2000).
- H. A. Otun, M. W. MacDougall, J. Bailey, G. N. Europe-Finner, S. C. Robson, Spatial and temporal expression of the myometrial mitogen-activated protein kinase 38 and ERK1/2 in the human uterus during pregnancy and labor. *J. Soc. Gynecol. Investig.* **12**, 185–190 (2005).
- A. Nohara, M. Ohmichi, K. Koike, N. Masumoto, M. Kobayashi, M. Akahane, H. Ikegami, K. Hirota, A. Miyake, Y. Murata, The role of mitogen-activated protein kinase in oxytocin-induced contraction of uterine smooth muscle in pregnant rat. *Biochem. Biophys. Res. Commun.* **229**, 938–944 (1996).
- Y. Li, H.-D. Je, S. Malek, K. G. Morgan, Role of ERK1/2 in uterine contractility and preterm labor in rats. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **287**, R328–R335 (2004).
- L. Pereira, S. Y. Lee, B. Gayraud, K. Andrikopoulos, S. D. Shapiro, T. Bunton, N. J. Biery, H. C. Dietz, L. Y. Sakai, F. Ramirez, Pathogenetic sequence for aneurysm revealed in mice underexpressing fibrillin-1. *Proc. Natl. Acad. Sci. U.S.A.* **96**, 3819–3823 (1999).
- M. Manning, A. Misicka, A. Olma, K. Bankowski, S. Stoev, B. Chini, T. Durroux, B. Mouillac, M. Corbani, G. Guillon, Oxytocin and vasopressin agonists and antagonists as research tools and potential therapeutics. *J. Neuroendocrinol.* **24**, 609–628 (2012).
- K. Nishimori, L. J. Young, Q. Guo, Z. Wang, T. R. Insel, M. M. Matzuk, Oxytocin is required for nursing but is not essential for parturition or reproductive behavior. *Proc. Natl. Acad. Sci. U.S.A.* **93**, 11699–11704 (1996).
- J. J. Doyle, A. J. Doyle, N. K. Wilson, J. P. Habashi, D. Bedja, R. E. Whitworth, M. E. Lindsay, F. Schoenhoff, L. Myers, N. Huso, S. Bachir, O. Squires, B. Rusholme, H. Ehsan, D. Huso, C. J. Thomas, M. J. Caulfield, J. E. Van Eyk, D. P. Judge, H. C. Dietz; GenTAC registry consortium, MIBAVA leducq consortium, A deleterious gene-by-environment interaction imposed by calcium channel blockers in Marfan syndrome. *eLife* **4**, e08648 (2015).
- D. A. Hovsepian, N. Sriram, H. Kamel, M. E. Fink, B. B. Navi, Acute cerebrovascular disease occurring after hospital discharge for labor and delivery. *Stroke* **45**, 1947–1950 (2014).
- M. S. Tweet, S. N. Hayes, E. Codi, R. Gulati, C. H. Rose, P. J. M. Best, Spontaneous coronary artery dissection associated with pregnancy. *J. Am. Coll. Cardiol.* **70**, 426–435 (2017).
- S. Barrigón, J. Tamargo, The effect of oxytocin on contractile responses and <sup>45</sup>Ca movements in rat isolated aortic strips. *Br. J. Pharmacol.* **87**, 763–770 (1986).
- C. A. Grotegut, L. Feng, L. Mao, R. P. Heine, A. P. Murtha, H. A. Rockman,  $\beta$ -Arrestin mediates oxytocin receptor signaling, which regulates uterine contractility and cellular migration. *Am. J. Physiol. Endocrinol. Metab.* **300**, E468–E477 (2011).
- A. Tohgo, K. L. Pierce, E. W. Choy, R. J. Lefkowitz, L. M. Luttrell,  $\beta$ -Arrestin scaffolding of the ERK cascade enhances cytosolic ERK activity but inhibits ERK-mediated transcription following angiotensin AT1a receptor stimulation. *J. Biol. Chem.* **277**, 9429–9436 (2002).
- R. Rouf, E. G. MacFarlane, E. Takimoto, R. Chaudhary, V. Nagpal, P. P. Rainer, J. G. Bindman, E. E. Gerber, D. Bedja, C. Schiefer, K. L. Miller, G. Zhu, L. Myers, N. Amat-Alarcon, D. I. Lee, N. Koitabashi, D. P. Judge, D. A. Kass, H. C. Dietz, Nonmyocyte ERK1/2 signaling contributes to load-induced cardiomyopathy in Marfan mice. *JCI Insight* **15**, 91588 (2017).

32. S. Anidjar, P. B. Dobrin, M. Eichorst, G. P. Graham, G. Chejfec, Correlation of inflammatory infiltrate with the enlargement of experimental aortic aneurysms. *J. Vasc. Surg.* **16**, 139–147 (1992).
33. A. Daugherty, D. L. Rateri, I. F. Charo, A. Phillip Owens III, D. A. Howatt, L. A. Cassis, Angiotensin II infusion promotes ascending aortic aneurysms: Attenuation by CCR2 deficiency in apoE-mice. *Clin. Sci.* **118**, 681–689 (2010).
34. A. E. Clipperton-Allen, A. W. Lee, A. Reyes, N. Devidze, A. Phan, D. W. Pfaff, E. Choleris, Oxytocin, vasopressin and estrogen receptor gene expression in relation to social recognition in female mice. *Physiol. Behav.* **105**, 915–924 (2012).

**Acknowledgments:** We thank R. Makineni, R. Tyner, F. Paulsen, and the University of Toledo College of Medicine for their generous research support (to M.M.). **Funding:** Supported by grants from the NIH (AR41135 to H.C.D.), the Howard Hughes Medical Institute (to H.C.D.), the Marfan Foundation (to J.P.H.), the Smilow Center for Marfan Syndrome Research (to H.C.D.), and the University of Toledo College of Medicine (to M.M.). E.G.M. is supported by the Loews-Dietz Syndrome Foundation and NIH grant R00HL121287, and C.B. is supported by NIH grant GM007309. **Author contributions:** J.P.H. and H.C.D. were responsible for all aspects of study design, data interpretation, and preparation of the manuscript. E.G.M. performed immunoblot and IF analyses. R.B., N.H., Y.C., T.J.C., and G.R. were responsible for mouse

husbandry and medication dosing. C.B. monitored OR expression. D.B. and J.P.H. performed echocardiograms, and J.P.H. interpreted the echocardiograms. D.H. performed pump implantation. M.M. provided OTA and antagonist and guidance regarding their use in animal studies. **Competing interests:** H.C.D. has equity interest in Blade Therapeutics that develops drugs to treat fibrosis and consults for GlaxoSmithKline. H.C.D. and J.P.H. hold a patent entitled “Methods and compositions for the treatment of Marfan syndrome and associated disorders” (US-2010-0034806). All other authors declare that they have no competing interests. **Data and materials availability:** All data associated with this study are present in the paper or Supplementary Materials.

Submitted 2 March 2018

Resubmitted 14 September 2018

Accepted 25 March 2019

Published 1 May 2019

10.1126/scitranslmed.aat4822

**Citation:** J. P. Habashi, E. G. MacFarlane, R. Bagirzadeh, C. Bowen, N. Huso, Y. Chen, D. Bedja, T. J. Creamer, G. Rykiel, M. Manning, D. Huso, H. C. Dietz, Oxytocin antagonism prevents pregnancy-associated aortic dissection in a mouse model of Marfan syndrome. *Sci. Transl. Med.* **11**, eaat4822 (2019).

## Oxytocin antagonism prevents pregnancy-associated aortic dissection in a mouse model of Marfan syndrome

Jennifer Pardo Habashi, Elena Gallo MacFarlane, Rustam Bagirzadeh, Caitlin Bowen, Nicholas Huso, Yichun Chen, Djahida Bedja, Tyler J. Creamer, Graham Rykiel, Maurice Manning, David Huso and Harry C. Dietz

*Sci Transl Med* 11, eaat4822.  
DOI: 10.1126/scitranslmed.aat4822

### Dissecting a risk of Marfan syndrome

Marfan syndrome is an autosomal dominant connective tissue disorder associated with manifestations in multiple organ systems. The aorta is a key site affected by this syndrome, and female patients are at risk for aortic dissection associated with pregnancy and childbirth. Traditionally, this risk has been ascribed to labor-induced stress, but Habashi *et al.* noted that aortic dissection often occurs relatively late postpartum, prompting them to examine possible additional explanations. Using mouse models of Marfan syndrome, the researchers discovered that oxytocin, the hormone involved in milk letdown, plays a key role in aortic dissection, and identified several potential approaches for preventing this complication.

ARTICLE TOOLS	<a href="http://stm.sciencemag.org/content/11/490/eaat4822">http://stm.sciencemag.org/content/11/490/eaat4822</a>
SUPPLEMENTARY MATERIALS	<a href="http://stm.sciencemag.org/content/suppl/2019/04/29/11.490.eaat4822.DC1">http://stm.sciencemag.org/content/suppl/2019/04/29/11.490.eaat4822.DC1</a>
RELATED CONTENT	<a href="http://stm.sciencemag.org/content/scitransmed/5/183/183ra58.full">http://stm.sciencemag.org/content/scitransmed/5/183/183ra58.full</a> <a href="http://stm.sciencemag.org/content/scitransmed/12/527/eaay1059.full">http://stm.sciencemag.org/content/scitransmed/12/527/eaay1059.full</a> <a href="http://stm.sciencemag.org/content/scitransmed/12/550/eaaz0131.full">http://stm.sciencemag.org/content/scitransmed/12/550/eaaz0131.full</a>
REFERENCES	This article cites 34 articles, 12 of which you can access for free <a href="http://stm.sciencemag.org/content/11/490/eaat4822#BIBL">http://stm.sciencemag.org/content/11/490/eaat4822#BIBL</a>
PERMISSIONS	<a href="http://www.sciencemag.org/help/reprints-and-permissions">http://www.sciencemag.org/help/reprints-and-permissions</a>

Use of this article is subject to the [Terms of Service](#)

*Science Translational Medicine* (ISSN 1946-6242) is published by the American Association for the Advancement of Science, 1200 New York Avenue NW, Washington, DC 20005. The title *Science Translational Medicine* is a registered trademark of AAAS.

Copyright © 2019 The Authors, some rights reserved; exclusive licensee American Association for the Advancement of Science. No claim to original U.S. Government Works